A Hot Spot for p53 Mutation in Transitional Cell Carcinoma of the Bladder: Clues to the Etiology of Bladder Cancer

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Abstract

Twenty-eight transitional cell carcinomas of the bladder, grade 2 or 3, were analyzed for the presence of p53 mutations. Thirteen tumors were found to contain 14 mutations. These were all base substitution mutations, of which nine were GC→AT transitions (three at CpG sites). The remaining five mutations were transversions (three GC→CG, one GC→TA, and one AT→TA). Four of the mutations were found at codon 280. A comparison with other studies of bladder tumors reveals that a region encompassing codons 280 and 285 represents a hot spot for p53 mutation in bladder cancer. The 280/285 hot spot lies within two purine-rich sequences that may provide some clues to the identity of potential bladder carcinogens. A comparison of mutations from bladder tumors of smokers and nonsmokers reveals no significant differences.

Introduction

Bladder cancer is the fifth most common neoplasm in the United Kingdom and the United States. The commonest form of bladder cancer is TCC. A number of genetic abnormalities have been found to be associated with TCC. Loss of markers on chromosome 9 (1) and chromosome 14 (2) occurs in approximately 50 and 30% of bladder tumors, respectively. In addition, p53 mutations have been identified with an incidence of around 50%. The remaining five sites (280/285 codons 280 and 285) represents a hot spot for p53 mutations. These were all transversions (three GC→CG, one GC→TA, and one AT→TA). Four of the mutations were found at codon 280. A comparison with other studies of bladder tumors reveals that a region encompassing codons 280 and 285 represents a hot spot for p53 mutation in bladder cancer. The 280/285 hot spot lies within two purine-rich sequences that may provide some clues to the identity of potential bladder carcinogens. A comparison of mutations from bladder tumors of smokers and nonsmokers reveals no significant differences.

Materials and Methods

Detection of p53 Mutations in Fresh Tumor Material. Grade 2 or 3 TCC samples were obtained by intraurethral resection from 28 patients over a 1-year period and frozen for subsequent DNA extraction using a Nucleon I DNA extraction kit (Scotlab). In one case, the DNA was extracted from urine sediment spun out of a 40-ml urine sample and washed four times with sucrose solution [0.25 M sucrose, 25 mM KCl, 1.8 mM CaCl2, and 50 mM Tris-HCl (pH 7.5)]. DNA sequences from exons 4→9 of the p53 gene were amplified using the PCR method. The oligonucleotide amplimers and amplification conditions were as described in Brash et al. (15), except that 30 cycles of amplification were performed. The downstream amplifier of each exon pair was biotinylated. The DNA amplification products were extracted from agarose gels by spinning the excised agarose blocks in Spin-X tubes (Costar). The DNA was then bound to streptavidin-coated magnetic Dynabeads (Dynal, Wirral, United Kingdom) and converted into single-strand DNA template. These templates were then directly sequenced using a T7 DNA polymerase sequencing kit (Pharmacia) with [33P]dATP. Sequence alterations were confirmed by repeating the PCR and sequencing reactions. The possibility of alterations being polymorphisms or germ-line mutations was discounted through sequencing of normal DNA samples. Standard precautions and controls were performed to exclude the possibility of cross-contamination.

Statistical Analysis. The p53 mutational spectra from smokers and nonsmokers were compared using a computer program for the analysis of mutational spectra data described in Cariello et al. (16). This program permits the comparison of independent...
A Hot Spot

An additional mutation was found in the urine sediment, arising from a urothelial tumor of unknown origin.

Statistical Analysis. The smoking status of each of the patients in the study was obtained through interview. The p53 mutations of patients classified as regular smokers or nonsmokers (never smoked) were added to those from previously published studies (Table 2). The mutational spectra from the smoking and nonsmoking groups were compared using the program described by Cariello et al. (16). Five separate analyses were carried out on the same set of data, and an average value of $P = 0.077 \pm 0.012$ was obtained for the probability that the two mutational spectra were derived from the same populations.

Discussion

Mutation Hot Spots in Bladder Tumors. The most striking feature of this relatively small group of mutations is the occurrence of four mutations at codon 280 (AGA), which included two GC→AT transitions (AGA→AAA) and two GC→CG transversions (AGA→ACA). This hot spot was first observed by Spruck et al. (5), but until now, it has not been confirmed in any other studies of bladder cancer. Fig. 1 shows the distribution of p53 mutations in TCC of the bladder from a number of published studies, including the mutations from this study. These data were extracted from the database of p53 mutations compiled by Hollstein et al. (17) and from three other studies (18–20). A total of 217 mutations distributed among 101 codons have been described for TCC.

Table 1. p53 mutations present in patients with TCC

<table>
<thead>
<tr>
<th>Patient</th>
<th>Stage/grade</th>
<th>Mutation Target</th>
<th>Codon</th>
<th>Amino acid change</th>
<th>Exon</th>
<th>LOH</th>
<th>Smoking status</th>
</tr>
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<tbody>
<tr>
<td>10</td>
<td>pT1/G2</td>
<td>A→T</td>
<td>Aga</td>
<td>Arg→Stop</td>
<td>4</td>
<td>LOH</td>
<td>S</td>
</tr>
<tr>
<td>533</td>
<td>pT3/G3</td>
<td>C→T</td>
<td>Cag</td>
<td>Gln→Stop</td>
<td>4</td>
<td>NI</td>
<td></td>
</tr>
<tr>
<td>382</td>
<td>pT1/G2</td>
<td>G→C</td>
<td>aaG</td>
<td>Lys→Asp</td>
<td>5</td>
<td>LOH</td>
<td>NS</td>
</tr>
<tr>
<td>420</td>
<td>pTx/G3</td>
<td>G→A</td>
<td>cGc</td>
<td>Arg→His</td>
<td>5</td>
<td>LOH</td>
<td>S</td>
</tr>
<tr>
<td>388</td>
<td>pT2/G2</td>
<td>G→T</td>
<td>Gaa</td>
<td>Glu→Stop</td>
<td>6</td>
<td>NI</td>
<td></td>
</tr>
<tr>
<td>439</td>
<td>pT1/G2</td>
<td>C→T</td>
<td>tCc</td>
<td>Ser→Pro</td>
<td>7</td>
<td>LOH</td>
<td>NS</td>
</tr>
<tr>
<td>316</td>
<td>pT2/G2</td>
<td>G→A</td>
<td>gGc</td>
<td>Gly→Asp</td>
<td>7</td>
<td>NI</td>
<td></td>
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<tr>
<td>394</td>
<td>pT1/G2</td>
<td>G→C</td>
<td>cGt</td>
<td>Arg→His</td>
<td>8</td>
<td>NI</td>
<td>Pipe</td>
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<tr>
<td>305</td>
<td>pT1/G2</td>
<td>G→A</td>
<td>agA</td>
<td>Arg→Lys</td>
<td>8</td>
<td>No LOH</td>
<td>NS</td>
</tr>
<tr>
<td>385</td>
<td>pT1/G2</td>
<td>G→A</td>
<td>aga</td>
<td>Arg→Lys</td>
<td>8</td>
<td>NI</td>
<td>NS</td>
</tr>
<tr>
<td>448</td>
<td>pT2/G3</td>
<td>G→C</td>
<td>aga</td>
<td>Arg→Thr</td>
<td>8</td>
<td>NI</td>
<td>exS</td>
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<tr>
<td>419</td>
<td>pTa/G3</td>
<td>C→T</td>
<td>Cgg</td>
<td>Arg→Trp</td>
<td>8</td>
<td>LOH</td>
<td>NS</td>
</tr>
<tr>
<td>429</td>
<td>pT2/G3</td>
<td>G→A</td>
<td>Gag</td>
<td>Glu→Lys</td>
<td>8</td>
<td>No LOH</td>
<td>exS</td>
</tr>
</tbody>
</table>

* Mutated base, upper case letter.

† LOH was assayed using informative polymorphisms at codons 72 and 213. NI, noninformative.

§ S. smoker; NS, nonsmoker; exS, ex-smoker; Pipe, pipe smoker; ?, smoking status unknown.

An additional mutation was found in the urine sediment, arising from a urothelial tumor of unknown origin.

Table 2. Comparison of p53 mutational spectra from TCCs of the bladder in smokers and nonsmokers

<table>
<thead>
<tr>
<th>Mutation</th>
<th>Nonsmoker (%)</th>
<th>Smoker (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>GC→AT*</td>
<td>16 (44)</td>
<td>10 (24)</td>
</tr>
<tr>
<td>CpG*</td>
<td>4 (11)</td>
<td>8 (19)</td>
</tr>
<tr>
<td>GC→TA</td>
<td>3 (8)</td>
<td>6 (14)</td>
</tr>
<tr>
<td>AT→GC</td>
<td>10 (28)</td>
<td>8 (19)</td>
</tr>
<tr>
<td>AT→TA</td>
<td>2 (6)</td>
<td>6 (14)</td>
</tr>
<tr>
<td>AT→CG</td>
<td>1 (3)</td>
<td>2 (5)</td>
</tr>
<tr>
<td>F/L/Del</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Total</td>
<td>36</td>
<td>42</td>
</tr>
</tbody>
</table>

* Data were compiled from this study and four previous studies (5, 6, 37, 38).

† GC→AT mutations at CpG sites are not included.

‡ GC→AT transitions at CpG sites.

§ F/L/Del, frameshift, insertion, and deletion mutations.

Discussion

Mutation Hot Spots in Bladder Tumors. The most striking feature of this relatively small group of mutations is the occurrence of four mutations at codon 280 (AGA), which included two GC→AT transitions (AGA→AAA) and two GC→CG transversions (AGA→ACA). This hot spot was first observed by Spruck et al. (5), but until now, it has not been confirmed in any other studies of bladder cancer. Fig. 1 shows the distribution of p53 mutations in TCC of the bladder from a number of published studies, including the mutations from this study. These data were extracted from the database of p53 mutations compiled by Hollstein et al. (17) and from three other studies (18–20). A total of 217 mutations distributed among 101 codons have been described for TCC. It can be seen that there are several mutational hot spots, with a particular concentration between codons 273 and 285. Twelve percent of the mutations occur at four CpG sites (codons 175, 248, 273, and 282).
Fig. 1. Distribution of p53 mutations in TCC of the bladder by codon position. The mutations are taken from the studies listed in the p53 mutation database compiled by Hollstein et al. (17) and from three other studies (18-20) as well as this one. The seven codons with the most mutations are labeled; those at CpG sites are boxed.

Fig. 2. Proportion of p53 mutations at codons 280 and 285 in various human tumors. The number of mutations found at codons 280 and 285 in various human tumors is expressed as a proportion of the total number of p53 mutations characterized in each tumor type. The data were obtained from the p53 mutation database compiled by Hollstein et al. (17) and from three other studies (18-20). The results of this study are also included.

whereas nearly 15% occur at three non-CpG hot spots, including the two most common sites of mutation (codons 280 and 285).

The significance of the codon 280 and 285 hot spots can be seen by comparing the frequency of mutation at these sites in a variety of human malignancies (Fig. 2). In this analysis, the numbers of mutations at codons 280 and 285 have been expressed as a percentage of the total number of characterized mutations for a variety of tumor types. Nearly 12% of bladder cancer mutations occur at these two hot spots. The next highest
frequency is less than 3%, which is seen in breast cancer and head and neck tumors. The codon 280/285 hot spot would therefore seem to be specific to TCC of the bladder.

Bladder-specific mutational hot spots may exist in the p53 gene for a variety of reasons. They may be a reflection of the interaction between bladder carcinogens and the p53 sequence; hot spots for mutation may represent sites of preferential DNA damage. Alternatively, there may be a tissue-specific biological selection that results in a bias toward mutations at particular sites: codon 280 and 285 mutations may confer a stronger selective growth advantage on bladder urothelial cells than other mutations. A third explanation is that bladder cells may lack some repair capability that repairs damage in the 280/285 region of the p53 gene more efficiently than that in other regions.

There are a number of documented links between preferential sites of damage and p53 mutation in human tumors. The distribution and type of mutations in skin tumors associated with sun exposure correlate well with the known sequence specificity of UV light mutagenesis (15). In addition, sites of frequent mutation in skin tumors seem to correlate with the sites of slowest repair of UV photoproducts (21). Other examples include sites of deamination at CpG sequences in colorectal cancer (codons 175, 245, 248, 273, and 282; reviewed in Ref. 22), aflatoxin in hepatocellular cancer (also reviewed in Ref. 22), and benzo(a)pyrene damage hot spots in lung tumors of smokers (23).

Different mutations of p53 are known to affect its transforming and suppressor functions to varying extents (24). Mutations in different domains can affect the biological properties of the resulting protein in different ways, depending on the cellular background (25). It is possible, therefore, that certain mutations have a stronger selective advantage in bladder urothelium than others. There is no evidence for this to date. In cell transformation studies, the p53 Thr280 mutation is dominant over wild type but has only moderate oncogenic activity (26). Overall, it would seem that the spectrum of p53 mutations seen in different tumors generally reflects the known mutational specificities of suspected etiological agents, e.g., UV light (15, 27) and tobacco smoke carcinogens (23, 28, 29). Nevertheless, the relative contribution of mutational specificity and biological selection of mutations to the final mutational spectrum in bladder cancer will need to be addressed by functional studies in appropriate cell lines using specific p53 mutations.

Epidemiological Clues from Bladder Hot Spots. All but 1 of 25 mutations occurring at the 280/285 hot spot in the p53 database involve mutations of GC bp. Of a total of 223 mutations in TCC of the bladder, 168 (75%) involve mutations of GC bp. In 108 of 168 (64%) cases, the guanine is on the nontranscribed strand. These observations are consistent with transcription-coupled repair of guanine adducts on the transcribed strand, resulting in a bias toward mutation at guanine adducts on the nontranscribed strand (30, 31). Guanines are favored sites of mutation by a wide range of mutagens such as nitrosoamines, 4-aminobiphenyl, polycyclic hydrocarbons, and oxygen radicals.

The two hot spots at codons 280 and 285 involve mutation of the first guanine within AGAG sequences occurring as part of extended homopurinic runs in the nontranscribed strand. GGGAGAG (codon 280, underlined) and AGAGGAG (codon 285, underlined). Six other AGAG sequences are present within the p53 coding sequence, and three of these sites lie within extended homopurinic runs at codons 14, 287, and 315. No base substitution mutations have been described at codons 14 or 315 of p53. However, three mutations involving GC→AT or CG substitutions of the first guanine in an AGAG sequence have been found at codon 287, and two of these were from bladder tumors. Thus, more than 12% of all p53 mutations in TCCs of the bladder are GC→AT or CG alterations occurring at the first guanine of an AGAG sequence within a homopurinic run. These observations could prove useful in discovering the identity of an important bladder carcinogen. For example, Mazur and Glickman (32) found that BPDE, a polycyclic hydrocarbon occurring in tobacco smoke, has a marked specificity for inducing mutations at AG(G)A sequences in Chinese hamster ovary cells. A study of BPDE mutagenesis in the supF gene in Escherichia coli found a similar hot spot for GC→CG mutations at the first guanine within an AGAG sequence (33). However, the same mutation hot spot was also induced by hydrogen peroxide (34).

Smoking versus Nonsmoking. Mutations at CpG sites are thought to arise by an endogenous deamination mechanism. Therefore, the relatively low proportion of mutations at CpG sites in TCC bladder tumors (15%), compared to approximately 50% of colon and germ-line mutations) is consistent with an important role for exogenous mutagenic agents in bladder carcinogenesis. Obvious candidates for such agents can be found in tobacco smoke, which is a risk factor in bladder cancer. An analysis of p53 mutations in human lung tumors shows a bias toward GC→TA transversions; 42 and 35% of mutations in small cell and non-small cell lung carcinoma, respectively, are of this type, compared to 16% in human tumors in general. A comparison of mutations from lung tumors of smokers and nonsmokers shows a significantly lower frequency of transitions and a significantly higher frequency of GC→TA transversions in smokers (22). The GC→TA transversion is induced in model systems by a number of known tobacco carcinogens such as benzo(a)pyrene and 4-aminobiphenyl. However, no particular bias toward GC→TA transversions (13% of total) is seen within bladder tumors (Fig. 3). A relatively high proportion of mutations are GC→CG transversions (18%), which are induced in model systems by aromatic amines such as 4-aminobiphenyl (35). Furthermore, DNA adducts derived from 4-aminobiphenyl have been detected in human exfoliated bladder cells (36).

Fig. 3. Mutational spectrum of p53 mutations in TCC of the bladder. The data are taken from the studies listed in the p53 mutation database compiled by Hollstein et al. (17), from three other studies (18–20), and from this study (n = 223).
Epidemiological information in the form of smoking history was available for most of the patients in this study (Table 1). It was possible, therefore, to combine our data with that from four other studies (5, 6, 37, 38) in which p53 mutations from smoking and nonsmoking bladder cancer patients were compared. Using a computer program designed by Cariello et al. (16), in which the distribution of mutations is taken into account as well as types of mutation, it would seem that the two spectra are not significantly different from each other (P = 0.077, average of five analyses). This confirms the result of an earlier analysis based on lower numbers (16).

The presence of no significant differences between the two spectra would suggest that similar mutagenic processes are involved in p53 mutagenesis within the bladder urothelium of smokers and nonsmokers. This conclusion is undermined somewhat by the presence in one study of double base substitutions in bladder tumors from four smokers (5). Double mutations of p53 occur often in other P animal-studies in which abnormally high levels of carcinogenic exposure are involved (29, 39). However, it is possible that p53 mutations in bladder cells is due to carcinogens in cigarette smoke to which nonsmokers are also exposed. The lack of any significant difference in the frequency of GC→TA transversions in smokers and nonsmokers suggests that different carcinogenic components of tobacco smoke may be involved in bladder cancerogenesis compared to lung and possibly rules out BPDE as a bladder carcinogen. However, there are a number of alternative carcinogenic agents present in tobacco smoke to which nonsmokers are also exposed, including nitrosamines, 4-aminobiphenyl, and free radicals. The codon 280/285 hot spot occurs in both smokers and nonsmokers, which supports the suggestion that it may be induced by some carcinogenic exposure common to both groups. The lack of any significant difference in mutational spectra between smokers and nonsmokers may suggest that tobacco-specific urothelial carcinogens are promoting agents rather than mutagenic agents.

Studies of p53 mutational spectra have clearly demonstrated the usefulness of such data in understanding the molecular epidemiology of certain types of cancer. Additional studies on bladder tumors, together with an analysis of mutagenesis at the codon 280 and 285 hot spots, may help to provide valuable clues to the identity of important bladder carcinogens.

References


A hot spot for p53 mutation in transitional cell carcinoma of the bladder: clues to the etiology of bladder cancer.

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