

1       **Global gene expression analysis in cord blood reveals gender-specific**  
2               **differences in response to carcinogenic exposure *in utero***

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60 **Abstract**

61 Background: It has been suggested that such foetal exposure might lead  
62 to predisposition to develop cancer during childhood or in later life possibly  
63 through modulation of the foetal transcriptome. Since gender effects in the  
64 incidence of childhood cancers have been described, we hypothesized  
65 differences at the transcriptomic level in cord blood between male and female  
66 newborns as a consequence of foetal carcinogenic exposure. The objective was  
67 to investigate whether transcriptomic responses to dietary genotoxic and non-  
68 genotoxic carcinogens demonstrate gender-specific mechanisms-of-action  
69 relevant for chemical carcinogenesis.

70 Methods: Global gene expression was applied in umbilical cord blood  
71 samples, the CALUX<sup>®</sup>-assay was used for measuring dioxin(-like), androgen(-  
72 like) and estrogen(-like) internal exposure, and acrylamide-haemoglobin adduct  
73 levels were determined by mass spectrometry adduct-FIRE-procedure<sup>TM</sup>. To link  
74 gene expression to an established phenotypic biomarker of cancer risk,  
75 micronuclei frequencies were investigated.

76 Results: While exposure levels did not differ between sexes at birth,  
77 important gender-specific differences were observed in gene expressions  
78 associated with these exposures linked with cell cycle, the immune system and  
79 more general cellular processes such as (post)-translation. Moreover, oppositely  
80 correlating leukemia/lymphoma genes between male and female newborns were  
81 identified in relation to the different biomarkers of exposure which might be  
82 relevant to male-specific predisposition to develop these cancers in childhood.

83           Conclusions/Impact: This study reveals different transcriptomic responses  
84 to environmental carcinogens between the sexes. In particular, male-specific  
85 TNF-alpha-NF-kB signaling upon dioxin exposure and activation of the Wnt-  
86 pathway in boys upon acrylamide exposure might represent possible mechanistic  
87 explanations for gender specificity in the incidence of childhood leukemia.  
88

## 89 **1. Introduction**

90 A gender effect in the incidence of childhood cancers has been well  
91 described (1, 2). The male-to-female incidence varies for several types of  
92 cancers; in particular, a male predominance in the age-adjusted incidence of all  
93 types of leukemias and lymphomas is apparent, with the highest ratio (M:F: 3.0)  
94 for non-Hodgkin lymphoma (2). Potential gender-specific risk factors include the  
95 faster growth rate of the male embryo due to accelerated rates of cell division  
96 and proliferation (3). For both sexes, birth weight have been positively associated  
97 with childhood leukemia, whereas the maternal dietary intake of DNA  
98 topoisomerase II inhibitors was associated with infant leukemia, altogether, they  
99 are the most common pediatric neoplasm (4). However, the aetiology of sex-  
100 specific childhood cancer still needs to be fully clarified.

101 In this context, a link between environmental/dietary exposure to  
102 carcinogenic chemicals and infant leukemia has been proposed (5). Statistics  
103 show significant increases in the incidence of cancer including leukemia, among  
104 children in Europe over the last decades (6) suggesting that (maternal) exposure  
105 to carcinogenic factors present an aetiological factor. Thus, investigations on  
106 possible mechanisms-of-action through which exposure to environmental  
107 carcinogens predispose to childhood cancer risk, is warranted. The latent period  
108 of e.g. leukemia, its very early onset in childhood might indicate the foetal period  
109 to be a critical window of exposure. It has been suggested that such foetal  
110 exposure might lead to predisposition to develop disease such as cancer and

111 immune diseases during childhood or in later life possibly through modulation of  
112 the foetal transcriptome (7, 8).

113       Upon ingestion or inhalation by pregnant women, many environmental  
114 carcinogens are capable of crossing the placental barrier and reaching the foetus  
115 (9). It has been shown that the placental barrier does not protect the unborn child  
116 in case of maternal exposure to acrylamide, dioxins and estrogen-like  
117 compounds such as organochlorine pesticides and amphenones (10-13).

118       Over the last decade, microarray-based genomics technology has  
119 significantly progressed enabling whole genome screening at the gene  
120 expression level, making this highly suitable for investigating mechanisms  
121 underlying e.g. the molecular effects of carcinogenic exposure. Since a gender  
122 effect in the incidence of childhood cancers exists, with cancer of the blood  
123 system being far more frequent in males than females, we hypothesized that  
124 differences at the transcriptomic level in cord blood will occur between male and  
125 female newborns as a consequence of foetal carcinogenic exposure, possibly  
126 leading to gender-specific predisposition to develop childhood leukemia.

127       For this, global transcriptomic responses in umbilical cord blood samples  
128 from Norwegian male and female newborns were examined in relation to  
129 markers of carcinogenic exposure and effect. Investigated non-genotoxic  
130 carcinogen exposures determined by means of DR CALUX<sup>®</sup>, include aryl  
131 hydrocarbon (Ah) receptor-mediated compounds such as polychlorinated  
132 dibenzodioxins (PCDDs), polychlorinated dibenzofurans (PCDFs), and dioxin like  
133 polychlorinated biphenyls (PCBs) (14). Measurements by CALUX<sup>®</sup> assay of ER $\alpha$

134 and AR receptor activation represent exposure to endocrine disruptors such as  
135 carcinogenic pesticides like 1,1,1-trichloro-2,2-bis(*p*-chlorophenyl)ethane (DDT)  
136 and (anti-) androgens (15).

137 Global transcriptomic responses in male and female newborns were also  
138 examined in relation to genotoxic acrylamide exposure. Humans are exposed to  
139 relatively high doses of acrylamide since this compound is formed in food during  
140 cooking (16, 17). Acrylamide can be converted to the epoxide glycidamide by  
141 cytochrome P450 2E1 (18). Both compounds are electrophilically reactive. The  
142 mutagenicity from acrylamide exposure arises from the reaction of glycidamide  
143 with nucleophilic centres in DNA, as the direct interaction of acrylamide with DNA  
144 is slow (19). Acrylamide and glycidamide also bind to circulating proteins such as  
145 haemoglobin (Hb) and form adducts. This Hb-bound acrylamide represents a  
146 well-accepted measure of internal dose and was, as well as the Hb-bound  
147 glycidamide, used as a biomarker of exposure to acrylamide in the present study.  
148 A recently developed liquid chromatography-tandem mass spectrometry method  
149 was used for measurement of the Hb adducts (20).

150 Gene expression was associated to a validated biomarker of cancer risk  
151 i.e. micronuclei frequencies (MN). MN is a well established biomarker  
152 of chromosomal breakage and/or whole chromosome loss that are unrepaired,  
153 misrepaired or malsegregated due to chromosome, cellular and nuclear  
154 dysfunction (21-23), and increased micronucleus frequencies in peripheral blood  
155 lymphocytes have been shown in prospective studies to predict the risk of cancer  
156 in humans (24).

157



158 **2. Materials and methods**

159

160 **2.1 Sample collection**

161 Umbilical cord blood samples were collected immediately after birth from  
162 the cord vein of 200 babies whose mothers participated in the Norwegian BraMat  
163 cohort. As detailed below in section 2.3, 45 male and 66 female newborns were  
164 included in the current study (Table 1). This BraMat sub cohort is nested within  
165 the Norwegian Mother and Child cohort (MoBa). Study protocols were approved  
166 by the Regional Committee for Ethics in Medical Research in Oslo, Norway.  
167 Informed consent was obtained from all participating mothers prior to sample  
168 collection. The samples were collected by trained nurses at the maternity wards  
169 of the Ulleval and Akershus hospitals in Oslo, Norway. For preserving RNA for  
170 microarray analysis, aliquots of heparin-anticoagulated whole blood of 0.4 ml  
171 were mixed with 1.2 ml of RNAlater (Ambion/Applied Biosystems, Nieuwerkerk  
172 aan den IJssel, the Netherlands) as soon as possible after blood collection.  
173 Samples were kept at -80°C until shipment on dry ice to the research laboratory.  
174 The remainder of the whole blood was utilized for CALUX<sup>®</sup> measurements,  
175 assessment of acrylamide- and glycidamide-Hb adducts and analysis of  
176 micronuclei frequencies.

177

178 **2.2 RNA isolation**

179 Total RNA was isolated from the RNAlater-preserved whole blood  
180 samples using the RiboPure-Blood system (Ambion) according to the

181 manufacturer's instructions. RNA yield and purity was measured by means of  
182 Nanodrop equipment (Wilmington, DE, USA) and integrity was verified by  
183 automated gel electrophoresis (2100 BioAnalyzer, Agilent Technologies,  
184 Amstelveen, the Netherlands).

185

### 186 **2.3 Gene expression analysis**

187 From the 200 BraMat samples, only RNA samples with a RNA Integrity  
188 Number (RIN)  $\geq 6$  and a bench time (time from collection until RNA stabilization)  
189 of  $\leq 6$  hours, i.e. 120 samples were used for transcriptomic analysis. Total RNA  
190 (1 microgram) was used to generate Cyanine-labeled cRNA using the Agilent  
191 Low RNA Input Linear Amplification kit according to the manufacturer's  
192 instructions. Each individual cord blood sample was labelled by means of  
193 Cyanine-5 and competitively hybridized against a common reference sample  
194 (pooled RNA cord blood samples, labelled with Cyanine-3) onto Agilent 4x44k  
195 human oligonucleotide microarrays (Agilent Technologies, Palo Alto, CA, USA)  
196 according to the manufacturer's instructions. After hybridization, microarrays  
197 were washed and scanned immediately using an Axon GenePix<sup>®</sup> 4000B  
198 Microarray Scanner (Molecular Devices, Sunnyvale, USA). Laser power was set  
199 to 100%. The photo multiplier tube (PMT) gain was set to a saturation tolerance  
200 of 0.02% to minimize background and saturated spots.

201 Additional sample exclusions were based on technical performance of the  
202 microarrays, as well as on absence of informed consent forms or reported

203 smoking of the mother at 30 weeks of pregnancy. The number of cases left in the  
204 study after exclusions was 111.

205

#### 206 ***2.4 Internal measurements of pollutants***

207 Dioxin-like activity in plasma was determined in a total of 43 samples using the  
208 dioxin responsive (DR) CALUX<sup>®</sup> reporter gene bioassay. The bioassay  
209 comprises a genetically modified H4IIE rat hepatoma cell line containing the  
210 luciferase gene under control of the AhR. Approximately 3 grams of plasma from  
211 umbilical cord blood was extracted by means of shake-solvent extraction  
212 (hexane:diethylether, 97:3). Extracted fat was used for clean-up on an acid silica  
213 column (20% and 33% H<sub>2</sub>SO<sub>4</sub>), topped with sodium sulphate. Cleaned extracts  
214 were dissolved in DMSO (8 µl); the DR CALUX<sup>®</sup> activity was determined  
215 following exposure to dilution series of cleaned extracts (0.8% DMSO) for 24 hrs.  
216 Estrogenic and androgenic activity in umbilical cord blood were determined in a  
217 total of 34 and 31 samples respectively, using a human U2OS cell line stably  
218 transfected with the luciferase gene that is under control of the estrogen receptor  
219 alpha (ER $\alpha$  CALUX<sup>®</sup>) or androgen receptor (AR CALUX<sup>®</sup>). Approximately 0.5 ml  
220 of cord blood plasma was extracted by means of shake-solvent extraction  
221 (MTBE; methyltertiarbutylether). Extracts were dissolved in DMSO (40 µl); the  
222 ER $\alpha$ - and AR CALUX<sup>®</sup> activity was determined following 24 hrs of exposure at  
223 various dilutions of the re-dissolved extracts (0.1% DMSO). All bioassays were  
224 corrected for procedure blanks and performed including a reference compound  
225 calibration curve. By interpolation, final results are expressed as reference

226 compound equivalents (eq.): DR CALUX<sup>®</sup> are expressed in pg 2,3,7,8-TCDD-like  
227 eq. (TEQ)/gram fat, ER $\alpha$  CALUX<sup>®</sup> in ng 17 $\beta$ -estradiol-like eq. (EEQ)/ml plasma,  
228 AR CALUX<sup>®</sup> in ng dihydrotestosterone-like eq. (AEQ)/ml plasma (25, 26).

229

### 230 **2.5 Haemoglobin-adduct levels**

231 Hb-adducts from acrylamide (AA Hb-adducts) and glycidamide (GA Hb-  
232 adducts) were measured as internal dose markers of exposure to acrylamide  
233 from food. Red blood cells were isolated and stored at -20 C until further  
234 processing. Adduct levels to N-terminal valines in Hb were measured in a total of  
235 84 samples applying the “adduct FIRE procedure” using liquid chromatography–  
236 tandem mass spectrometry (LC–MS/MS) (Shimadzu Prominence, AB SCIEX  
237 3200 qtrap) for final detection (20). Method performance parameters and its  
238 application to measurements of acrylamide and glycidamide Hb-adducts in cord  
239 blood samples has previously been described (20, 27).

240

### 241 **2.6 Cytokinesis-block micronucleus assay**

242 Micronuclei frequencies (MN) per 1000 binucleated cells (%MNBN) were  
243 analyzed in peripheral blood T-lymphocytes of 33 samples. MNBN, in addition to  
244 *in vivo* accumulated MN, reflect damage present on DNA or key proteins and  
245 expressed as MN during *in vitro* cell division. MN in mononucleated cells  
246 (MNMONO) only reflect chromosomal damage before the start of the assay.  
247 Since gene expression is not purely a cumulative measure, and MNMONO  
248 frequencies were low, MNBN were used. The *in vitro* cytokinesis blocked MN

249 assay (CBMN assay) was carried out according to the standardized protocol  
250 developed for semi-automated image analysis system (Decordier et al. 2009).  
251 Sampling and making of cultures occurred in Oslo, Norway. The protocol was  
252 performed as recently described by Vande Loock et al (28).

253

## 254 **2.7 Data analysis**

255 The data discussed in this publication have been deposited in NCBI's  
256 Gene Expression Omnibus (29) and are accessible through GEO Series  
257 accession number GSE31836 (30). Scan images of Cy5- and Cy3- channels  
258 were loaded into ImaGene software version 8.0.1 (BioDiscovery, El Segundo,  
259 USA) for the extraction of raw pixel intensities and local backgrounds. The data  
260 were preprocessed using the Refiner Array module from the Genedata  
261 Expressionist system (Genedata AG, Basel, Switzerland). Raw data were  
262 assessed for quality and preprocessed as follows. Signals were corrected by  
263 subtracting backgrounds without producing negative values. For each feature  
264 and array, the contrast, i.e., the quality for each signal according to its signal-to-  
265 noise ratio, the distortion and imbalance were determined. Defective or saturated  
266 features detected by the scanner software or by the Refiner Array module were  
267 diagnosed, and masked. LOWESS correction was used to correct all features so  
268 that the signal distortion and imbalance of the two channels is minimized. Quality  
269 criteria for the data analysis were set as follows: signal-to-noise ratios >2, relative  
270 errors <0.5, saturated and features flagged as masked filtered out.

271 Dose-response relationships were investigated with the available samples  
272 by Pearson's correlation analyses of individual gene expressions (Cy5/Cy3  
273 ratios) and individual values for CALUX<sup>®</sup> measurements, haemoglobin-adducts  
274 and micronuclei frequencies, as mentioned in Table 2. Correlation analyses were  
275 performed only for transcripts with least 50 % valid expression values

276 Significantly correlating transcripts were selected using two cut-off's.  
277 Minimal correlation coefficients were set to  $\geq 0.60$  or  $\leq -0.60$  and an unadjusted p-  
278 value of  $< 0.05$ . To ensure maximum specificity, a higher cut-off of  $\geq 0.75$  or  $\leq -0.75$   
279 was used. No false discovery rates were used, since correlation coefficients were  
280 considered more important.

281 To identify biological processes in which significantly correlating genes are  
282 involved, the software suite Metacore<sup>™</sup> was used. Gene Ontology (GO) terms,  
283 pathways and gene networks containing at least two significantly correlating  
284 genes and a p-value  $\leq 0.01$  were considered significantly enriched. In addition, a  
285 false discovery rate cut-off of 0.25 was applied. For enabling further biological  
286 interpretation of correlation data, correlation coefficients were uploaded in Tox-  
287 Profiler without any statistical pre-selection of correlating genes. Tox-Profiler  
288 uses the T-test to score differences between mean input values (usually  
289 expression values, but in our case correlation coefficients) of predefined gene  
290 sets and that of all other genes (31). For biological interpretation, gene sets  
291 based on annotations from different databases were used, i.e. GO terms, KEGG,  
292 Biocarta and WikiPathways. To identify significant effects, a Wilcoxon non-  
293 parametric statistical test was used, using an unadjusted E-value cut-off of 0.05.

294 To evaluate differences at the transcriptomic level between male and  
295 female newborns in relation to blood cancer risk, the effects on human cancer  
296 genes, i.e. genes in which mutations that are causally implicated in oncogenesis  
297 have been reported, were investigated (32). For this, only genes connected to  
298 leukemia and lymphoma were included which showed an opposite correlation  
299 coefficient between male and female neonates. Additionally, the correlation  
300 coefficient had to be  $\geq 0.60$  or  $\leq -0.60$  in either male or female newborns.  
301

302 **3. Results**

303

304 **3.1 Biomarker analyses**

305 From a total of 200 samples from the BraMat cohort, 120 RNA samples  
306 were selected based on a RNA Integrity Number (RIN)  $\geq 6$  and a bench time (time  
307 from collection until RNA stabilization) of  $\leq 6$  hours. These 120 samples were  
308 hybridized onto microarrays. Sample/microarray exclusions were based on  
309 technical performance of the microarrays, as well as on absence of informed  
310 consent forms or reported smoking of the mother at 30 weeks of pregnancy. The  
311 number of cases left in the study after exclusions was 111. Characteristics of the  
312 remaining study population are presented in Table 1A, for the whole study  
313 population as well as for male and female infants separately. All mothers of the  
314 included 111 newborns are non-smokers. Table 1B presents RNA Integrity  
315 Numbers (RIN) and bench time, i.e. time from collection until RNA stabilization,  
316 while Table 1C presents the data distributions for the investigated parameters on  
317 CALUX<sup>®</sup>, Hb-adducts and micronuclei frequencies. The number of available  
318 samples for each biomarker varies as presented in Table 1C. For all these  
319 measurements, no significant differences were observed between male and  
320 female newborns.

321

322 **3.2 Number of significantly correlating genes**



323 Correlation analyses of gene expression levels with biomarkers of  
324 exposure and with MN measured showed variable numbers of significantly  
325 correlating genes (Table 2). Due to missing values, the numbers of available  
326 samples for correlation analysis vary per biomarker and were significantly lower  
327 than the 111 mentioned in Table 1. Overall, separate analyses for male and  
328 female newborns resulted in higher numbers of significantly correlating genes per  
329 gender with low overlap of similarly expressed genes between the two sexes,  
330 thus indicating a clear gender-specific toxicogenomic response.

331

### 332 **3.3 Biological evaluation**

333

#### 334 **3.3.1 DR CALUX<sup>®</sup>**

335 Correlation analysis demonstrated that male infants show a higher number  
336 of significantly correlating transcripts in relation to dioxin and dioxin-like  
337 exposure, assessed by DR CALUX<sup>®</sup> assay, when compared to females. For  
338 males, functional interpretation revealed significantly affected processes relevant  
339 to carcinogenicity, such as programmed cell death/apoptosis, spindle/nuclear  
340 division and the ubiquitination–proteasome pathway (Table 3). In females, mainly  
341 epigenetic responses were found, such as nucleotide and chromatin assembly  
342 (Table 4). The mitotic cell cycle appeared to be affected in both males and  
343 females.

344 Tox-Profiler results showed more processes to be significantly affected in  
345 female neonates compared to males (Table 5). Female-specific processes

346 included translation and post-translational modification. Tox-Profiler also  
347 confirmed the epigenetic responses in females, although these were also (not-  
348 significantly) positively correlated in males. T and B cell receptor Signaling  
349 Pathways were found to be suppressed in females. Translational processes were  
350 also found for male newborns. Interestingly, these were oppositely regulated  
351 when compared to females. Also, proteasome degradation was found to be  
352 oppositely regulated between the sexes as well as the TNF-alpha-NF-kB  
353 Signaling Pathway.

354

### 355 **3.3.2 ER $\alpha$ CALUX<sup>®</sup>**

356 Female newborns showed a higher number of significantly correlating  
357 transcripts in relation to estrogen and estrogen-like exposure when compared to  
358 males. Accordingly, more processes were found to be significantly affected in  
359 females, of which the majority was immune-related. Other processes included  
360 more general (post-) translation events, while estrogen (-like) exposure also  
361 appeared to affect apoptosis Table 4). Tox-Profiler-based analysis in particular  
362 showed the immune response to be positively correlated in females. Also, more  
363 general cellular processes mostly related to transcription and post-translational  
364 processes were identified. Interestingly, all processes found using Tox-Profiler  
365 were oppositely regulated between the sexes, although mainly reaching  
366 significance in females only (Table 5).

367

### 368 **3.3.3 AR CALUX<sup>®</sup>**

369 Male newborns showed a higher number of significantly correlating genes with  
370 AR CALUX<sup>®</sup> involved in immune response, cell cycle regulation, ubiquitin-  
371 proteasomal proteolysis and cytoskeletal-related processes (Table 3). Functional  
372 enrichment analyses on significantly correlating genes for females resulted in an  
373 extensive list comprising general cellular processes like (post-) translation,  
374 metabolic and biosynthesis processes (Table 4). This finding is confirmed by the  
375 Tox-Profiler analysis, which also shows general cellular processes to be  
376 significantly affected in female newborns (Table 5).

377

#### 378 **3.3.4 Haemoglobin adduct level**

379 Analyses revealed only weak correlation of GA Hb-adducts with gene  
380 expression, leaving too few significantly correlating genes to perform biological  
381 interpretation analyses. For AA Hb-adducts, in particular for males, regulation of  
382 developmental process, exocytosis and regulation of T-cell activation and neuron  
383 differentiation was found (Tables 3 and 4). Tox-Profiler results revealed general  
384 processes oppositely regulated between the two sexes, although significance  
385 was generally found for females, except for mRNA splicing, which was  
386 significantly (negatively) correlated with AA Hb-adducts in males (Table 5).

387

#### 388 **3.3.5 Micronuclei frequency**

389 Male newborns showed a higher number of genes significantly correlating  
390 with micronuclei frequencies, when compared to female newborns (1397 vs 95;  
391 Table 2). Functional enrichment analyses using a correlation coefficient cut-off of

392 0.60 identified general cellular processes like translation and RNA processing,  
393 next to a number of immune-related processes. A higher cut-off of 0.75,  
394 decreasing the number of genes to 270 and thus increasing specificity, resulted  
395 in more specific processes such as nuclear fragmentation during apoptosis and  
396 cell cycle regulation (Table 3). Despite the limited number of significantly  
397 correlating genes for female newborns, cell cycle regulation and DNA damage  
398 removal appeared to be enriched (Table 4). Tox-Profiler analysis confirmed the  
399 significant correlation of immune-related processes in males. Also, the  
400 translational and post-translational processes found by using significantly  
401 correlating transcripts were confirmed, showing negatively correlation with  
402 micronuclei frequencies in males, where a significant positive correlation was  
403 found in females.

404 No significant correlations were observed between micronuclei  
405 frequencies and the exposure markers when the subjects were grouped together  
406 as well as for females separately. Only in males, acrylamide and glycidamide  
407 hemoglobin adducts correlated significantly with micronuclei frequencies  
408 (CC=0.75 with  $p=0.019$  and CC=0.73 with  $p=0.025$ , respectively).

409

### 410 ***3.3.6 Inversely correlating effects on leukemia- and lymphoma-associated*** 411 ***genes***

412 Finally, to evaluate differences at the transcriptomic level between male  
413 and female newborns in relation to blood cancer risk, we investigated the effects  
414 on human cancer genes. Figure 1 presents the top four (based on deltaCC)

415 oppositely correlating cancer genes between male and female newborns in  
416 association with the biomarkers. In total, four transcripts correlated inversely in  
417 relation to DR CALUX<sup>®</sup>, 13 in relation to ER $\alpha$  CALUX<sup>®</sup>, 7 in relation to AR  
418 CALUX<sup>®</sup>, 18 in relation to micronuclei frequencies, 2 in relation to acrylamide and  
419 none in relation to glycidamide (supplementary data Table S4).  
420

## 421 **4. Discussion**

422           Since a gender effect in the incidence of childhood cancers is known to  
423 exist, we hypothesized differences at the transcriptomic level in cord blood  
424 between male and female newborns as a consequence of foetal carcinogenic  
425 exposure. Concordantly, different transcriptomic responses to environmental  
426 carcinogens between the sexes were observed. Taking into account that the  
427 largest differences between sexes in childhood cancer are found for leukemias  
428 and lymphomas (2), and that we measured transcriptomics, exposure biomarkers  
429 and effect biomarkers in blood of newborns, this discussion will focus on the  
430 relation between transcriptome data and childhood risks for acute lymphoblastic  
431 leukemia (ALL).

432

### 433 **4.1 DR CALUX<sup>®</sup>**

434           The non-genotoxic carcinogen exposures determined by means of DR  
435 CALUX<sup>®</sup>, comprise compounds that activate the Ah receptor, such as PCDDs,  
436 PCDFs, and dioxin like PCBs (14). In our study, far more genes correlated with  
437 DR CALUX<sup>®</sup> in males than in females (Table 2). In addition, functional  
438 interpretation demonstrates large gender differences in dose-dependent  
439 transcriptomics responses.

440           An epigenetic response to Ah-receptor-mediated exposure was found,  
441 showing nucleosome-related processes positively correlating with DR CALUX<sup>®</sup>  
442 mainly in females. Effects on the nucleosome are in concordance with the known  
443 mechanism-of-action of dioxins; activation of transcription by dioxin is

444 accompanied by changes in chromatin structure, which depend upon a functional  
445 aromatic hydrocarbon (Ah) receptor (33). It is, however, unclear why this is sex-  
446 specific and how this could affect ALL susceptibility. The (mitotic) cell cycle was  
447 mainly affected in males. Although it is generally accepted that Ah receptor  
448 activation causes toxic and carcinogenic effects through its role in detoxification  
449 by modulating expression of metabolizing enzymes, activation of the Ah receptor  
450 possibly also participates in pathways such as cell cycle regulation, apoptosis  
451 and immune response (34). Possibly, the activation of cell cycle regulating genes  
452 via the Ah-receptor in male neonates is stronger than in females, and therefore  
453 males are more susceptible to develop ALL due to exposure to PCB and other  
454 Ah-receptor activators.

455 T and B cell Receptor Signaling Pathways are negatively correlated with  
456 dioxin exposure in female neonates. Foetal TCDD exposure in animals has been  
457 linked to inhibition of cellular differentiation and maturation, primarily targeting T  
458 lymphocytes, leading to thymic atrophy and immunosuppression in offspring (35).  
459 This may suggest that the immune system is more suppressed in females  
460 exposed to Ah-activators, than in males. That would imply a higher risk in  
461 females, which is not the case in childhood ALL. However, a higher incidence  
462 has been reported in female infants (36) possibly suggesting T and B cell  
463 Receptor Signaling Pathways to mechanistically underlie the increased risk of  
464 developing of infant ALL.

465 Interestingly, the TNF-alpha-NF-kB Signaling Pathway is negatively  
466 correlated with dioxin exposure in females, while (not significantly) positively

467 correlated in male newborns. The association of inflammation and cancer has  
468 been studied extensively (37) showing TNF- $\alpha$  acts as a key mediator of  
469 inflammation and cancer (38). Although this pathway is involved in general  
470 cancer susceptibility, NF-kB activation has been consistently associated with  
471 childhood ALL (39, 40). The involvement of the NF-kB pathway has been  
472 suggested to play a role in the sex-specific character of childhood leukemia  
473 based on the association of the *IRF4* gene with higher male-specific risk with  
474 childhood ALL (41). IRF4 was however not found to correlate with dioxin  
475 exposure although we found several other genes showing differential gender-  
476 specific correlation within the TNF-alpha-NF-kB Signaling Pathway, including e.g.  
477 *TRAF1*, *TRAF3*, *MAP3k3*, *MAP3k14* and *CASP8*. This might further indicate the  
478 TNF-alpha-NF-kB Signaling Pathway as a possible contributing factor in relation  
479 to the male predominance in childhood ALL. Moreover, the unaffected or even  
480 non-significantly increased signaling by tumor necrosis factor (TNF)- $\alpha$  in males  
481 might also result in a failure of the lymphocytes to undergo apoptosis (42),  
482 possibly contributing to the increased risk of childhood ALL in males. This is  
483 underlined by the male-specific negative correlation of *CASP8* with dioxin  
484 exposure. Accordingly, apoptosis was found to be exclusively affected in males.  
485 Nine human blood cancer-associated genes were found to be inversely  
486 correlating strongly indicating a gender-specific response to dioxins in relation to  
487 tumorigenesis. The human anti-apoptotic cancer genes *JAK2* and *VHL* are  
488 positively correlated with dioxin exposure in males, while negatively in females  
489 contributing to apoptosis blockade, possibly predisposing males to develop



490 childhood ALL. These cancer genes have been associated with childhood ALL  
491 (43-44).

492 In conclusion, the female-specific suppression of the TNF- $\alpha$ -NF- $\kappa$ B  
493 Signaling Pathway upon dioxin exposure might protect female neonates from  
494 developing childhood ALL, while their male counterparts are at higher risk,  
495 possibly due to consequent blockade of apoptosis.

496

#### 497 **4.2 ER $\alpha$ CALUX<sup>®</sup>**

498 Estrogens have been linked to B and T cell activation, enhanced antigen  
499 presentation by dendritic cells, and cytokine production (45). Accordingly, we  
500 found immune-related processes to be affected, with a higher response in  
501 females compared to males. In fact, results using unfiltered correlation data  
502 suggest an inverse regulation upon estrogen receptor activation with a significant  
503 stimulation of the GO term 'immune system' in females and (non-significant)  
504 suppression in males. GO terms like B and T cell signalling show similar inverse  
505 trends. The male-specific immune suppression may contribute to the higher  
506 incidence of childhood ALL since leukemia is associated with immune  
507 dysfunction (47). Further confirming gender-specificity, we found leukemia- and  
508 lymphoma-related genes differentially correlating with estrogen exposure (Table  
509 7). Not all genes conclusively point to a male predisposition for developing ALL,  
510 in fact we found a female-specific positive correlation of *MLL1*. Mixed lineage  
511 leukemias are histone methyl-transferases that regulate gene activation and are  
512 also well known to be rearranged in acute myeloid and lymphoid leukemias (47).

513 Interestingly, the female-specific correlation might be in line with the literature:  
514 Unlike childhood leukemia diagnosed in childhood, where there is a  
515 predominance in males, high frequencies of MLL rearrangements in female  
516 Infant Leukemia have been reported, as well as a higher risk for females to  
517 develop infant ALL and (36, 48).

518

### 519 **4.3 AR CALUX<sup>®</sup>**

520 Next to androgens, certain environmental pollutants may influence AR  
521 signaling pathways by acting as or interfering with endogenous androgens,  
522 regulating the expression of various genes involved in cell  
523 cycle control, apoptosis, cell growth and differentiation (49). We found the cell  
524 cycle GO term to correlate significantly to AR CALUX<sup>®</sup> in males. Possibly, the  
525 activation of cell cycle genes via the AR-receptor in male neonates is stronger  
526 than in females, and leading to a higher males-specific susceptibility. In contrast,  
527 the immune response was significantly suppressed in only females implying a  
528 higher cancer risk in females. Concordantly, differentially correlating leukemia  
529 and lymphoma genes with AR receptor activation do not conclusively support a  
530 male predisposition. In fact, next to the *MLLT6* gene correlating with the ER $\alpha$   
531 receptor, a female-specific positive correlation of *MIIT3* with AR receptor  
532 activation was found which might be related to female predisposition to develop  
533 infant leukemia (36, 47, 48).

534

### 535 **4.4 Hb-adducts**

536 Tox-Profiler revealed the Wnt signaling pathway and a number of general  
537 processes oppositely regulated between the two sexes upon acrylamide  
538 exposure. Aberrations of the Wnt pathway have been associated with various  
539 types of cancer including ALL. A male-specific activation of the Wnt pathway  
540 might thus contribute to the higher incidence of cancer in the blood system.

541

#### 542 **4.5 Micronuclei frequencies**

543       Micronuclei formation in peripheral blood T-lymphocytes may be regarded  
544 as a phenotypic endpoint that reflects the deleterious effects on chromosomes  
545 due to the (combined) exposure to genotoxic carcinogens (50). Previous studies  
546 report that the baseline MN frequency is relatively low in newborns and no  
547 differences in gender have been identified (28, 51), which is confirmed in our  
548 study. Despite this, the sex-specific transcriptomic profiles associated with  
549 micronuclei frequencies differ dramatically. Overall, a higher number of  
550 significantly correlating genes in male newborns in relation to MN was observed.  
551 Correlations were frequently opposite between males and females. The male-  
552 specific profile contains many more genes relevant to ALL and carcinogenicity in  
553 general. This might contribute to the higher childhood cancer risk in males.  
554 Interestingly, 6 genes of our MN correlating genes are also present in the  
555 recently published MN gene network (52). They were found to be oppositely  
556 correlating between male and female neonates, i.e. *BCL2*, *CDC20*, *PTTG1*, *BAX*,  
557 *PCNA*, and *LMNA*, suggesting sex-specific contribution to the network. Lastly,  
558 MN-associated sex-specific differences in general cellular processes such as

559 transcription, post-translational processes, RNA processing and splicing might be  
560 of relevance with regard to the reported gender differences in the incidence of  
561 childhood cancers, however interpreting these findings is difficult.

562

#### 563 **45. Conclusions**

564 While exposure to acrylamide and endocrine disruptors did not differ  
565 significantly between sexes at birth, distinct gender-specific dose-response  
566 related transcriptomic profiles were identified. In particular, male-specific TNF-  
567 alpha-NF-kB signaling upon dioxin exposure and subsequent failure of the  
568 lymphocytes to undergo apoptosis, and activation of the Wnt pathway in boys  
569 upon acrylamide exposure might represent possible mechanistic explanations for  
570 the gender-specificity in the incidence of childhood ALL. Interestingly, oppositely  
571 correlating leukemia and lymphoma genes between male and female newborns  
572 were identified in relation to the different biomarkers of exposure which might be  
573 relevant to male-specific predisposition to develop childhood ALL and female-  
574 specific predisposition to develop infant ALL .

575 Our study has the strength of considering multiple parameters in relation  
576 to gene expression, although we recognize that the correlation analyses in the  
577 current study of gene expression with the various biomarkers are based on a  
578 relatively small number of samples making multivariate correlation analyses  
579 impractical. This is due to the limited availability of samples with biomarker  
580 measurements and gender-specific analyses, which reduces statistical power.  
581 Nevertheless, since the gender-specific responses occur for multiple biomarkers,

582 we are confident about our conclusions. To confirm the generated hypotheses,  
583 future research based on higher number of samples is needed.

584

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592

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752

753 **Tables**

754 Table 1. Characteristics of the study population (A), RNA Integrity Number and  
755 bench time (B) and data distribution of the various parameters (C).

756

757 Table 2. Numbers of genes correlating with CALUX<sup>®</sup>, Hb-adducts and  
758 micronuclei.

759

760 Table 3. The GO terms, pathways and networks enriched among the significantly  
761 correlating genes for the various biomarkers for males

762

763 Table 4. The GO terms, pathways and networks enriched among the significantly  
764 correlating genes for the various biomarkers for females

765

766 Table 5. GO terms and pathways enriched for males among all correlating genes  
767 using Tox-Profiler.

768

769

770 Figure 1. Human leukemia and lymphoma associated genes with opposite  
771 correlations with biomarkers between male and female newborns. Only the top 4,  
772 based on deltaCC is presented; for complete lists refer to supplementary data  
773 Table S4. \* indicates a significant correlation, with an unadjusted p-value < 0.05.  
774 \*\* indicates the query terms used to search the GO term annotations for each  
775 gene.



Table 1. Characteristics of the study population (A), RNA Integrity Number and bench time (B) and data distribution of the various parameters (C).

	All	Males	Females
<b>A</b>			
N	111	45	66
Birth weight	3629 ± 434	3722 ± 416	3564 ± 438
Gestation	39.8 ± 1.1	39.9 ± 1.1	39.7 ± 1.1
Age - mother	31.3 ± 4.2	31.0 ± 4.3	31.5 ± 4.2
- father	32.8 ± 6.4	32.3 ± 4.3	33.7 ± 5.7
Mother BMI BP	23.4 ± 3.9	23.1 ± 3.6	23.6 ± 4.1
Mother was pregnant before			
- no	35%	34%	36%
- yes	65%	66%	64%
Mother smoked BP			
- no	84%	82%	85%
- occasional	9%	11%	8%
- daily	7%	7%	6%
Delivery start			
- Spontaneous	81%	86%	77%
- Induced	12%	7%	16%
- Caesarean section	7%	7%	8%
Caesarean sections			
- Planned	7	2	5
- Emergency	7	4	3
<b>B</b>			
RIN (0-10)	8.0 ± 1.3	7.98 ± 1.30	7.97 ± 1.30
Bench time	77.8 ± 172.4	58.77 ± 116.66	90.63 ± 201.29
<b>C</b>			
DR CALUX <sup>®</sup>	36.4 ± 9.8 (n=43)	37.4 ± 10.5 (n=14)	35.9 ± 9.6 (n=29)
ERα CALUX <sup>®</sup>	15.6 ± 14.4 (n=34)	16.1 ± 10.6 (n=17)	15.2 ± 17.8 (n=17)

AR CALUX <sup>®</sup>	0.08 ± 0.06 (n=31)	0.07 ± 0.03 (n=16)	0.09 ± 0.08 (n=15)
AA Hb-adducts	16.5 ± 6.6 (n=84)	16.0 ± 7.1 (n=34)	16.7 ± 6.4 (n=50)
GA Hb-adducts	10.0 ± 4.0 (n=84)	9.5 ± 3.8 (n=34)	10.3 ± 4.0 (n=50)
%MNBN	1.2 ± 0.9 (n=33)	1.1 ± 0.8 (n=13)	1.2 ± 0.9 (n=16)

Data are presented as mean ± SD unless stated otherwise. BP: before pregnancy, RIN: RNA Integrity Number, Bench time: time from blood collection to RNA stabilisation. Birth weight expressed in g, gestation in weeks, age in years, bench time in minutes, DR CALUX<sup>®</sup> in pg TEQ/gr fat, ER $\alpha$  CALUX<sup>®</sup> (ng EEQ/ml plasma), AR CALUX<sup>®</sup> (ng AEQ/ml plasma), Hb-adducts in pmol/g Hb, %MNBN in MN per 1000 binucleated cells.

Table 2. Numbers of genes correlating with CALUX<sup>®</sup>, Hb-adducts and micronuclei.

	# of genes	Pos. correlation	Neg. correlation	Gender	# of genes	Pos. correlation	Neg. correlation	Overlap boys-girls
DR CALUX <sup>®</sup>	<b>3</b>	0	3	Boys (n=14) Girls (n=29)	<b>371</b> <b>39</b>	163 29	208 10	1
ER $\alpha$ CALUX <sup>®</sup>	<b>20</b>	14	6	Boys (n=17) Girls (n=17)	<b>493</b> <b>626</b>	311 459	182 167	3
AR CALUX <sup>®</sup>	<b>83</b>	69	14	Boys (n=16) Girls (n=15)	<b>1293</b> <b>508</b>	946 366	347 142	29
AA Hb-adducts	<b>0</b>	0	0	Boys (n=34) Girls (n=50)	<b>23</b> <b>0</b>	18 0	5 0	0
GA Hb-adducts	<b>0</b>	0	0	Boys (n=34) Girls (n=50)	<b>6</b> <b>0</b>	2 0	4 0	0
%MNBN	<b>22</b>	4	18	Boys (n=13) Girls (n=16)	<b>1397</b> <b>95</b>	662 28	735 67	1

Numbers are presented for sexes grouped together and separately, correlation coefficients >0.60 or <-0.60 and an unadjusted p-value  $\leq$ 0.05).

Table 3. The GO terms, pathways and networks enriched among the significantly correlating genes for the various biomarkers for males

GO term / Pathway / Network	# significant processes (CC≥0.60/ CC≥0.75)	P-value (# genes affected/total # in pathway)	Database
DR CALUX <sup>®</sup>	29/16		
cell cycle		3.76E-04 (31/679)	GO
apoptosis		5.63E-04 (26/543)	GO
ERα CALUX <sup>®</sup>	50/9		
antigen processing and presentation		2.92E-05 (9/61)	GO
regulation of adaptive immune response		1.69E-04 (8/60)	GO
AR CALUX <sup>®</sup>	7/1		
Cell cycle_G1-S		6.64E-03 (17/122)	Pathways
Proteolysis_Ubiquitin-proteasomal proteolysis		7.11E-03 (19/143)	Pathways
AA Hb-adducts	4/0		
regulation of developmental process		5.51E-03 (4/637)	GO
regulation of T cell activation		8.89E-03 (2/121)	GO
GA Hb-adducts	0/0		
%MNBN	22/53		
DNA damage_Checkpoint		9.22E-03 (16/92)	Networks
regulation of cell cycle <sup>a</sup>		3.49E-04 (14/314)	GO
Apoptosis and survival_p53-dependent apoptosis <sup>a</sup>		1.06E-03 (4/25)	Pathways

Only processes mentioned in the text are presented; for complete lists refer to supplementary data Table S1.

<sup>a</sup>Found when using the 0.75 correlation coefficient cut-off.

Table 4. The GO terms, pathways and networks enriched for males among the significantly correlating genes for the various biomarkers for females

GO term / Pathway / Network	# significant processes (CC $\geq$ 0.60/ CC $\geq$ 0.75)	P-value (# genes affected/total # in pathway)	Database
DR CALUX <sup>®</sup>	18/0		
nucleosome assembly		1.22E-05 (4/69)	GO
Cell cycle_ Initiation of mitosis		6.37E-03 (2/18)	Pathways
ER $\alpha$ CALUX <sup>®</sup>	74/88		
immune system process		8.49E-06 (58/863)	GO
regulation of lymphocyte activation		1.99E-05 (18/155)	GO
AR CALUX <sup>®</sup>	39/33		
translational elongation		8.72E-19 (28/104)	GO
ubiquitin-dependent protein catabolic process		4.70E-06 (21/214)	GO
AA Hb-adducts	0/0		
GA Hb-adducts	0/0		
%MNBN	3/6		
nucleotide-excision repair, DNA damage removal		1.30E-04 (3/18)	GO
Cell cycle_ Initiation of mitosis		6.37E-03 (2/18)	Pathways

Only processes mentioned in the text are presented; for complete lists refer to supplementary data Table S2.

Table 5. GO terms and pathways enriched for males among all correlating genes using Tox-Profiler.

Biomarker	Process	# significant		T-value males	p-value males	T-value females	p-value females	Database
		processes	males/females					
DR CALUX <sup>®</sup>			5/29					
	nucleosome assembly			4.1	0.154	6.4	<0.001	GO
	T cell receptor signaling pathway			-2.7	1.000	-4.5	0.001	KEGG
	B cell receptor signaling pathway			-0.6	1.000	-4.1	0.005	KEGG
	TNF-alpha-NF-kB Signaling Pathway			2.9	0.551	-4.2	0.010	WikiPathways
ERα CALUX <sup>®</sup>			2/41					
	immune response			-1.4	1.000	6.8	<0.001	GO
	T Cell Receptor Signaling Pathway			-1.5	1.000	6.8	<0.001	WikiPathways
	B Cell Receptor Signaling Pathway			-1.9	1.000	4.9	<0.001	WikiPathways
AR CALUX <sup>®</sup>			12/24					
	immune response			-2.5	1.000	-6.6	<0.001	GO
	ubiquitin-dependent protein catabolic process			0.9	1.000	5.9	<0.001	GO
	nucleosome			-4.7	0.020	-1.2	1.000	GO
AA Hb-adducts			4/17					
	translational elongation			-1.7	1.000	8.8	<0.001	GO
	RNA splicing			-5.6	<0.001	1.7	1.0000	GO
GA Hb-adducts			8/12					
	Wnt signaling pathway			4.2	0.032	0.2	1.000	KEGG

%MNBN		30/13				
	translational elongation	8.5	<0.001	-5.7	<0.001	GO
	Spliceosome	4.6	0.002	-5.4	<0.001	KEGG
	mRNA processing	4.0	0.012	-4.1	0.002	WikiPathways
	Pathways in cancer	4.8	0.001	1.8	1.000	KEGG
	translational elongation	8.5	<0.001	-5.7	<0.001	GO

Only processes mentioned in the text are presented; for complete lists refer to supplementary data Table S3.





1 **Tables**

2 Table 1. Characteristics of the study population (A), RNA Integrity Number and  
3 bench time (B) and data distribution of the various parameters (C).

4

5 Table 2. Numbers of genes correlating with CALUX<sup>®</sup>, Hb-adducts and  
6 micronuclei.

7

8 Table 3. The GO terms, pathways and networks enriched among the significantly  
9 correlating genes for the various biomarkers for males

10

11 Table 4. The GO terms, pathways and networks enriched among the significantly  
12 correlating genes for the various biomarkers for females

13

14 Table 5. GO terms and pathways enriched for males among all correlating genes  
15 using Tox-Profiler.

16

17 Table 6. Human leukemia and lymphoma associated genes with opposite  
18 correlations with biomarkers between male and female newborns

Table 1. Characteristics of the study population (A), RNA Integrity Number and bench time (B) and data distribution of the various parameters (C).

	All	Males	Females
<b>A</b>			
N	111	45	66
Birth weight	3629 ± 434	3722 ± 416	3564 ± 438
Gestation	39.8 ± 1.1	39.9 ± 1.1	39.7 ± 1.1
Age - mother	31.3 ± 4.2	31.0 ± 4.3	31.5 ± 4.2
- father	32.8 ± 6.4	32.3 ± 4.3	33.7 ± 5.7
Mother BMI BP	23.4 ± 3.9	23.1 ± 3.6	23.6 ± 4.1
Mother was pregnant before			
- no	35%	34%	36%
- yes	65%	66%	64%
Mother smoked BP			
- no	84%	82%	85%
- occasional	9%	11%	8%
- daily	7%	7%	6%
Delivery start			
- Spontaneous	81%	86%	77%
- Induced	12%	7%	16%
- Caesarean section	7%	7%	8%
Caesarean sections			
- Planned	7	2	5
- Emergency	7	4	3
<b>B</b>			
RIN (0-10)	8.0 ± 1.3	7.98 ± 1.30	7.97 ± 1.30
Bench time	77.8 ± 172.4	58.77 ± 116.66	90.63 ± 201.29
<b>C</b>			
DR CALUX <sup>®</sup>	36.4 ± 9.8 (n=43)	37.4 ± 10.5 (n=14)	35.9 ± 9.6 (n=29)
ERα CALUX <sup>®</sup>	15.6 ± 14.4 (n=34)	16.1 ± 10.6 (n=17)	15.2 ± 17.8 (n=17)

AR CALUX <sup>®</sup>	0.08 ± 0.06 (n=31)	0.07 ± 0.03 (n=16)	0.09 ± 0.08 (n=15)
AA Hb-adducts	16.5 ± 6.6 (n=84)	16.0 ± 7.1 (n=34)	16.7 ± 6.4 (n=50)
GA Hb-adducts	10.0 ± 4.0 (n=84)	9.5 ± 3.8 (n=34)	10.3 ± 4.0 (n=50)
%MNBN	1.2 ± 0.9 (n=33)	1.1 ± 0.8 (n=13)	1.2 ± 0.9 (n=16)

Data are presented as mean ± SD unless stated otherwise. BP: before pregnancy, RIN: RNA Integrity Number, Bench time: time from blood collection to RNA stabilisation. Birth weight expressed in g, gestation in weeks, age in years, bench time in minutes, DR CALUX<sup>®</sup> in pg TEQ/gr fat, ER $\alpha$  CALUX<sup>®</sup> (ng EEQ/ml plasma), AR CALUX<sup>®</sup> (ng AEQ/ml plasma), Hb-adducts in pmol/g Hb, %MNBN in MN per 1000 binucleated cells.

Table 2. Numbers of genes correlating with CALUX<sup>®</sup>, Hb-adducts and micronuclei.

	# of genes	Pos. correlation	Neg. correlation	Gender	# of genes	Pos. correlation	Neg. correlation	Overlap boys-girls
DR CALUX <sup>®</sup>	<b>3</b>	0	3	Boys (n=14) Girls (n=29)	<b>371</b> <b>39</b>	163 29	208 10	1
ER $\alpha$ CALUX <sup>®</sup>	<b>20</b>	14	6	Boys (n=17) Girls (n=17)	<b>493</b> <b>626</b>	311 459	182 167	3
AR CALUX <sup>®</sup>	<b>83</b>	69	14	Boys (n=16) Girls (n=15)	<b>1293</b> <b>508</b>	946 366	347 142	29
AA Hb-adducts	<b>0</b>	0	0	Boys (n=34) Girls (n=50)	<b>23</b> <b>0</b>	18 0	5 0	0
GA Hb-adducts	<b>0</b>	0	0	Boys (n=34) Girls (n=50)	<b>6</b> <b>0</b>	2 0	4 0	0
%MNBN	<b>22</b>	4	18	Boys (n=13) Girls (n=16)	<b>1397</b> <b>95</b>	662 28	735 67	1

Numbers are presented for sexes grouped together and separately, correlation coefficients >0.60 or <-0.60 and an unadjusted p-value  $\leq$ 0.05).

Table 3. The GO terms, pathways and networks enriched among the significantly correlating genes for the various biomarkers for males

GO term / Pathway / Network	# significant processes (CC≥0.60/ CC≥0.75)	P-value (# genes affected/total # in pathway)	Database
DR CALUX <sup>®</sup>	29/16		
cell cycle		3.76E-04 (31/679)	GO
apoptosis		5.63E-04 (26/543)	GO
ERα CALUX <sup>®</sup>	50/9		
antigen processing and presentation		2.92E-05 (9/61)	GO
regulation of adaptive immune response		1.69E-04 (8/60)	GO
AR CALUX <sup>®</sup>	7/1		
Cell cycle_G1-S		6.64E-03 (17/122)	Pathways
Proteolysis_Ubiquitin-proteasomal proteolysis		7.11E-03 (19/143)	Pathways
AA Hb-adducts	4/0		
regulation of developmental process		5.51E-03 (4/637)	GO
regulation of T cell activation		8.89E-03 (2/121)	GO
GA Hb-adducts	0/0		
%MNBN	22/53		
DNA damage_Checkpoint		9.22E-03 (16/92)	Networks
regulation of cell cycle <sup>a</sup>		3.49E-04 (14/314)	GO
Apoptosis and survival_p53-dependent apoptosis <sup>a</sup>		1.06E-03 (4/25)	Pathways

Only processes mentioned in the text are presented; for complete lists refer to supplementary data Table S1.

<sup>a</sup>Found when using the 0.75 correlation coefficient cut-off.

Table 4. The GO terms, pathways and networks enriched for males among the significantly correlating genes for the various biomarkers for females

GO term / Pathway / Network	# significant processes (CC $\geq$ 0.60/ CC $\geq$ 0.75)	P-value (# genes affected/total # in pathway)	Database
DR CALUX <sup>®</sup>	18/0		
nucleosome assembly		1.22E-05 (4/69)	GO
Cell cycle_ Initiation of mitosis		6.37E-03 (2/18)	Pathways
ER $\alpha$ CALUX <sup>®</sup>	74/88		
immune system process		8.49E-06 (58/863)	GO
regulation of lymphocyte activation		1.99E-05 (18/155)	GO
AR CALUX <sup>®</sup>	39/33		
translational elongation		8.72E-19 (28/104)	GO
ubiquitin-dependent protein catabolic process		4.70E-06 (21/214)	GO
AA Hb-adducts	0/0		
GA Hb-adducts	0/0		
%MNBN	3/6		
nucleotide-excision repair, DNA damage removal		1.30E-04 (3/18)	GO
Cell cycle_ Initiation of mitosis		6.37E-03 (2/18)	Pathways

Only processes mentioned in the text are presented; for complete lists refer to supplementary data Table S2.

Table 5. GO terms and pathways enriched for males among all correlating genes using Tox-Profiler.

Biomarker	Process	# significant		T-value males	p-value males	T-value females	p-value females	Database
		processes	males/females					
DR CALUX <sup>®</sup>			5/29					
	nucleosome assembly			4.1	0.154	6.4	<0.001	GO
	T cell receptor signaling pathway			-2.7	1.000	-4.5	0.001	KEGG
	B cell receptor signaling pathway			-0.6	1.000	-4.1	0.005	KEGG
	TNF-alpha-NF-kB Signaling Pathway			2.9	0.551	-4.2	0.010	WikiPathways
ERα CALUX <sup>®</sup>			2/41					
	immune response			-1.4	1.000	6.8	<0.001	GO
	T Cell Receptor Signaling Pathway			-1.5	1.000	6.8	<0.001	WikiPathways
	B Cell Receptor Signaling Pathway			-1.9	1.000	4.9	<0.001	WikiPathways
AR CALUX <sup>®</sup>			12/24					
	immune response			-2.5	1.000	-6.6	<0.001	GO
	ubiquitin-dependent protein catabolic process			0.9	1.000	5.9	<0.001	GO
	nucleosome			-4.7	0.020	-1.2	1.000	GO
AA Hb-adducts			4/17					
	translational elongation			-1.7	1.000	8.8	<0.001	GO
	RNA splicing			-5.6	<0.001	1.7	1.0000	GO
GA Hb-adducts			8/12					
	Wnt signaling pathway			4.2	0.032	0.2	1.000	KEGG

%MNBN		30/13					
	translational elongation		8.5	<0.001	-5.7	<0.001	GO
	Spliceosome		4.6	0.002	-5.4	<0.001	KEGG
	mRNA processing		4.0	0.012	-4.1	0.002	WikiPathways
	Pathways in cancer		4.8	0.001	1.8	1.000	KEGG
	translational elongation		8.5	<0.001	-5.7	<0.001	GO

Only processes mentioned in the text are presented; for complete lists refer to supplementary data Table S3.





Fig 1

	GeneSymbol	Agilent Gene ID	CC Males	CC Females	Regulation of progression through cell cycle**	Proliferation/replication**	Apoptosis**	Regulation of transcription**	Cromatin/histone**	Immune response**
DR										
	JAK2	Janus kinase 2 a protein tyrosine kinase	0.77*	-0.18	Orange	Yellow	Blue			
	EWSR1	Ewing sarcoma breakpoint region 1	0.69*	-0.20				Purple		
	HSP90AB1	heat shock protein 90kDa alpha cytosolic, class B member 1	0.62*	-0.26						
	ZNF384	zinc finger protein 384	0.71*	-0.12				Purple		
Era										
	FUS	fusion involved in t 12;16 in malign liposarcoma	-0.66*	0.70*						
	CD79B	CD79b molecule, immunoglobulin-associated beta	-0.50	0.70*						Dark Red
	LCK	lymphocyte-specific protein tyrosine kinase	-0.25	0.80*	Orange		Blue			
	MLLT6	myeloid/lymphoid or mixed-lineage leukemia trithorax homolog, Drosophila; translocated to, 6	0.39	-0.63*				Purple		
AR										
	DEK	DEK oncogene DNA binding	-0.43	0.72*		Yellow		Purple	Grey	
	MLLT3	myeloid/lymphoid or mixed-lineage leukemia trithorax homolog, Drosophila; translocated to, 3	-0.50	0.65*				Purple		
	KRAS	v-Ki-ras2 Kirsten rat sarcoma viral oncogene homolog	-0.49	0.63*	Orange					
	IGH@	immunoglobulin heavy locus	0.62*	-0.41						
%dMNB										
	BCL2	B-cell CLL/lymphoma 2	-0.77*	0.49	Orange		Blue			Dark Red
	RUNX1	runt-related transcription factor 1 acute myeloid leukemia 1; aml1 oncogene	-0.80*	0.33				Purple		
	TPM3	tropomyosin 3	0.72*	-0.34						
	EIF4A2	eukaryotic translation initiation factor 4A, isoform 2	0.69*	-0.35						
AA Hb-adducts										
	NIN	ninein GSK3B interacting protein	-0.64*	0.12						
	PBX1	pre-B-cell leukemia homeobox 1	0.66*	-0.09		Yellow		Purple		
GA Hb-adducts										

# Cancer Epidemiology, Biomarkers & Prevention

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## Global gene expression analysis in cord blood reveals gender-specific differences in response to carcinogenic exposure *in utero*

Kevin Hochstenbach, Danitsja M. van Leeuwen, Hans Gmuender, et al.

*Cancer Epidemiol Biomarkers Prev* Published OnlineFirst August 9, 2012.

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