Short Communication

Fine-Mapping IGF1 and Prostate Cancer Risk in African Americans: The Multiethnic Cohort Study

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Abstract

Genetic variation at insulin-like growth factor I (IGF1) has been linked to prostate cancer risk. However, the specific predisposing variants have not been identified. In this study, we fine-mapped the IGF1 locus for prostate cancer risk in African Americans. We conducted targeted Roche GS-Junior 454 resequencing of a 156-kb region of IGF1 in 80 African American aggressive prostate cancer cases. Three hundred and thirty-four IGF1 SNPs were examined for their association with prostate cancer risk in 1,000 African American prostate cancer cases and 991 controls. The top associated SNP in African Americans, rs148371593, was examined in an additional 3,465 prostate cancer cases and 3,425 controls of non-African American ancestry—European Americans, Japanese Americans, Latinos, and Native Hawaiians. The overall association of 334 IGF1 SNPs and prostate cancer risk was assessed using logistic kernel-machine methods. The association between each SNP and prostate cancer risk was evaluated through unconditional logistic regression. A false discovery rate threshold of \( q < 0.1 \) was used to determine statistical significance of associations. We identified 8 novel IGF1 SNPs. The cumulative effect of the 334 IGF1 SNPs was not associated with prostate cancer risk (\( P = 0.13 \)) in African Americans. Twenty SNPs were nominally associated with prostate cancer at \( P < 0.05 \). The top associated SNP among African Americans, rs148371593 [minor allele frequency (MAF) = 0.03; \( P = 0.0014; q > 0.1 \)], did not reach our criterion of statistical significance. This polymorphism was rare in non-African Americans (MAF < 0.003) and was not associated with prostate cancer risk (\( P = 0.98 \)). Our findings do not support the role of IGF1 variants and prostate cancer risk among African Americans. Cancer Epidemiol Biomarkers Prev; 23(9); 1928–32. ©2014 AACR.

Introduction

Prostate cancer is the most common cancer in U.S. men. African Americans have the highest incidence rate of prostate cancer and at least twice the mortality rate of disease in comparison with other racial/ethnic groups (1). Insulin-like growth factor I (IGF1) is a potentially interesting candidate gene for prostate cancer as it stimulates cellular proliferation and inhibits apoptosis.

Men with higher levels of circulating IGF1 have been reported to have an increased risk of prostate cancer compared with men with lower levels of IGF1 (2–5). Past research and previous studies have linked genetic variation at IGF1 to prostate cancer risk (2, 3, 6); however, the specific predisposing IGF1 variants have not been identified. Detailed fine-mapping of the IGF1 locus may refine the genetic signal and aid in prioritizing risk variants for further follow-up and functional studies. Moreover, studying African Americans is an efficient means of localizing predisposing alleles given their high rates of prostate cancer and lower levels of linkage disequilibrium. These features provide for greater resolution in identifying IGF1 risk alleles and evaluating their effects among a population with the greatest burden of disease. Here, we conducted a fine-mapping study of the IGF1 locus and prostate cancer risk among African Americans.

Materials and Methods

Study subjects

The Multiethnic Cohort (MEC) Study is a large population-based cohort study of more than 215,000 men and women from Hawaii and Los Angeles. The cohort is
composed predominantly of individuals from five racial/ethnic groups: African Americans, Native Hawaiians, Japanese, Latinos, and Whites. Further methodological details of this study are provided elsewhere (7). Briefly, incident prostate cancer cases were identified by cohort linkage to population-based Surveillance, Epidemiology and End Results cancer registries covering Hawaii and California. Information on stage of disease and Gleason grade at the time of diagnosis was also collected from the cancer registries. Aggressive prostate cancer was defined as either regional, metastatic disease, or localized disease with Gleason grade >8. Controls had no diagnosis of prostate cancer and were randomly selected from the control pool of participants that provided blood specimens for genetic analysis. Controls were frequency matched to cases by age (±5 years) and ethnicity. For this study, our African American and non-African American case–control studies of prostate cancer nested in the MEC included 1,098 cases and 1,081 controls and 3,480 cases and 3,447 controls, respectively. This study was approved by the institutional review boards at the University of Hawaii, the University of Southern California, and the California Prevention Institute of California.

SNP discovery and selection
We used RainDance Technologies Custom Primer Library Design, and utilized Roche GS-Junior 454 next-generation sequencing technology to target and resequence 156 kb of IGF1 (including 50 kb downstream and 25 kb upstream, chromosome 12: 102,741,896–102,898,083, human genome assembly 18) in pooled samples of 80 African American prostate cancer cases with aggressive disease (8 pools of 10 samples each). Variant analysis was performed with Roche Amplicon Variant Analysis (AVA) software. For the eight pools, a total of 395 SNPs were identified as high-quality variants (maximum variant allele frequency (VAF) > 5% with maximum minor allele frequency (MAF) > 10% in regions of possible off-target reads). To increase our coverage of SNPs in this region, we selected 316 additional SNPs that had an MAF of >1% in Yorubans (YRI) from the 1000 Genomes Project, resulting in a total of 711 SNPs. These 711 SNPs were scored for Illumina GoldenGate assay design; 259 SNPs did not meet assay criteria of a sufficient design score and/or located within >60 base pairs of a neighboring SNP, resulting in 452 SNPs. To reach 384 SNPs for the Illumina GoldenGate genotyping assay, an additional 68 SNPs were excluded based on lower design scores and proximity to neighboring SNP.

Case–control genotyping
All assays were undertaken by laboratory personnel blinded to prostate case–control status. For African Americans, the Illumina GoldenGate assay was used to genotype 384 IGF1 variants in 1,098 African American prostate cancer cases and 1,081 controls. We excluded subjects with a call rate of <95% (n = 43) and those missing genetic ancestry estimates (n = 145), resulting in a final analysis set of 1,000 African American cases and 991 African American controls. Of the 384 variants selected for Illumina genotyping, 38 SNPs failed genotyping (call rate < 95%) or were not in Hardy–Weinberg equilibrium (P < 0.0001). Of the 38 SNPs that failed Illumina genotyping, 32 SNPs were prioritized for genotyping by the Taqman assay on the OpenArray platform. Between the Illumina GoldenGate and Taqman OpenArray genotyping, a total of 384 variants were genotyped in African American cases and controls, of which 6 SNP assays failed, 1 SNP was not in Hardy–Weinberg equilibrium (P < 0.0001), 6 SNPs were excluded due to a low call rate (<95%), and 41 were monomorphic. For the 13 SNPs that did not pass the quality control (QC), data were abstracted for 4 SNPs from a previous genome-wide association study (GWAS) of prostate cancer among these subjects (8). This resulted in a final analysis set of 334 SNPs with an average genotyping success rate of 99.83% and average genotype concordance rate for QC duplicates (~2% of all samples) of 98.5%.

For replication in non-African American samples, the Taqman assay was used to genotype the top associated SNP (rs148371593) in 3,480 prostate cases and 3,447 controls of European, Hawaiian, Japanese, and Latino ancestry. We excluded subjects (n = 37) that had no genotyping call, resulting in a total of 3,465 prostate cases and 3,425 controls. For non-African Americans, the average genotyping success rate was 99.5%, and the genotype concordance rate for QC duplicates (~2% of all samples) was 99.3%.

Ancestry estimation
For African Americans, to correct for genetic ancestry, we included in the regression analysis the first two eigenvectors from principal components analysis of previously collected GWAS data (genomic inflation factor λ = 1.08; ref. 8). For non-African Americans, 93 ancestry informative markers that captured the major continental genetic diversity (9) were previously genotyped (10), principal components of ancestry were estimated by EIGENSTRAT (11), and the first four eigenvectors were included in the regression analysis.

Association testing
For African Americans, to test for the overall effect of 334 IGF1 SNPs together, we performed a logistic kernel-machine test, which is better powered than the omnibus test in the presence of correlation between SNPs (6). To examine single SNP association with prostate cancer, we conducted unconditional logistic regression, adjusting for age, family history, and genetic ancestry. The P values were estimated by a 1-degree-of-freedom Wald test for trend. To correct for multiple testing, we applied the false discovery rate (FDR) method and used a threshold of q < 0.1 to determine statistical significance (5). All analyses were run using the R software (7) with the packages lme4 for the regression models, GenABEL to compute the genomic inflation factor λ, and SKAT for the
kernel-machine test (12). Manhattan and correlation plots were also done using the R software.

Results

The average ages for African American prostate cancer cases and controls were 69.3 and 69.8 years, respectively (Supplementary Table S1). Among all cases, 89.6% presented with localized disease and the remaining had regional/distant disease. Study characteristics for the non-African American subjects are presented in Supplementary Table S1.

Of the 334 IGF1 SNPs identified by sequencing African Americans, 8 SNPs were novel and not found in the 1000 Genomes (13) database (one was found in cases only with an MAF of 0.001, and the remaining 7 SNPs had an MAF in controls ranging from 0.0005 to 0.021; Supplementary Tables S2 and S3), and 74 SNPs were rare with an MAF of <0.01 (Supplementary Tables S1 and S2). In a kernel-machine test, there was no overall cumulative effect of the 334 IGF1 SNPs on prostate cancer risk among African Americans (P = 0.13). In single SNP analysis, 20 SNPs were nominally associated with prostate cancer risk at P < 0.05 (Fig. 1 and Table 1). However, none were statistically significant (q < 0.1) after performing FDR correction for multiple testing (5). The top association was with SNP rs148371593, located in the 3′ region of IGF1, for which the minor A allele (MAF = 0.027) was found to have an inverse association with prostate cancer risk in African Americans (OR, 0.83; 95% CI, 0.75–0.93; P = 0.0014). SNP rs148371593 was weakly or not correlated (r² < 0.45) with other SNPs in our set (Supplementary Fig. S1). In a non-African American sample set of 3,465 prostate cases and 3,425 controls, only 7 heterozygotes for rs148371593 were observed—2 Hawaiian (1 case and 1 control) and 5 of Latino ancestry (2 cases and 3 controls). The minor A allele was absent in European and Japanese ancestry populations, and it was extremely rare in Latino and Native Hawaiian populations (MAF of 0.003 and 0.001, respectively). Combining Latinos and Native Hawaiians, no significant association was observed (OR, 0.98; 95% CI, 0.53–1.82; P = 0.95; data not shown).

Discussion

In this fine-mapping study of prostate cancer risk among African Americans, we examined 334 IGF1 variants—8 of which were novel—and observed no statistically significant association between IGF1 variants and prostate cancer risk after correcting for multiple testing. The top association, a less common SNP, rs148371593, did not replicate in non-African American samples, for which it was found to be rare among Latinos and Native Hawaiians and nonexistent among Japanese and European Americans.

Numerous studies have indicated a possible involvement of IGF1 in prostate development and associated higher circulating levels of IGF1 with an increased risk of developing prostate cancer (2–5). In our previous haplotype-based study (2, 3, 6), we identified two upstream IGF1 variants associated with prostate cancer risk in the MEC Study (2). These two variants, rs7978742 and rs7965399, were originally reported to be associated with overall prostate cancer risk (P = 0.003 for both; ref. 2). In this study, with a larger sample size of African Americans, we observed nominal positive associations for these SNPs in African Americans (rs7978742, P = 0.03; rs7965399, P = 0.05), which was in line with our previously reported pattern of associations.

Subsequent studies confirmed an association at the IGF1 locus and prostate cancer risk in populations of European ancestry, in particular with variants rs7136446 and rs2033178 (13), and rs4764695 (14, 15). We observed a marginal nominal association with rs7136446 (P = 0.06) and could not confirm an association with the other two reported variants (P > 0.05) in African Americans.

Of the two variants (rs6220 and rs7136446) reported significantly associated with higher levels of circulating IGF1 by Johansson and colleagues (6), rs6220 was not examined in this study, though one correlated SNP, rs5009837 (r² = 0.91 among African Americans), was tested in our study and was not associated with prostate cancer risk (P = 0.47). The second variant, rs7136446, showed a nominal borderline association with prostate cancer risk in our study (P = 0.06). Two additional SNPs
rs1520220 and rs10735380, previously associated with high circulating levels of IGFI (16, 17), were not associated with prostate cancer risk in our study (P > 0.05).

Several GWASs of prostate cancer have failed to observe an association with IGFI and prostate cancer (18). The absence of an IGFI association in GWAS of prostate cancer coupled with our findings here would suggest that common IGFI variants do not play a role in prostate cancer susceptibility.

Sufficient statistical power is a major challenge of fine-mapping studies that aim to identify less common and rare risk alleles. While our study was able to observe nominal associations with variants that have an MAF ranging from <0.01 to 0.27, reduced study power is a limitation of our study. For less common variants with an MAF of 0.03, we had 80% to detect odds ratios of 1.60 among our African American sample, while for more common variants with an MAF of 0.20, we had 80% power to detect smaller odds ratios of 1.24.

Despite these limitations, the identification of novel and less common variants may be useful in further investigating the difference in prevalence and risk of prostate cancer between African Americans and other racial/ethnic groups. Large study populations of diverse ancestral populations are needed to confirm associations for less common variants.

### Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

### Authors' Contributions

Conception and design: D.O. Stram, D. Taverna, I. Cheng


Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): A. Lum-Jones, M. Tirikainen, D. Duggan, L. Le Marchand, I. Cheng


Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases): D. Taverna, A. Lum-Jones, C. Caberto, I. Cheng

Study supervision: I. Cheng

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### Table 1. Associations for 20 IGFI SNPs and prostate cancer risk among African Americans at P < 0.05

<table>
<thead>
<tr>
<th>SNP</th>
<th>Positiona</th>
<th>MAb</th>
<th>MAFc</th>
<th>OR (95% CI)</th>
<th>P</th>
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<tbody>
<tr>
<td>rs148371593</td>
<td>102773400</td>
<td>A/G</td>
<td>0.027</td>
<td>0.83 (0.75–0.93)</td>
<td>0.001</td>
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<td>chr12_102748461</td>
<td>102748461</td>
<td>A/G</td>
<td>0.001</td>
<td>1.43 (1.10–1.90)</td>
<td>0.008</td>
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<tr>
<td>rs78360701</td>
<td>102762488</td>
<td>A/G</td>
<td>0.001</td>
<td>1.42 (1.10–1.90)</td>
<td>0.012</td>
</tr>
<tr>
<td>rs5742687</td>
<td>102812676</td>
<td>G/A</td>
<td>0.001</td>
<td>1.42 (1.10–1.90)</td>
<td>0.012</td>
</tr>
<tr>
<td>rs140149239</td>
<td>102888147</td>
<td>A/C</td>
<td>0.004</td>
<td>1.27 (1.10–1.50)</td>
<td>0.013</td>
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<tr>
<td>rs140231439</td>
<td>102753563</td>
<td>G/A</td>
<td>0.009</td>
<td>1.18 (1.00–1.40)</td>
<td>0.016</td>
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<tr>
<td>rs138668850</td>
<td>102871977</td>
<td>A/C</td>
<td>0.025</td>
<td>0.87 (0.78–0.98)</td>
<td>0.018</td>
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<tr>
<td>rs7965399</td>
<td>102891686</td>
<td>G/A</td>
<td>0.267</td>
<td>1.04 (1.00–1.10)</td>
<td>0.030</td>
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<tr>
<td>rs76845058</td>
<td>102836572</td>
<td>G/C</td>
<td>0.009</td>
<td>0.81 (0.67–0.98)</td>
<td>0.033</td>
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<tr>
<td>rs146567847</td>
<td>102864270</td>
<td>A/G</td>
<td>0.009</td>
<td>1.17 (1.00–1.30)</td>
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</tr>
<tr>
<td>rs5742681</td>
<td>102813915</td>
<td>G/C</td>
<td>0.004</td>
<td>0.69 (0.49–0.97)</td>
<td>0.035</td>
</tr>
<tr>
<td>rs73187920</td>
<td>102752878</td>
<td>A/C</td>
<td>0.048</td>
<td>0.92 (0.84–0.99)</td>
<td>0.037</td>
</tr>
<tr>
<td>rs60741636</td>
<td>102792368</td>
<td>C/G</td>
<td>0.008</td>
<td>0.81 (0.66–0.99)</td>
<td>0.039</td>
</tr>
<tr>
<td>rs77458341</td>
<td>102894716</td>
<td>G/A</td>
<td>0.272</td>
<td>1.04 (1.00–1.10)</td>
<td>0.043</td>
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<tr>
<td>rs1787342</td>
<td>102835435</td>
<td>G/A</td>
<td>0.024</td>
<td>0.89 (0.79–1.00)</td>
<td>0.043</td>
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<tr>
<td>rs35638230</td>
<td>102742243</td>
<td>A/C</td>
<td>0.012</td>
<td>1.14 (1.00–1.30)</td>
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<tr>
<td>rs11111256</td>
<td>102761471</td>
<td>A/G</td>
<td>0.046</td>
<td>0.92 (0.85–1.00)</td>
<td>0.046</td>
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<td>rs56060310</td>
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<td>0.009</td>
<td>0.82 (0.67–1.00)</td>
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<td>rs11111285</td>
<td>102895256</td>
<td>G/A</td>
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<td>1.04 (1.00–1.10)</td>
<td>0.049</td>
</tr>
</tbody>
</table>

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*aChromosomal location based on NCBI build 37/hg19.

*bMajor/minor allele.

*cMinor allele frequency among controls.
References


