

GALAD Score for Hepatocellular Carcinoma Detection in Comparison with Liver Ultrasound and Proposal of GALADUS Score



Ju Dong Yang¹, Benyam D. Addissie¹, Kristin C. Mara², William S. Harmsen², Jianliang Dai³, Ning Zhang¹, Nicha Wongjarupong¹, Hawa M. Ali¹, Hamdi A. Ali¹, Fatima A. Hassan¹, Sravanthi Lavu¹, Jessica L. Cvinar¹, Nasra H. Giama¹, Catherine D. Moser¹, Katsuyuki Miyabe¹, Loretta K. Allotey¹, Alicia Algeciras-Schimmich⁴, J. Paul Theobald⁵, Melissa M. Ward⁵, Mindie H. Nguyen⁶, Alex S. Befeler⁷, K. Rajender Reddy⁸, Myron Schwartz⁹, Denise M. Harnois¹⁰, Hiroyuki Yamada¹¹, Sudhir Srivastava¹², Jo Ann Rinaudo¹², Gregory J. Gores¹, Ziding Feng³, Jorge A. Marrero¹³, and Lewis R. Roberts¹

Abstract

Background: The GALAD score is a serum biomarker-based model that predicts the probability of having hepatocellular carcinoma (HCC) in patients with chronic liver disease. We aimed to assess the performance of the GALAD score in comparison with liver ultrasound for detection of HCC.

Methods: A single-center cohort of 111 HCC patients and 180 controls with cirrhosis or chronic hepatitis B and a multicenter cohort of 233 early HCC and 412 cirrhosis patients from the Early Detection Research Network (EDRN) phase II HCC Study were analyzed.

Results: The area under the ROC curve (AUC) of the GALAD score for HCC detection was 0.95 [95% confidence interval (CI), 0.93–97], which was higher than the AUC of ultrasound (0.82, $P < 0.01$). At a cutoff of -0.76 , the GALAD score had a

sensitivity of 91% and a specificity of 85% for HCC detection. The AUC of the GALAD score for early-stage HCC detection remained high at 0.92 (95% CI, 0.88–0.96; cutoff -1.18 , sensitivity 92%, specificity 79%). The AUC of the GALAD score for HCC detection was 0.88 (95% CI, 0.85–0.91) in the EDRN cohort. The combination of GALAD and ultrasound (GALADUS score) further improved the performance of the GALAD score in the single-center cohort, achieving an AUC of 0.98 (95% CI, 0.96–0.99; cutoff -0.18 , sensitivity 95%, specificity 91%).

Conclusions: The performance of the GALAD score was superior to ultrasound for HCC detection. The GALADUS score further enhanced the performance of the GALAD score.

Impact: The GALAD score was validated in the United States.

¹Division of Gastroenterology and Hepatology, Mayo Clinic College of Medicine and Science, Rochester, Minnesota. ²Division of Biomedical Statistics and Informatics, Mayo Clinic College of Medicine and Science, Rochester, Minnesota. ³Department of Biostatistics, MD Anderson Cancer Center, Houston, Texas. ⁴Department of Laboratory Medicine and Pathology, Mayo Clinic College of Medicine and Science, Rochester, Minnesota. ⁵Clinical Immunoassay Laboratory, Mayo Clinic College of Medicine and Science, Rochester, Minnesota. ⁶Division of Gastroenterology and Hepatology, Stanford University Medical Center, Palo Alto, California. ⁷Division of Gastroenterology and Hepatology, Saint Louis University, St. Louis, Missouri. ⁸Division of Gastroenterology, Hospital of the University of Pennsylvania, Philadelphia, Pennsylvania. ⁹Department of General Surgery, The Mount Sinai Medical Center, New York, New York. ¹⁰Division of Gastroenterology and Hepatology, Mayo Clinic College of Medicine and Science, Jacksonville, Florida. ¹¹Wako Diagnostics, Mountain View, California. ¹²Division of Cancer Prevention, National Cancer Institute, Rockville, Maryland. ¹³Division of Digestive and Liver Disease, UT Southwestern Medical Center, Dallas, Texas.

Note: Supplementary data for this article are available at Cancer Epidemiology, Biomarkers & Prevention Online (<http://cebp.aacrjournals.org/>).

J.D. Yang and B.D. Addissie contributed equally to this article.

Corresponding Author: Lewis R. Roberts, Mayo Clinic College of Medicine and Science, 200 First Street SW, Rochester, MN 55905. Phone: 507-266-3239; Fax: 507-284-0762; E-mail: roberts.lewis@mayo.edu

doi: 10.1158/1055-9965.EPI-18-0281

©2018 American Association for Cancer Research.

Introduction

Hepatocellular carcinoma (HCC) has been rising in incidence and mortality rates in the United States. HCC is a leading cause of cancer-related death in many parts of the world, and liver cancer is the second global cause of cancer-related years of life lost after lung cancer (1). The increasing disease burden from HCC has been attributed to the rise in prevalence of chronic liver diseases that put patients at risk of developing HCC (2). Most HCCs occur in patients with cirrhotic liver disease, whose annual rate of liver cancer incidence is 1% to 5% (3). About a quarter of cirrhosis patients die from HCC as the tumors are frequently detected at an advanced symptomatic stage at which effective treatment options are limited (3–5). In order to achieve early detection of HCC in asymptomatic patients, HCC surveillance is recommended for high-risk patients in order to improve outcomes (6–8). A number of studies have shown that HCC surveillance is associated with earlier detection of HCCs and improved outcomes of patients with HCC (9–11).

Liver ultrasound is a standard HCC surveillance test, and biannual liver sonography has been the strategy recommended by the American Association for the Study of Liver Diseases (AASLD), the European Association for the Study of the Liver (EASL), and the Asian Pacific Association for the Study of the Liver

Yang et al.

(APASL; refs. 6–8, 12). However, the sensitivity of ultrasound for HCC detection has been reported to be 40% to 80% (9, 13). The sensitivity of ultrasound for HCC detection can be compromised by severity of liver disease, particular etiologies of liver disease (NASH or alcohol), and high body mass index (BMI; ref. 14). For this reason, up to 30% to 40% of tumors detected under surveillance are beyond early-stage HCC (9, 13, 15). Given the limited performance of ultrasound as a surveillance test, alpha-fetoprotein (AFP) is often used in addition to ultrasound, and a recent study from Taiwan showed that the use of AFP in addition to ultrasound significantly improves the sensitivity of surveillance compared with the ultrasound alone without significant loss of specificity (16). However, serum AFP level is affected by hepatic inflammation. Elevation of AFP in the absence of HCC can often be seen in the presence of enhanced hepatocyte destruction and regeneration, particularly in patients with chronic viral hepatitis. Pregnant woman or patients with gonadal tumors may have falsely elevated AFP, which limits its utility as a surveillance test (17, 18).

Recently, the GALAD score was proposed as a statistical model for estimating the likelihood of the presence of HCC in individual patients with chronic liver disease (19, 20). The GALAD score is derived from Gender, Age, AFP-L3, AFP, and Des-carboxy-prothrombin (DCP) and was shown to be a highly accurate model for the detection of HCC (19, 20). Prior studies have shown that biomarker performance varies with geographic region, being influenced by differences in the etiologic and patient factors in different regions. For example, AFP was shown to be a highly effective surveillance test in Alaska and Asia where Hepatitis B virus (HBV) is a major etiology. Performance of AFP was suboptimal in studies from Western countries where Hepatitis C virus (HCV) is the leading etiology (21, 22). The performance of the GALAD score has been validated in the UK, Germany, Japan, and Hong Kong, but has not as yet been evaluated in the United States. Further, the performance of the GALAD score in comparison with ultrasound remains to be determined.

The aim of this study was (1) to investigate the performance of the GALAD score for the detection of HCC in U.S. cohorts, (2) to compare the performance of the GALAD score with liver ultrasound, and (3) to develop a novel risk score model combining the GALAD score and liver ultrasound results.

Materials and Methods

Patients in the Mayo test cohort

The study was approved by the Mayo Clinic's Institutional Review Board. Individuals who gave research authorization and written-informed consent were included in the current study. For the test cohort, eligible patients were identified using the Mayo Clinic's institutional clinical database.

The control group consisted of patients who were candidates for HCC surveillance, namely those with cirrhosis or chronic hepatitis B without HCC seen at Mayo Clinic between October 2013 and October 2016, (1) who were tested for AFP, AFP-L3%, and DCP as part of their regular clinical care or (2) had provided stored serum with research consent authorization for the measurement of AFP, AFP-L3%, and DCP. Control patients were required to have at least 6 months of follow-up after GALAD score assessment to confirm the absence of HCC or have a negative contrast-enhanced multiphase CT, MRI, or liver biopsy at the time of GALAD score assessment.

The case group consisted of patients with newly diagnosed HCC in the setting of cirrhosis or chronic hepatitis B during the same study period, (1) who were tested for AFP, AFP-L3%, and DCP as part of their regular clinical care or (2) had provided stored serum with research consent authorization for the measurement of AFP, AFP-L3%, and DCP at the time of tumor diagnosis. Any patients who were on warfarin were excluded as warfarin can elevate the DCP level in the absence of HCC.

Clinical information

Demographic and clinical characteristics of patients were abstracted closest to the time of blood collection within a maximum time window of 3 months. Cirrhosis was defined by (1) histology or (2) findings characteristic of cirrhosis or portal hypertension in cross-sectional imaging studies (nodular configuration of the liver or signs of portal hypertension such as intraabdominal or esophageal varices, splenomegaly, dilated portal vein, or ascites) and/or thrombocytopenia (platelets <150 K; refs. 10, 23).

HBV infection as the underlying cause of the liver disease was confirmed based on positive HBsAg. HCV was confirmed by HCV RNA or anti-HCV with chronic liver disease. Alcohol was considered as a cause of HCC when a patient had a documented history of alcoholic liver disease or significant history of alcohol abuse or alcohol addiction (24). Nonalcoholic fatty liver disease (NAFLD) or nonalcoholic steatohepatitis (NASH) was diagnosed with radiologic or histologic evidence of fatty infiltration or inflammation in the setting of risk factors for fatty liver disease (metabolic syndrome) without any history of significant alcohol intake (<20 g per day) at HCC diagnosis or at any time before HCC diagnosis (25). When more than one etiology were identified in a patient (e.g., alcohol abuse and HCV), one was chosen using the best judgment of the clinician as the factor contributing most to the patient's liver dysfunction. The diagnosis of HCC was made by dynamic contrast CT or MRI of the liver or biopsy based on the guidelines of the AASLD (7).

Ultrasound results at the time of blood collection were abstracted from medical records in order to compare the performance of GALAD with ultrasound. A positive ultrasound result was defined by the presence of a solid hepatic lesion. Serum biomarkers were measured using the WAKO μ TASWako i30 Immunoanalyzer.

Multicenter Early Detection Research Network cohort

Data from the National Cancer Institute Early Detection Research Network (NCI EDRN) phase II biomarker case-control study for HCC were obtained for further multicenter evaluation of the performance of the GALAD score in the United States. New biomarkers are typically evaluated through five different phases of biomarker development (phase I, preclinical exploratory studies; phase II, clinical assay development and validation with case-control studies; phase III, retrospective longitudinal repository studies; phase IV, prospective screening studies; phase V, randomized cancer control studies; ref. 26). The aim of a phase II biomarker study is to develop a novel clinical assay and validate it in the setting of a case and control study design.

Briefly, the study included 233 consecutive early-stage HCC patients and 412 cirrhosis controls without HCC seen between February 2005 and August 2007 at 7 tertiary referral centers in the United States (27). The presence of cirrhosis was defined by histology or clinical evidence of portal hypertension in subjects with chronic liver disease. Subjects in the control group had an ultrasound, CT, or MRI showing no evidence of a hepatic mass

within 6 months prior to enrollment. Patients with an AFP ≥ 20 ng/mL at enrollment were also required to have a CT or MRI showing no mass suggestive of HCC within the 3 months prior to enrollment or up to 2 weeks after consent. All controls were assessed by AFP and imaging 6 months after enrollment to ensure that they did not have HCC. Serum AFP was measured by automated systems (Wako) at the time of enrollment prior to HCC-specific treatment.

Statistical analysis

The χ^2 test or Fisher exact test was used to compare categorical variables and the Student *t* or Wilcoxon rank-sum test for continuous variables. The GALAD score was calculated as $-10.08 + 1.67 \times [\text{Gender} (1 \text{ for male, } 0 \text{ for female})] + 0.09 \times [\text{Age}] + 0.04 \times [\text{AFP-L3}\%] + 2.34 \times \log[\text{AFP}] + 1.33 \times \log[\text{DCP}]$ (19). A web-based calculator is available at <http://www.mayoclinic.org/medical-professionals/model-end-stage-liver-disease/galad>. The area under the ROC curve (AUC) was calculated to assess the performance of the GALAD score for the detection of HCC. Subgroup analyses were performed to investigate the performance of the GALAD score in subgroups of patients with different gender, age, AFP, etiologies, BMI, cirrhosis status, hepatic dysfunction, and tumor stage.

Youden's index was used to find the optimal GALAD cutoffs from which sensitivity, specificity, and their corresponding 95% confidence intervals (CI) were calculated and compared with that of liver ultrasound (28). The GALADUS score was developed combining GALAD and liver ultrasound for the detection of HCC using multivariable logistic regression analysis containing all variables in the GALAD score and liver ultrasound.

As the GALAD and GALADUS are constructed based on tests performed on the same individuals, statistical analysis on differences between curves should take into account the correlated nature of the data. Therefore, the comparison of the AUCs was performed using Delong test, which is a nonparametric statistical test comparing the areas under two or more correlated ROCs (29).

Statistical analysis was carried out using SAS 9.4 (SAS Institute) and R version 3.0.2 (R Foundation).

Results

Clinical characteristics of Mayo test cohort

A total of 291 patients (111 cases and 180 controls) were eligible and analyzed. Demographic and clinical characteristics of the patients were summarized in Table 1. The proportion of males was higher among HCC patients (78%) than patients in the control group (53%; $P < 0.01$). HCC patients were older than patients in the control group (64 vs. 57, $P < 0.01$). HCV was the leading etiology of HCC (43%), whereas NASH was the most common etiology of liver disease in the control group (27%; $P < 0.01$). A small number of patients did not have liver cirrhosis (2 HCCs and 26 controls) in the setting of HBV etiology.

As expected, the mean GALAD score was higher in HCC patients than in control patients (3.8 vs. -2.6 , $P < 0.01$). More than half of the HCC patients had very early ($n = 16$, 14%) or early-stage ($n = 44$, 40%) HCC at diagnosis based on the Barcelona Clinic Liver Cancer (BCLC) staging classification.

Performance of GALAD for the detection of HCC in comparison with liver ultrasound

The AUC of the GALAD score for HCC detection was 0.95 (95% CI, 0.93–0.97; Table 2; Fig. 1A). At an optimal cutoff of -0.76 , the

Table 1. Clinical characteristics of Mayo Clinic cohort

	HCC (N = 111)	Control (N = 180)	P value
Age, mean (SD)	63.9 (8.6)	57.1 (11.6)	<0.01
Age <60	32 (28.8%)	91 (50.6%)	
Age ≥ 60	79 (71.2%)	89 (49.4%)	
Gender			<0.01
Female	25 (22.5%)	84 (46.7%)	
Male	86 (77.5%)	96 (53.3%)	
Race			0.39
White	86 (77.5%)	148 (82.2%)	
Asian	8 (7.2%)	14 (7.8%)	
Non-White/Asian	17 (15.3%)	18 (10.0%)	
Child-Pugh			0.56
Missing	4	19	
A	78 (72.9%)	112 (69.6%)	
B-C	29 (27.1%)	49 (30.4%)	
Etiology			<0.01
HCV	48 (43.2%)	33 (18.3%)	
HBV	11 (9.9%)	27 (15.0%)	
Alcohol	14 (12.6%)	37 (20.6%)	
NASH	30 (27.0%)	49 (27.2%)	
Others	8 (7.2%)	34 (18.9%)	
Cirrhosis	109 (98.2%)	154 (85.6%)	<0.01
BMI			0.72
≤ 30	58 (52.3%)	96 (55.8%)	
30–35	26 (23.4%)	41 (23.8%)	
>35	27 (24.3%)	35 (20.3%)	
Platelet count, median (IQR)	99.0 (70.0–195.0)	130.0 (93.0–180.0)	0.023
ALT, median (IQR)	51.0 (36.5–85.0)	32.0 (22.0–49.0)	<0.01
ALT <40	37 (33.3%)	119 (67.2%)	
ALT ≥ 40	74 (66.7%)	58 (32.8%)	
FIB-4, median (IQR)	7.0 (4.2–10.0)	3.5 (1.9–6.1)	<0.01
AFP, median (IQR)	39.8 (7.4–462.0)	3.3 (2.2–5.5)	<0.01
AFP <20	47 (42.3%)	176 (97.8%)	
AFP ≥ 20	64 (57.7%)	4 (2.2%)	
DCP, median (IQR)	9.5(2.2–151.0)	0.6 (0.3–2.0)	<0.01
L3, median (IQR)	12.8 (6.9–32.3)	1.0 (1.0–9.2)	<0.01
Ascites			0.77
0 (no ascites)	76 (68.5%)	117 (65.4%)	
1 (controlled ascites)	28 (25.2%)	47 (26.3%)	
2 (refractory ascites)	7 (6.3%)	15 (8.4%)	
GALAD_Z, mean (SD)	3.8 (4.4)	-2.6 (1.9)	<0.01
Ultrasound lesion			<0.01
Missing	21	7	
Negative	10 (11.1%)	136 (78.6%)	
Positive	80 (88.9%)	37 (21.4%)	
Tumor stage			NA
Very early (BCLC 0)	16 (14.5%)		
Early (BCLC A)	44 (39.6%)		
Nonearly (BCLC B-D)	51 (45.9%)		

GALAD score had a sensitivity of 91% and a specificity of 85% for HCC detection. The AUC of the GALAD score was higher than that of ultrasound for HCC detection (0.95 vs. 0.82, $P < 0.01$).

The AUC for GALAD remained high in subgroups of patients with different gender, age, AFP level, and HCC etiologies (Table 2). For instance, the GALAD score had a sensitivity of 89% and a specificity of 81% for AFP-negative tumor detection at a cutoff of -1.18 , and the ROC of the GALAD score was higher than that of ultrasound (0.90 vs. 0.80, $P < 0.01$). On the other hand, the AUC of ultrasound for HCC detection was low at 0.72 in patients with alcohol etiology. Ascites and high Child-Turcotte-Pugh (CTP) class compromised the performance of ultrasound, but did not affect the performance of the GALAD score (Table 2).

When the analysis was limited to early-stage HCC (BCLC 0-A), the AUC for the GALAD score remained high at 0.92 (95% CI,

Yang et al.

Table 2. Performance of GALAD within subgroups of Mayo Clinic cohort

Subgroups	GALAD cutoff	Sensitivity (95% CI)		Specificity (95% CI)		AUC (95% CI) ^a		P ^b
		GALAD	Ultrasound	GALAD	Ultrasound	GALAD	Ultrasound	
Overall	-0.76	0.91 (0.84-0.96)	0.89 (0.81-0.95)	0.85 (0.79-0.90)	0.79 (0.72-0.84)	0.95 (0.93-0.97)	0.82 (0.77-0.87)	<0.01
Gender								
Male	0.27	0.84 (0.74-0.91)	0.87 (0.77-0.94)	0.92 (0.84-0.96)	0.80 (0.71-0.88)	0.95 (0.92-0.97)	0.83 (0.77-0.89)	<0.001
Female	-1.04	0.88 (0.69-0.97)	0.95 (0.76-1.00)	0.95 (0.88-0.99)	0.77 (0.66-0.85)	0.94 (0.88-1.00)	0.80 (0.71-0.88)	0.001
Age								
Age <60	-1.41	0.97 (0.84-1.00)	0.89 (0.71-0.98)	0.89 (0.81-0.95)	0.78 (0.67-0.86)	0.98 (0.96-1.00)	0.80 (0.72-0.87)	<0.001
Age ≥60	0.34	0.82 (0.72-0.90)	0.89 (0.78-0.95)	0.91 (0.83-0.96)	0.80 (0.70-0.88)	0.93 (0.89-0.97)	0.84 (0.77-0.89)	0.006
AFP								
AFP <20	-1.18	0.89 (0.77-0.96)	0.87 (0.72-0.96)	0.81 (0.74-0.86)	0.79 (0.72-0.85)	0.90 (0.86-0.95)	0.80 (0.74-0.85)	<0.01
AFP ≥20	3.91	0.69 (0.56-0.80)	0.90 (0.79-0.97)	1.00 (0.28-1.00)	0.75 (0.19-0.99)	0.87 (0.72-1.00)	0.89 (0.78-0.96)	0.37
Etiology								
HCV	-0.53	0.94 (0.83-0.99)	0.92 (0.80-0.98)	0.85 (0.68-0.95)	0.74 (0.55-0.88)	0.95 (0.90-1.00)	0.85 (0.74-0.92)	0.02
HBV	-1.68	1.00 (0.62-1.00)	1.00 (0.47-1.00)	1.00 (0.82-1.00)	0.85 (0.66-0.96)	1.00 (1.00-1.00)	0.88 (0.73-0.97)	0.01
Alcohol	-1.18	1.00 (0.68-1.00)	0.77 (0.46-0.95)	0.73 (0.56-0.86)	0.70 (0.53-0.84)	0.93 (0.86-0.99)	0.72 (0.58-0.84)	<0.01
NASH	-0.85	0.87 (0.69-0.96)	0.85 (0.66-0.96)	0.84 (0.70-0.93)	0.83 (0.70-0.93)	0.89 (0.81-0.97)	0.84 (0.74-0.91)	0.18
Others	0.27	1.00 (0.52-1.00)	1.00 (0.19-1.00)	1.00 (0.85-1.00)	0.80 (0.61-0.92)	1.00 (1.00-1.00)	0.82 (0.65-0.93)	<0.01
BMI								
≤30	0.34	0.867 (0.75-0.94)	0.91 (0.79-0.98)	0.95 (0.88-0.98)	0.75 (0.65-0.83)	0.97 (0.95-0.99)	0.80 (0.72-0.86)	<0.01
30-35	0.11	0.88 (0.70-0.98)	0.89 (0.67-0.99)	0.95 (0.83-0.99)	0.75 (0.59-0.87)	0.97 (0.93-1.00)	0.80 (0.67-0.89)	<0.01
>35	-1.42	0.89 (0.71-0.98)	0.85 (0.65-0.96)	0.80 (0.63-0.92)	0.88 (0.73-0.97)	0.87 (0.78-0.97)	0.87 (0.75-0.94)	0.50
Ascites								
0 (No ascites)	-0.18	0.95 (0.87-0.99)	0.91 (0.81-0.96)	0.91 (0.84-0.96)	0.78 (0.69-0.85)	0.97 (0.95-0.99)	0.83 (0.76-0.88)	<0.01
1 (Controlled ascites)	-0.27	0.95 (0.75-0.97)	0.85 (0.62-0.97)	0.85 (0.72-0.94)	0.81 (0.67-0.91)	0.96 (0.92-1.00)	0.82 (0.71-0.90)	<0.01
2 (Refractory ascites)	-0.20	1.00 (0.42-1.00)	0.83 (0.36-1.00)	1.00 (0.70-1.00)	0.73 (0.45-0.92)	1.00 (1.00-1.00)	0.76 (0.53-0.92)	0.01
Child-Pugh								
A	-0.85	0.92 (0.84-0.97)	0.94 (0.85-0.98)	0.84 (0.76-0.90)	0.77 (0.67-0.84)	0.94 (0.91-0.98)	0.83 (0.76-0.88)	<0.01
B-C	0.42	0.83 (0.64-0.94)	0.74 (0.52-0.90)	0.96 (0.86-1.00)	0.77 (0.62-0.88)	0.95 (0.91-0.99)	0.76 (0.64-0.85)	<0.01
ALT								
<40	-1.01	0.95 (0.82-0.99)	0.84 (0.66-0.95)	0.86 (0.78-0.91)	0.84 (0.76-0.90)	0.95 (0.92-0.99)	0.84 (0.77-0.89)	<0.01
≥40	0.38	0.84 (0.73-0.91)	0.92 (0.81-0.97)	0.95 (0.86-0.99)	0.67 (0.53-0.79)	0.94 (0.91-0.98)	0.80 (0.71-0.87)	<0.01
Tumor stage								
Very early (BCLC 0)	-1.70	1.00 (0.70-1.00)	0.67 (0.35-0.90)	0.72 (0.64-0.78)	0.79 (0.72-0.84)	0.91 (0.85-0.96)	0.78 (0.71-0.84)	<0.01
Early (BCLC 0-A)	-1.18	0.92 (0.82-0.97)	0.92 (0.81-0.98)	0.79 (0.73-0.85)	0.79 (0.72-0.84)	0.92 (0.88-0.96)	0.82 (0.76-0.87)	<0.01
Nonearly (BCLC B-D)	0.27	0.92 (0.81-0.98)	0.85 (0.69-0.94)	0.94 (0.90-0.97)	0.79 (0.72-0.85)	0.99 (0.97-1.00)	0.80 (0.74-0.85)	<0.01

^aFor the calculation of AUC, the continuous GALAD score is used, whereas for sensitivity and specificity, we used the GALAD cutoff.^bP value looking at difference in AUC between GALAD and ultrasound.

0.88-0.96; best cutoff -1.18, sensitivity 92%, specificity 79%; Table 2; Fig. 1B). The AUC of the GALAD score was higher than that of ultrasound for detection of early-stage (0.92 vs. 0.82, $P < 0.01$) and very early-stage HCC (0.91 vs. 0.78, $P < 0.001$). Overall results remain excellent when the analysis was limited to cirrhotic cases and controls only (109 HCC; 154 controls) or after frequency matching of the Child-Pugh Score between cases and controls (105 HCC; 105 controls; Supplementary Tables S1 and S2).

When the GALAD score cutoff was set at -0.63, the best cutoff of GALAD score reported in the previous study, the performance of GALAD score remains excellent (Supplementary Table S3). The GALAD score had a sensitivity of 89% and a specificity of 86% for HCC detection with a sensitivity of 82% and a specificity of 86% for early-stage and a sensitivity of 73% and a specificity of 86% for very early-stage tumor detection.

Performance of GALAD for the detection of early-stage HCC in the multicenter EDRN cohort

A total of 412 patients with cirrhosis without HCC and 233 patients with early-stage HCC, 10 of whom had very early-stage tumor, included in the EDRN cohort were analyzed. The clinical characteristics of the patients are summarized in Supplementary Table S4. The AUC of the GALAD score for HCC detection was

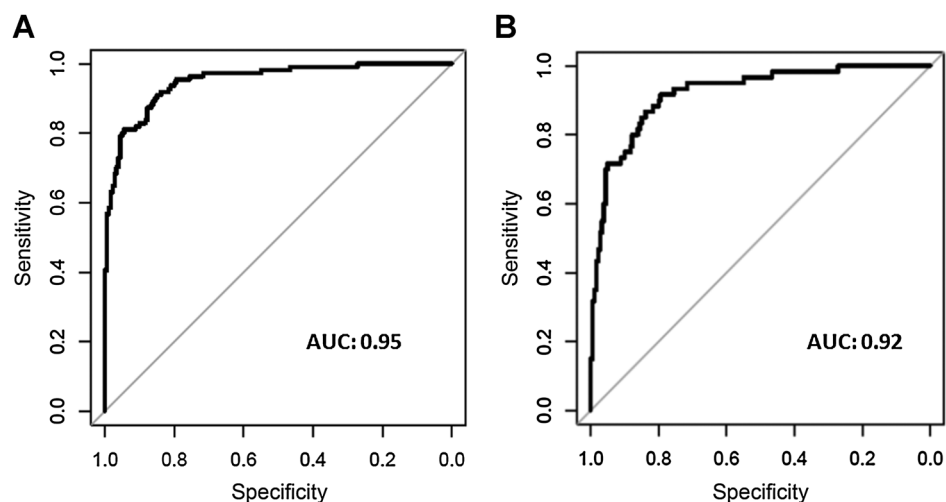
0.88 (95% CI, 0.85-91; Table 3). At a cutoff of -0.17, the GALAD score had a sensitivity of 76% and a specificity of 86% for HCC detection. The AUC of the GALAD score for detection of very early stage was 0.86 (95% CI, 0.76-96; Table 3). The AUC of GALAD remained excellent in subgroups of patients with different HCC etiologies (Table 3; Fig. 2). When the GALAD score cutoff was set at -0.63, the GALAD score had a sensitivity of 79% and a specificity of 79% for HCC detection (Table 3).

Proposal of GALADUS score in the Mayo test cohort

Next, we evaluated whether the combination of liver ultrasound and GALAD score further enhances the performance of HCC detection compared with the performance of either test alone. Combining GALAD and liver ultrasound significantly improved the performance of the model (Supplementary Table S5). The new model was named GALADUS score, and the equation is as follows: $\text{GALADUS} = -12.79 + 0.09 \times \text{age} + 1.74 \times (1 \text{ for male, } 0 \text{ for female}) + 2.44 \times \log_{10}(\text{AFP}) + 0.04 \times \text{AFP-L3} + 1.39 \times \log_{10}(\text{DCP}) + 3.56 \times (1 \text{ for positive ultrasound, } 0 \text{ for negative ultrasound})$. The AUC of the GALADUS score for HCC detection was 0.98 (95% CI, 0.96-0.99; Fig. 3A). At the GALADUS cutoff of -0.179, the sensitivity was 95% with a specificity of 0.91%. When the analysis was limited to early-stage HCC (BCLC 0-A), the AUC for the GALADUS score remained high at 0.97

Figure 1.

The ROC of GALAD scores for HCC diagnosis in each subgroup (Mayo cohort). X axis: specificity; Y axis: sensitivity. **A**, The ROC of GALAD score for detection of HCC. **B**, The ROC of GALAD score for detection of early-stage HCC.



(95% CI, 0.95–0.99; best cutoff -0.5 , sensitivity 88%, specificity 94%; Fig. 3B).

Discussion

In the current study, the excellent performance of the GALAD score for the detection of HCC was confirmed for the first time in a U.S. cohort. First, the performance of the GALAD score was shown to be superior to that of ultrasound. The performance of the GALAD score remained excellent for the detection of early-stage HCC including tumors with negative AFP. Gender, age group, underlying etiology of HCC, and severity of liver dysfunction did not affect the performance of the GALAD score, whereas the performance of ultrasound was hampered in patients with poorly controlled ascites or Child–Pugh class of B or C cirrhosis. The outstanding performance of the GALAD score was confirmed in an independent multicenter U.S. cohort of early-stage HCC and cirrhosis patients, confirming the utility of the GALAD score as an excellent tool for HCC detection. Finally, we proposed the

GALADUS score, which combines the GALAD score and liver ultrasound for the detection of HCC, and showed improved performance over liver ultrasound or GALAD score alone, although the minimal improvement of the AUC of GALADUS over GALAD may not be clinically relevant.

The GALAD score was first developed from a single UK center using a statistical model that could determine the risk of HCC in individual patients with chronic liver diseases using objective measures, particularly serological tumor markers (19). The AUC of the GALAD score for detection of all HCCs was 0.97, with an AUC of 0.96 for detection of early-stage HCC and an AUC of 0.98 for detection of advanced-stage HCC (19). In a German cohort, the GALAD score achieved a sensitivity of 92% and a specificity of 90% at the cutoff of -0.63 . The GALAD score was subsequently validated in a larger multicenter, multicontinent study (20). This study included 2,430 patients with HCC and 4,404 with chronic liver disease recruited from Germany, Japan, and Hong Kong. Similar to the original study, the overall AUCs of GALAD for HCC detection were 0.93 (95% CI, 0.92–0.94) and 0.94 (95% CI,

Table 3. Cutoffs for GALAD score at the maximum sensitivity and specificity in the ROC curve in the EDNRN multicenter cohort

Subgroups	GALAD cutoff	Sensitivity (95% CI)	Specificity (95% CI)	GALAD cutoff	Sensitivity (95% CI)	Specificity (95% CI)	AUC (95% CI)
Overall (233 HCCs, 412 controls)	-0.17	76% (70%–82%)	86% (82%–89%)	-0.63	79% (74%–84%)	79% (75%–83%)	0.88 (0.85–0.91)
Etiology subgroup							
Alcohol (24 HCCs, 48 controls)	0.82	71% (49%–87%)	92% (80%–98%)	-0.63	83% (67%–96%)	67% (54%–79%)	0.89 (0.81–0.97)
HBV (46 HCCs, 23 controls)	-1.25	83% (69%–92%)	100% (85%–100%)	-0.63	72% (59%–85%)	100% (100%–100%)	0.94 (0.88–0.99)
HCV (122 HCCs, 240 controls)	-0.17	79% (70%–85%)	85% (80%–89%)	-0.63	82% (75%–88%)	77% (72%–82%)	0.89 (0.86–0.92)
Nonviral/alcohol ^a (41 HCCs, 101 controls)	-0.16	78% (62%–89%)	93% (86%–97%)	-0.63	78% (63%–90%)	85% (78%–92%)	0.88 (0.81–0.96)
AFP							
AFP <20 (95 HCCs, 360 controls)	-1.27	77% (67%–85%)	73% (68%–78%)	-0.63	55% (44%–64%)	85% (81%–88%)	0.80 (0.75–0.85)
AFP ≥ 20 (137 HCCs, 48 controls)	2.39	72% (63%–79%)	90% (77%–97%)	-0.63	96% (93%–99%)	38% (25%–52%)	0.87 (0.81–0.93)
Tumor stage							
Very early (BCLC 0; 10 HCCs, 412 controls)	-0.16	80% (44%–97%)	86% (83%–89%)	-0.63	80% (50%–100%)	79% (75%–83%)	0.86 (0.76–0.96)

^aNonviral/alcohol refers to all other etiologies combined other than alcohol, HBV, and HCV.

Yang et al.

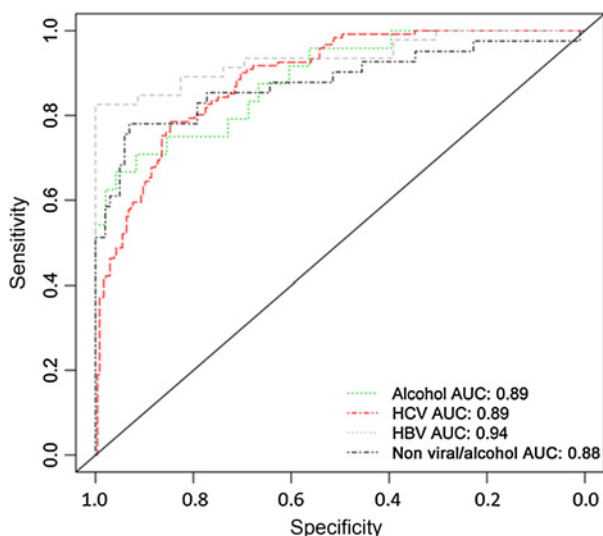


Figure 2. The ROC of GALAD scores for HCC diagnosis in each subgroup of patients with different etiologies (EDRN cohort). X axis: specificity; Y axis: sensitivity.

0.93–0.96) for the Japanese and German validation cohorts, respectively. Similar to our study results, the GALAD score provided a sensitivity of 88% and specificity of 88% at the cutoff of -0.68 in the German cohort (20). Another study of 98 patients from Italy (44 chronic liver disease patients and 54 HCC patients) reported that the overall AUC of GALAD for HCC detection was 0.98 (30).

Liver ultrasound is a standard HCC surveillance test endorsed by several societies including AASLD, EASL, and APASL (6, 7, 12). However, there are several major limitations of ultrasound as a surveillance test for HCC (9). The performance of ultrasound in surveillance is highly dependent on the skill of the sonographer.

In addition, detecting an early HCC nodule can be challenging, particularly in a patient with a nodular cirrhotic liver.

A recent single-center study of 941 patients with cirrhosis showed that 20% of ultrasounds in those patients were inadequate for excluding HCC (14). Similar to our current study, decreased performance of ultrasound for detection of HCC was independently demonstrated in patients with alcohol-induced liver cirrhosis (OR, 2.11; 95% CI, 1.33–3.37) or patients with advanced stages of liver disease (Child–Pugh class B or C cirrhosis; OR, 1.93; 95% CI, 1.32–2.81). The GALAD score may therefore serve as an invaluable surveillance test in these subgroups as its performance did not significantly decrease with etiology or severity of liver disease. Furthermore, use of the GALAD score may decrease the risk of surveillance-related physical harms by identifying patients with false-positive ultrasound results and may maximize the potential benefit by identifying patients with false-negative ultrasound results (15).

From the public or global health standpoint, the GALAD score can be used in the parts of the world where medical resources are limited and liver ultrasound is not widely available or easily affordable. The GALAD score is easily calculated and may serve as an excellent screening test, thus increasing the uptake and compliance with surveillance and consequently improving the effectiveness of the surveillance program.

Our current study has several limitations. Given the retrospective design of this phase II biomarker study, the results could have been affected by unmeasured potential biases. As the study was performed on subjects seen at major referral centers, the results could be susceptible to referral bias. In order to minimize the concerns related to single-center study designs, we also analyzed data from the NCI EDRN phase II HCC biomarker and confirmed the excellent performance of the GALAD score for HCC detection in this multicenter U.S. cohort. A recent interim report of a randomized controlled trial (ultrasound + biomarker vs. ultrasound alone) showed that addition of three biomarkers (AFP, AFP-L3%, and DCP) increased the sensitivity of HCC surveillance in comparison with ultrasound alone, although number of new

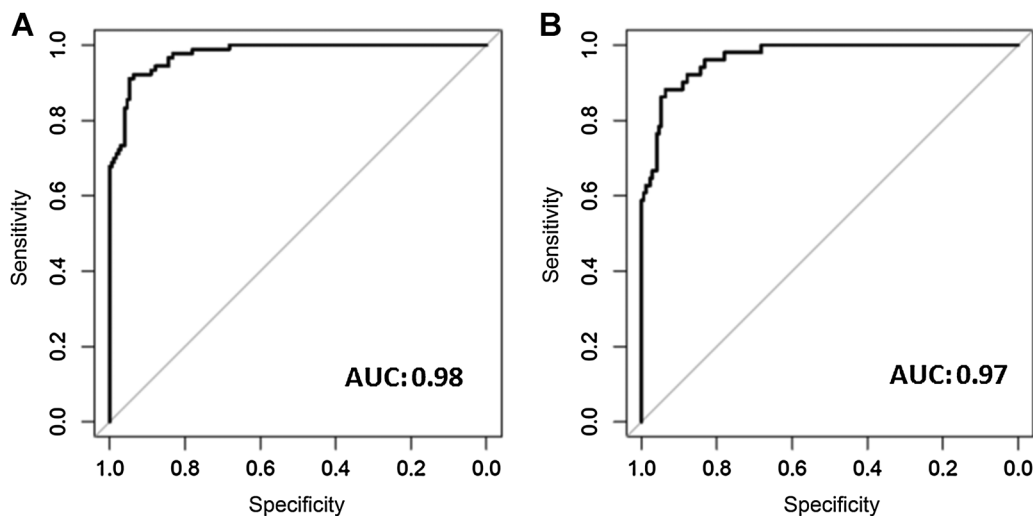


Figure 3. The ROC of GALADUS scores for HCC diagnosis. X axis: specificity; Y axis: sensitivity. **A**, The ROC of GALADUS score for detection of HCC. **B**, The ROC of GALADUS score for detection of early-stage HCC.

HCC cases was small ($n = 20$; ref. 31). Similarly, an interim analysis of a small phase III biomarker study showed that a GALAD score >1.95 had a sensitivity of 90% 6 months prior to HCC diagnosis. Similar to the previous study, the small number of incident cases ($n = 14$) is a major limitation (32). Both studies have been reported in abstract form. A larger phase III prospective multicenter Early Detection Research Network–HCC Early Detection Strategy (HEDS) study is currently underway to identify novel biomarkers for HCC. We plan to further validate the performance of GALAD and GALADUS scores in comparison with ultrasound in this large prospective cohort of cirrhotic patients in which the optimum cutoffs of the GALAD and GALADUS scores will be further defined. We expect that the optimum cutoff of the GALAD/GALADUS score will vary depending on the baseline characteristics of the at-risk population as it does for AFP. The small sample size in the current study prohibited robust analysis to propose a specific cutoff for each subgroup. Nonetheless, our data clearly showed that the performance of the GALAD score remains excellent at a cut off of -0.63 , the best cutoff for the GALAD score reported in the previous study. Therefore, this would be a reasonable cutoff to use until more robust data become available.

In conclusion, the excellent performance of the GALAD score was shown for the first time in the United States. In addition, in this first study comparing the performance of the GALAD score with ultrasound, we showed that the GALAD score has superior performance to ultrasound for HCC detection. The GALAD score is complementary to ultrasound for the detection of HCC, and it will be particularly important in group of patients with advanced-stage hepatic dysfunction or obesity who are at risk for false-negative ultrasound. Finally, we proposed the GALADUS score incorporating the GALAD score and liver ultrasound result, which was shown to be superior to either ultrasound or GALAD score alone. Although the benefit of GALADUS over GALAD was minimal in this case–control study, any change in clinical practice would require ultrasound + GALAD versus ultrasound alone in a prospective setting, and judgment as to the relative values of the two approaches (individually) or as a combination (as in GALADUS) should await the outcome of such study. Further evaluation of the GALAD and GALADUS scores will be performed in a larger phase III prospective multicenter cohort biomarker study, where their utility for detection of early HCC will be tested in the setting of HCC surveillance. Future studies should investigate the cost-effectiveness of GALAD or GALADUS over liver ultrasound with/without AFP before its wide implementation in clinical practice.

Disclosure of Potential Conflicts of Interest

M.H. Nguyen reports receiving commercial research support from Bristol-Myers Squibb, Janssen Pharmaceuticals, Gilead Sciences, Laboratory for Advanced Medicine, and Exact Science; received honoraria from the speakers'

bureau of Bristol-Myers Squibb, Janssen Pharmaceuticals, Intercept Pharmaceuticals, Roche Laboratories, Dynavax Laboratory, and Alnylam Pharmaceuticals; and is a consultant/advisory board member for Bristol-Myers Squibb, Janssen Pharmaceuticals, Exact Science, Bayer, Gilead Sciences, Intercept Pharmaceuticals, Roche Laboratories, Dynavax Laboratory, Alnylam Pharmaceuticals, Novartis, Laboratory for Advanced Medicine, and Eisai Science. L.R. Roberts reports receiving commercial research funding from Ariad, Wako, Gilead, BTG, and Bayer, and Redhill; received honoraria from the speakers' bureau of Wako, Medscape, NACCME, and OncoLive; and is a consultant/advisory board member for Bayer, Exact Sciences, Tavec, and Grail. No other potential conflicts of interest were disclosed by the other authors.

Disclaimer

The content of this article is solely the responsibility of the authors and does not necessarily represent the official views of the NIH.

Authors' Contributions

Conception and design: J.D. Yang, B.D. Addissie, B.D. M. Schwartz, D.M. Harnois, S. Srivastava, J.A. Rinaudo, Z. Feng, J.A. Marrero, L.R. Roberts
Development of methodology: J.D. Yang, B.D. Addissie, W.S. Harmsen, J.A. Rinaudo, J.A. Marrero, L.R. Roberts
Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): J.D. Yang, B.D. Addissie, N. Zhang, N. Wongjarupong, H.M. Ali, H.A. Ali, F.A. Hassan, S. Lavu, J.L. Cvinar, N.H. Giama, C.D. Moser, K. Miyabe, L.K. Allotey, A. Algeciras-Schimnich, J.P. Theobald, A.S. Befeler, K.R. Reddy, M. Schwartz, H. Yamada, G.J. Gores, L.R. Roberts
Analysis and interpretation of data (e.g., statistical analysis, bio-statistics, computational analysis): J.D. Yang, B.D. Addissie, K.C. Mara, W.S. Harmsen, J. Dai, N.H. Giama, K.R. Reddy, G.J. Gores, Z. Feng, J.A. Marrero, L.R. Roberts
Writing, review, and/or revision of the manuscript: J.D. Yang, B.D. Addissie, K.C. Mara, W.S. Harmsen, J. Dai, H.A. Ali, A. Algeciras-Schimnich, A.S. Befeler, K.R. Reddy, M. Schwartz, D.M. Harnois, S. Srivastava, Z. Feng, J.A. Marrero, L.R. Roberts
Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases): J.D. Yang, B.D. Addissie, J.L. Cvinar, N.H. Giama, Z. Feng, L.R. Roberts
Study supervision: M.M. Ward, M.H. Nguyen, S. Srivastava, J.A. Rinaudo, G.J. Gores, Z. Feng, L.R. Roberts

Acknowledgments

This publication was supported by grant number T32 DK07198 from the National Institute of Diabetes and Digestive and Kidney Diseases (NIDDK; to J.D. Yang) and CA165076 from the NCI, the Mayo Clinic Center for Cell Signaling in Gastroenterology (NIDDK P30DK084567), the Mayo Clinic Cancer Center (CA15083), the Mayo Clinic Center for Translational Science Activities (NIH/NCRR CTSa grant number UL1 TR000135, to L.R. Roberts), and the Early Detection Research Network (NCI U24CA086368). Biomarker assays were performed by Wako Diagnostics.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked *advertisement* in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received March 16, 2018; revised May 17, 2018; accepted November 12, 2018; published first November 21, 2018.

References

1. Fitzmaurice C, Allen C, Barber RM, Barregard L, Bhutta ZA, Brenner H, et al. Global, regional, and national cancer incidence, mortality, years of life lost, years lived with disability, and disability-adjusted life-years for 32 cancer groups, 1990 to 2015: a systematic analysis for the Global Burden of Disease Study. *JAMA Oncol* 2017;3:524–48.
2. Ryerson AB, Ehemann CR, Altekruse SF, Ward JW, Jemal A, Sherman RL, et al. Annual report to the nation on the status of cancer, 1975–2012, featuring the increasing incidence of liver cancer. *Cancer* 2016;122:1312–37.
3. Yang JD, Roberts LR. Hepatocellular carcinoma: a global view. *Nat Rev Gastroenterol Hepatol* 2010;7:448–58.
4. Yang JD, Kim B, Sanderson SO, St Sauver JL, Yawn BP, Pedersen RA, et al. Hepatocellular carcinoma in Olmsted county, Minnesota, 1976–2008. *Mayo Clin Proc* 2012;87:9–16.
5. Yang JD, Mannalithara A, Piscitello AJ, Kisel JB, Gores GJ, Roberts LR, et al. Impact of surveillance for hepatocellular carcinoma on survival in patients with compensated cirrhosis. *Hepatology* 2018;68:78–88.

Yang et al.

6. European Association For The Study Of The Liver; European Organisation For Research And Treatment Of Cancer. EASL-EORTC clinical practice guidelines: management of hepatocellular carcinoma. *J Hepatol* 2012; 56:908–43.
7. Bruix J, Sherman M. Management of hepatocellular carcinoma: an update. *Hepatology* 2011;53:1020–2.
8. Omata M, Cheng AL, Kokudo N, Kudo M, Lee JM, Jia J, et al. Asia-Pacific clinical practice guidelines on the management of hepatocellular carcinoma: a 2017 update. *Hepatol Int* 2017;11:317–70.
9. Yang JD, Kim WR. Surveillance for hepatocellular carcinoma in patients with cirrhosis. *Clin Gastroenterol Hepatol* 2012;10:16–21.
10. Yang JD, Harmsen WS, Slettedahl SW, Chaiteerakij R, Enders FT, Therneau TM, et al. Factors that affect risk for hepatocellular carcinoma and effects of surveillance. *Clin Gastroenterol Hepatol* 2011;9:617–23.e1.
11. Singal AG, Pillai A, Tiro J. Early detection, curative treatment, and survival rates for hepatocellular carcinoma surveillance in patients with cirrhosis: a meta-analysis. *PLoS Med* 2014;11:e1001624.
12. Tan CH, Low SC, Thng CH. APASL and AASLD consensus guidelines on imaging diagnosis of hepatocellular carcinoma: a review. *Int J Hepatol* 2011;2011:519783.
13. Singal AG, Conjeevaram HS, Volk ML, Fu S, Fontana RJ, Askari F, et al. Effectiveness of hepatocellular carcinoma surveillance in patients with cirrhosis. *Cancer Epidemiol Biomarkers Prev* 2012;21:793–9.
14. Simmons O, Fetzer DT, Yokoo T, Marrero JA, Yopp A, Kono Y, et al. Predictors of adequate ultrasound quality for hepatocellular carcinoma surveillance in patients with cirrhosis. *Aliment Pharmacol Ther* 2017;45:169–77.
15. Atiq O, Tiro J, Yopp AC, Muffler A, Marrero JA, Parikh ND, et al. An assessment of benefits and harms of hepatocellular carcinoma surveillance in patients with cirrhosis. *Hepatology* 2017;65:1196–1205.
16. Chang TS, Wu YC, Tung SY, Wei KL, Hsieh YY, Huang HC, et al. Alpha-fetoprotein measurement benefits hepatocellular carcinoma surveillance in patients with cirrhosis. *Am J Gastroenterol* 2015;110:836–44.
17. Kakisaka K, Kataoka K, Onodera M, Suzuki A, Endo K, Tatemichi Y, et al. Alpha-fetoprotein: a biomarker for the recruitment of progenitor cells in the liver in patients with acute liver injury or failure. *Hepatol Res* 2015;45: E12–20.
18. Yang JD, Dai J, Singal AG, Gopal P, Addissie BD, Nguyen MH, et al. Improved performance of serum alpha-fetoprotein for hepatocellular carcinoma diagnosis in HCV cirrhosis with normal alanine transaminase. *Cancer Epidemiol Biomarkers Prev* 2017;26:1085–92.
19. Johnson PJ, Pirrie SJ, Cox TF, Berhane S, Teng M, Palmer D, et al. The detection of hepatocellular carcinoma using a prospectively developed and validated model based on serological biomarkers. *Cancer Epidemiol Biomarkers Prev* 2014;23:144–53.
20. Berhane S, Toyoda H, Tada T, Kumada T, Kagebayashi C, Satomura S, et al. Role of the GALAD and BALAD-2 serologic models in diagnosis of hepatocellular carcinoma and prediction of survival in patients. *Clin Gastroenterol Hepatol* 2016;14:875–86.e6.
21. McMahon BJ, Bulkow L, Harpster A, Snowball M, Lanier A, Sacco F, et al. Screening for hepatocellular carcinoma in Alaska natives infected with chronic hepatitis B: a 16-year population-based study. *Hepatology* 2000;32:842–6.
22. Lok AS, Sterling RK, Everhart JE, Wright EC, Hoefs JC, Di Bisceglie AM, et al. Des-gamma-carboxy prothrombin and alpha-fetoprotein as biomarkers for the early detection of hepatocellular carcinoma. *Gastroenterology* 2010;138:493–502.
23. Yang JD, Kim WR, Coelho R, Mettler TA, Benson JT, Sanderson SO, et al. Cirrhosis is present in most patients with hepatitis B and hepatocellular carcinoma. *Clin Gastroenterol Hepatol* 2011;9:64–70.
24. Donato F, Tagger A, Gelatti U, Parrinello G, Boffetta P, Albertini A, et al. Alcohol and hepatocellular carcinoma: the effect of lifetime intake and hepatitis virus infections in men and women. *Am J Epidemiol* 2002;155: 323–31.
25. Adams LA, Lymp JF, St Sauver J, Sanderson SO, Lindor KD, Feldstein A, et al. The natural history of nonalcoholic fatty liver disease: a population-based cohort study. *Gastroenterology* 2005;129:113–21.
26. Chaiteerakij R, Addissie BD, Roberts LR. Update on biomarkers of hepatocellular carcinoma. *Clin Gastroenterol Hepatol* 2015;13: 237–45.
27. Marrero JA, Feng Z, Wang Y, Nguyen MH, Befeler AS, Roberts LR, et al. Alpha-fetoprotein, des-gamma carboxyprothrombin, and lectin-bound alpha-fetoprotein in early hepatocellular carcinoma. *Gastroenterology* 2009;137:110–8.
28. Youden WJ. Index for rating diagnostic tests. *Cancer* 1950;3:32–5.
29. DeLong ER, DeLong DM, Clarke-Pearson DL. Comparing the areas under two or more correlated receiver operating characteristic curves: a nonparametric approach. *Biometrics* 1988;44:837–45.
30. Caviglia GP, Abate ML, Petrini E, Gaia S, Rizzetto M, Smedile A. Highly sensitive alpha-fetoprotein, Lens culinaris agglutinin-reactive fraction of alpha-fetoprotein and des-gamma-carboxyprothrombin for hepatocellular carcinoma detection. *Hepatol Res* 2016;46:E130–5.
31. Sherman M, Feld J, Yamada H, Mori Y, Jassen H. A randomized controlled trial of US vs. US+biomarkers for the diagnosis of hepatocellular carcinoma: interim report. *J Hepatol* 2017;66:S13.
32. El-Serag H, White D, Tayob N, Alsarraj A, Mori Y, Yamada H, et al. Phase 3 biomarker study for HCC surveillance using AFP, AFP L-3 and DCP. A prospective collection with retrospective blinded evaluation. *J Hepatol* 2017;66:S222–S3.

Cancer Epidemiology, Biomarkers & Prevention

GALAD Score for Hepatocellular Carcinoma Detection in Comparison with Liver Ultrasound and Proposal of GALADUS Score

Ju Dong Yang, Benyam D. Addissie, Kristin C. Mara, et al.

Cancer Epidemiol Biomarkers Prev 2019;28:531-538. Published OnlineFirst November 21, 2018.

Updated version Access the most recent version of this article at:
doi:[10.1158/1055-9965.EPI-18-0281](https://doi.org/10.1158/1055-9965.EPI-18-0281)

**Supplementary
Material** Access the most recent supplemental material at:
<http://cebp.aacrjournals.org/content/suppl/2018/11/21/1055-9965.EPI-18-0281.DC1>

Cited articles This article cites 32 articles, 3 of which you can access for free at:
<http://cebp.aacrjournals.org/content/28/3/531.full#ref-list-1>

Citing articles This article has been cited by 2 HighWire-hosted articles. Access the articles at:
<http://cebp.aacrjournals.org/content/28/3/531.full#related-urls>

E-mail alerts [Sign up to receive free email-alerts](#) related to this article or journal.

**Reprints and
Subscriptions** To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department
at pubs@aacr.org.

Permissions To request permission to re-use all or part of this article, use this link
<http://cebp.aacrjournals.org/content/28/3/531>.
Click on "Request Permissions" which will take you to the Copyright Clearance Center's (CCC)
Rightslink site.