Research Article

Global Gene Expression Analysis in Cord Blood Reveals Gender-Specific Differences in Response to Carcinogenic Exposure *In Utero*

Kevin Hochstenbach[1], Danitsja M. van Leeuwen[1], Hans Gmuender[4], Ralf W. Gottschalk[1], Martinus Løvik[5], Berit Granum[5], Unni Nygaard[5], Ellen Namork[5], Micheline Kirsch-Volders[6], Ilse Decordier[6], Kim Vande Loock[6], Harrie Besselink[2], Margareta Törnqvist[7], Hans von Stedingk[7], Per Rydberg[7], Jos C.S. Kleinjans[1], Henk van Loveren[1,3], and Joost H.M. van Delft[1]

Abstract

Background: It has been suggested that fetal carcinogenic exposure might lead to predisposition to develop cancer during childhood or in later life possibly through modulation of the fetal transcriptome. Because gender effects in the incidence of childhood cancers have been described, we hypothesized differences at the transcriptomic level in cord blood between male and female newborns as a consequence of fetal carcinogenic exposure. The objective was to investigate whether transcriptomic responses to dietary genotoxic and nongenotoxic carcinogens show gender-specific mechanisms-of-action relevant for chemical carcinogenesis.

Methods: Global gene expression was applied in umbilical cord blood samples, the CALUX-assay was used for measuring dioxin(-like), androgen(-like), and estrogen(-like) internal exposure, and acrylamide–hemoglobin adduct levels were determined by mass spectrometry adduct-FIRE-procedure^{TM}. To link gene expression to an established phenotypic biomarker of cancer risk, micronuclei frequencies were investigated.

Results: While exposure levels did not differ between sexes at birth, important gender-specific differences were observed in gene expressions associated with these exposures linked with cell cycle, the immune system and more general cellular processes such as posttranslation. Moreover, oppositely correlating leukemia/lymphoma genes between male and female newborns were identified in relation to the different biomarkers of exposure that might be relevant to male-specific predisposition to develop these cancers in childhood.

Conclusions/Impact: This study reveals different transcriptomic responses to environmental carcinogens between the sexes. In particular, male-specific TNF-alpha-NF-kB signaling upon dioxin exposure and activation of the Wnt-pathway in boys upon acrylamide exposure might represent possible mechanistic explanations for gender specificity in the incidence of childhood leukemia. *Cancer Epidemiol Biomarkers Prev; 21(10); 1756–67. ©2012 AACR.*

Introduction

A gender effect in the incidence of childhood cancers has been well described (1, 2). The male-to-female inci-dence varies for several types of cancers; in particular, a male predominance in the age-adjusted incidence of all types of leukemias and lymphomas is apparent, with the highest ratio (M:F: 3.0) for non-Hodgkin lymphoma (2). Potential gender-specific risk factors include the faster growth rate of the male embryo because of accelerated rates of cell division and proliferation (3). For both sexes, birth weight have been positively associated with childhood leukemia, whereas the maternal dietary intake of DNA topoisomerase II inhibitors was associated with infant leukemia, altogether, they are the most common pediatric neoplasm (4). However, the etiology of sex-specific childhood cancer still needs to be fully clarified.

In this context, a link between environmental/dietary exposure to carcinogenic chemicals and infant leukemia has been proposed (5). Statistics show significant increases in the incidence of cancer including leukemia, among children in Europe over the last decades (6) suggesting that (maternal) exposure to carcinogenic factors present an etiological factor. Thus, investigations on

Authors' Affiliations: [1]Department of Toxicogenomics, Maastricht University, Maastricht, [2]BioDetection Systems B.V., Amsterdam, [3]Laboratory for Health Protection Research, National Institute of Public Health and the Environment, 3720 BA Bilthoven, The Netherlands;[4]Genedata, Basel, Switzerland; [5]Department of Food, Water and Cosmetics, Norwegian Institute of Public Health, Oslo, Norway; [6]Laboratory of Cellular Genetics, Free University Brussels, Brussels, Belgium; and [7]Department of Materials and Environmental Chemistry, Arrhenius Laboratory, Stockholm University, Stockholm, Sweden

Corresponding Author: Joost van Delft, Department of Toxicogenomics, Maastricht University, Universiteitssingel 50, PO Box 616, 6200 MD, Maastricht, the Netherlands. Phone: +31 (0)433882127; Fax: +31 (0) 433881092; E-mail: j.vandelft@maastrichtuniversity.nl

American Association for Cancer Research

possible mechanisms-of-action through which exposure to environmental carcinogens predispose to childhood cancer risk, is warranted. The latent period of, for example, leukemia, its very early onset in childhood might indicate the fetal period to be a critical window of exposure. It has been suggested that such fetal exposure might lead to predisposition to develop disease such as cancer and immune diseases during childhood or in later life possibly through modulation of the fetal transcriptome (7, 8).

Upon ingestion or inhalation by pregnant women, many environmental carcinogens are capable of crossing the placental barrier and reaching the fetus (9). It has been shown that the placental barrier does not protect the unborn child in case of maternal exposure to acrylamide, dioxins, and estrogen-like compounds such as organochlorine pesticides and amphenones (10–13).

Over the last decade, microarray-based genomics technology has significantly progressed enabling whole genome screening at the gene expression level, making this highly suitable for investigating mechanisms underlying, for example, the molecular effects of carcinogenic exposure. Because a gender effect in the incidence of childhood cancers exists, with cancer of the blood system being far more frequent in males than females, we hypothesized that differences at the transcriptomic level in cord blood will occur between male and female newborns as a consequence of fetal carcinogenic exposure, possibly leading to gender-specific predisposition to develop childhood leukemia.

For this, global transcriptomic responses in umbilical cord blood samples from Norwegian male and female newborns were examined in relation to markers of carcinogenic exposure and effect. Investigated nongenotoxic carcinogen exposures determined by means of DR CALUX, include aryl hydrocarbon (Ah) receptor-mediated compounds such as polychlorinated dibenzodioxins (PCDD), polychlorinated dibenzofurans (PCDF), and dioxin-like polychlorinated biphenyls (PCB; ref. 14). Measurements by CALUX assay of ERα and AR receptor activation represent exposure to endocrine disruptors such as carcinogenic pesticides like 1,1,1-trichloro-2,2-bis(*p*-chlorophenyl)ethane (DDT) and antiandrogens (15).

Global transcriptomic responses in male and female newborns were also examined in relation to genotoxic acrylamide exposure. Humans are exposed to relatively high doses of acrylamide because this compound is formed in food during cooking (16, 17). Acrylamide can be converted to the epoxide glycidamide by cytochrome P450 2E1 (18). Both compounds are electrophilically reactive. The mutagenicity from acrylamide exposure arises from the reaction of glycidamide with nucleophilic centers in DNA, as the direct interaction of acrylamide with DNA is slow (19). Acrylamide and glycidamide also bind to circulating proteins such as hemoglobin (Hb) and form adducts. This Hb-bound acrylamide represents a well-accepted measure of internal dose and was, as well as the Hb-bound glycidamide, used as a biomarker of exposure

to acrylamide in the present study. A recently developed liquid chromatography-tandem mass spectrometry method was used for measurement of the Hb adducts (20).

Gene expression was associated to a validated biomarker of cancer risk, that is, micronuclei frequencies (MN). MN is a well established biomarker of chromosomal breakage and/or whole chromosome loss that are unrepaired, misrepaired, or malsegregated because of chromosome, cellular, and nuclear dysfunction (21–23), and increased micronucleus frequencies in peripheral blood lymphocytes have been shown in prospective studies to predict the risk of cancer in humans (24).

Materials and Methods

Sample collection

Umbilical cord blood samples were collected immediately after birth from the cord vein of 200 babies whose mothers participated in the Norwegian BraMat cohort. As detailed below in section "Gene expression analysis", 45 male and 66 female newborns were included in the current study (Table 1). This BraMat subcohort is nested within the Norwegian Mother and Child cohort (MoBa). Study protocols were approved by the Regional Committee for Ethics in Medical Research in Oslo, Norway. Informed consent was obtained from all participating mothers before sample collection. The samples were collected by trained nurses at the maternity wards of the Ulleval and Akershus hospitals in Oslo, Norway. For preserving RNA for microarray analysis, aliquots of heparin-anticoagulated whole blood of 0.4 mL were mixed with 1.2 mL of RNAlater (Ambion/Applied Biosystems, Nieuwerkerk aan den Ijssel, the Netherlands) as soon as possible after blood collection. Samples were kept at $-80°C$ until shipment on dry ice to the research laboratory. The remainder of the whole blood was utilized for CALUX measurements, assessment of acrylamide- and glycidamide-Hb adducts, and analysis of MN.

RNA isolation

Total RNA was isolated from the RNAlater-preserved whole blood samples using the RiboPure-Blood system (Ambion) according to the manufacturer's instructions. RNA yield and purity was measured by means of NanoDrop equipment and integrity was verified by automated gel electrophoresis (2100 BioAnalyzer, Agilent Technologies).

Gene expression analysis

From the 200 BraMat samples, only RNA samples with a RNA integrity number (RIN) 6 or greater and a bench time (time from collection until RNA stabilization) of 6 or less hours, that is, 120 samples were used for transcriptomic analysis. Total RNA (1 microgram) was used to generate Cyanine-labeled cRNA using the Agilent Low RNA Input Linear Amplification Kit according to the manufacturer's instructions. Each individual cord blood sample was labeled by means of Cyanine-5 and

Table 1. Characteristics of the study population (A), RIN and bench time (B), and data distribution of the various parameters (C)

A			
	All	**Males**	**Females**
N	111	45	66
Birth weight	3629 ± 434	3722 ± 416	3564 ± 438
Gestation	39.8 ± 1.1	39.9 ± 1.1	39.7 ± 1.1
Age			
Mother	31.3 ± 4.2	31.0 ± 4.3	31.5 ± 4.2
Father	32.8 ± 6.4	32.3 ± 4.3	33.7 ± 5.7
Mother BMI BP	23.4 ± 3.9	23.1 ± 3.6	23.6 ± 4.1
Mother was pregnant before			
no	35%	34%	36%
yes	65%	66%	64%
Mother smoked BP			
no	84%	82%	85%
occasional	9%	11%	8%
daily	7%	7%	6%
Delivery start			
Spontaneous	81%	86%	77%
Induced	12%	7%	16%
Caesarean section	7%	7%	8%
Caesarean sections			
Planned	7	2	5
Emergency	7	4	3
B			
RIN (0–10)	8.0 ± 1.3	7.98 ± 1.30	7.97 ± 1.30
Bench time	77.8 ± 172.4	58.77 ± 116.66	90.63 ± 201.29
C			
DR CALUX	36.4 ± 9.8 (*n* = 43)	37.4 ± 10.5 (*n* = 14)	35.9 ± 9.6 (*n* = 29)
ERα CALUX	15.6 ± 14.4 (*n* = 34)	16.1 ± 10.6 (*n* = 17)	15.2 ± 17.8 (*n* = 17)
AR CALUX	0.08 ± 0.06 (*n* = 31)	0.07 ± 0.03 (*n* = 16)	0.09 ± 0.08 (*n* = 15)
AA Hb-adducts	16.5 ± 6.6 (*n* = 84)	16.0 ± 7.1 (*n* = 34)	16.7 ± 6.4 (*n* = 50)
GA Hb-adducts	10.0 ± 4.0 (*n* = 84)	9.5 ± 3.8 (*n* = 34)	10.3 ± 4.0 (*n* = 50)
‰MNBN	1.2 ± 0.9 (*n* = 33)	1.1 ± 0.8 (*n* = 13)	1.2 ± 0.9 (*n* = 16)

Abbreviation: BP, before pregnancy; RIN, RNA integrity number; Bench time, time from blood collection to RNA stabilization.
Data are presented as mean ± SD unless stated otherwise. Birth weight expressed in g, gestation in weeks, age in years, bench time in minutes, DR CALUX in pg TEQ/gr fat, ERα CALUX (ng EEQ/mL plasma), AR CALUX (ng AEQ/mL plasma), Hb-adducts in pmol/g Hb, ‰ MNBN in MN per 1,000 binucleated cells.

competitively hybridized against a common reference sample (pooled RNA cord blood samples, labeled with Cyanine-3) onto Agilent 4 × 44 k human oligonucleotide microarrays (Agilent Technologies) according to the manufacturer's instructions. After hybridization, microarrays were washed and scanned immediately using an Axon GenePix 4000B Microarray Scanner (Molecular Devices). Laser power was set to 100%. The photo multiplier tube (PMT) gain was set to a saturation tolerance of 0.02% to minimize background and saturated spots.

Additional sample exclusions were based on technical performance of the microarrays, as well as on the absence of informed consent forms or reported smoking of the mother at 30 weeks of pregnancy. The number of cases left in the study after exclusions was 111.

Internal measurements of pollutants

Dioxin-like activity in plasma was determined in a total of 43 samples using the dioxin responsive (DR) CALUX reporter gene bioassay. The bioassay comprises a genetically modified H4IIE rat hepatoma cell line containing the luciferase gene under control of the AhR. Approximately 3 grams of plasma from umbilical cord blood was extracted by means of shake-solvent extraction (hexane:

diethylether, 97:3). Extracted fat was used for clean-up on an acid silica column (20% and 33% H_2SO_4), topped with sodium sulphate. Cleaned extracts were dissolved in dimethyl sulfoxide (DMSO; 8 µL); the DR CALUX activity was determined following exposure to dilution series of cleaned extracts (0.8% DMSO) for 24 hours. Estrogenic and androgenic activity in umbilical cord blood were determined in a total of 34 and 31 samples, respectively, using a human U2OS cell line stably transfected with the luciferase gene that is under control of the estrogen receptor alpha (ERα CALUX) or androgen receptor (AR CALUX). Approximately 0.5 mL of cord blood plasma was extracted by means of shake-solvent extraction (MTBE; methyltertiairbutylether). Extracts were dissolved in DMSO (40 µL); the ERα and AR CALUX activity was determined following 24 hours of exposure at various dilutions of the redissolved extracts (0.1% DMSO). All bioassays were corrected for procedure blanks and conducted including a reference compound calibration curve. By interpolation, final results are expressed as reference compound equivalents (eq.): DR CALUX are expressed in pilogram 2,3,7,8-TCDD-like eq. (TEQ)/gram fat, ERα CALUX in ng 17β-estradiol-like eq. (EEQ)/mL plasma, AR CALUX in nanogram dihydrotestosterone-like eq. (AEQ)/mL plasma (25, 26).

Hemoglobin-adduct levels

Hb-adducts from acrylamide (AA Hb-adducts) and glycidamide (GA Hb-adducts) were measured as internal dose markers of exposure to acrylamide from food. Red blood cells were isolated and stored at $-20°C$ until further processing. Adduct levels to N-terminal valines in Hb were measured in a total of 84 samples applying the "adduct FIRE procedure" using liquid chromatography–tandem mass spectrometry (LC–MS/MS; Shimadzu Prominence, AB SCIEX 3200 qtrap) for final detection

(20). Method performance parameters and its application to measurements of acrylamide and glycidamide Hb-adducts in cord blood samples has previously been described (20, 27).

Cytokinesis-block micronucleus assay

MNs per 1,000 binucleated cells (‰MNBN) were analyzed in peripheral blood T-lymphocytes of 33 samples. MNBN, in addition to *in vivo*-accumulated MN, reflect damage present on DNA or key proteins and expressed as MN during *in vitro* cell division. MN in mononucleated cells (MNMONO) only reflect chromosomal damage before the start of the assay. Because gene expression is not purely a cumulative measure, and MNMONO frequencies were low, MNBN were used. The *in vitro* cytokinesis-blocked MN assay (CBMN assay) was carried out according to the standardized protocol developed for semiautomated image analysis system. (Decordier et al. 2009) (28). Sampling and making of cultures occurred in Oslo, Norway. The protocol was conducted as recently described by Vande Loock and colleagues (29).

Data analysis

The data discussed in this publication have been deposited in NCBI's Gene Expression Omnibus (30) and are accessible through GEO Series accession number GSE31836 (31). Scan images of Cy5- and Cy3- channels were loaded into ImaGene software version 8.0.1 (BioDiscovery) for the extraction of raw pixel intensities and local backgrounds. The data were preprocessed using the Refiner Array module from the Genedata Expressionist system (Genedata AG). Raw data were assessed for quality and preprocessed as follows. Signals were corrected by subtracting backgrounds without producing negative values. For each feature and array, the contrast, that is, the quality for each signal according to its signal-to-noise

Table 2. Numbers of genes correlating with CALUX, Hb-adducts, and micronuclei

	No. of genes	Pos. correlation	Neg. correlation	Gender	No. of genes	Pos. correlation	Neg. correlation	Overlap boys-girls
DR CALUX	**3**	0	3	Boys ($n = 14$)	**371**	163	208	1
				Girls ($n = 29$)	**39**	29	10	
ERα CALUX	**20**	14	6	Boys ($n = 17$)	**493**	311	182	3
				Girls ($n = 17$)	**626**	459	167	
AR CALUX	**83**	69	14	Boys ($n = 16$)	**1293**	946	347	29
				Girls ($n = 15$)	**508**	366	142	
AA Hb-adducts	**0**	0	0	Boys ($n = 34$)	**23**	18	5	0
				Girls ($n = 50$)	**0**	0	0	
GA Hb-adducts	**0**	0	0	Boys ($n = 34$)	**6**	2	4	0
				Girls ($n = 50$)	**0**	0	0	
‰MNBN	**22**	4	18	Boys ($n = 13$)	**1397**	662	735	1
				Girls ($n = 16$)	**95**	28	67	

Numbers are presented for sexes grouped together and separately, correlation coefficients more than 0.60 or less than -0.60, and an unadjusted P value ≤ 0.05.

ratio, the distortion, and imbalance were determined. Defective or saturated features detected by the scanner software or by the Refiner Array module were diagnosed and masked. LOWESS correction was used to correct all features so that the signal distortion and imbalance of the 2 channels is minimized. Quality criteria for the data analysis were set as follows: signal-to-noise ratios more than 2, relative errors less than 0.5, and saturated and features flagged as masked filtered out.

Dose–response relationships were investigated with the available samples by Pearson's correlation analyses of individual gene expressions (Cy5/Cy3 ratios) and individual values for CALUX measurements, Hb-adducts, and MN, as mentioned in Table 2. Correlation analyses were conducted only for transcripts with least 50% valid expression values.

Significantly correlating transcripts were selected using 2 cut-offs. Minimal correlation coefficients were set to 0.60 or more or −0.60 or less and an unadjusted P value of less than 0.05. To ensure maximum specificity, a higher cut-off of 0.75 or more or −0.75 or less was used. No false discovery rates were used, because correlation coefficients were considered more important.

To identify biological processes in which significantly correlating genes are involved, the software suite Metacore™ was used. Gene ontology (GO) terms, pathways, and gene networks containing at least 2 significantly correlating genes and a P-value 0.01 or less were considered significantly enriched. In addition, a false discovery-rate cut-off of 0.25 was applied. For enabling further biological interpretation of correlation data, correlation coefficients were uploaded in Tox-Profiler without any statistical pre-selection of correlating genes. Tox-Profiler uses the T-test to score differences between mean input values (usually expression values, but in our case correlation coefficients) of predefined gene sets and that of all other genes (32). For biological interpretation, gene sets based on annotations from different databases were used, that is, GO terms, KEGG, Biocarta, and Wiki-Pathways. To identify significant effects, a Wilcoxon non-parametric statistical test was used, using an unadjusted E-value cut-off of 0.05.

To evaluate differences at the transcriptomic level between male and female newborns in relation to blood cancer risk, the effects on human cancer genes, that is, genes in which mutations that are causally implicated in oncogenesis have been reported, were investigated (33). For this, only genes connected to leukemia and lymphoma were included which showed an opposite correlation coefficient between male and female neonates. Additionally, the correlation coefficient had to be 0.60 or more or −0.60 or less in either male or female newborns.

Results

Biomarker analyses

From a total of 200 samples from the BraMat cohort, 120 RNA samples were selected on the basis of a RIN 6 or

Table 3. The GO terms, pathways, and networks enriched among the significantly correlating genes for the various biomarkers for males

GO term/Pathway/Network	# significant processes (CC $\geq$ 0.60/CC $\geq$ 0.75)	P-value (# genes affected/total # in pathway)	Database
DR CALUX	29/16		
Cell cycle		3.76E-04 (31/679)	GO
Apoptosis		5.63E-04 (26/543)	GO
ERα CALUX	50/9		
Antigen processing and presentation		2.92E-05 (9/61)	GO
Regulation of adaptive immune response		1.69E-04 (8/60)	GO
AR CALUX	7/1		
Cell cycle_G1-S		6.64E-03 (17/122)	Pathways
Proteolysis_Ubiquitin–proteasomal proteolysis		7.11E-03 (19/143)	Pathways
AA Hb-adducts	4/0		
Regulation of developmental process		5.51E-03 (4/637)	GO
Regulation of T-cell activation		8.89E-03 (2/121)	GO
GA Hb-adducts	0/0		
‰MNBN	22/53		
DNA damage_Checkpoint		9.22E-03 (16/92)	Networks
Regulation of cell cycle[a]		3.49E-04 (14/314)	GO
Apoptosis and survival_p53-dependent apoptosis[a]		1.06E-03 (4/25)	Pathways

Only processes mentioned in the text are presented; for complete lists refer to Supplementary data Table S1.
[a]Found when using the 0.75 correlation coefficient cut-off.

more and a bench time (time from collection until RNA stabilization) of 6 or less hours. These 120 samples were hybridized onto microarrays. Sample/microarray exclusions were based on technical performance of the microarrays, as well as on the absence of informed consent forms or reported smoking of the mother at 30 weeks of pregnancy. The number of cases left in the study after exclusions was 111. Characteristics of the remaining study population are presented in Table 1A, for the whole study population as well as for male and female infants separately. All mothers of the included 111 newborns are nonsmokers. Table 1B presents RIN and bench time, that is, time from collection until RNA stabilization, whereas Table 1C presents the data distributions for the investigated parameters on CALUX, Hb-adducts, and MN. The number of available samples for each biomarker varies as presented in Table 1C. For all these measurements, no significant differences were observed between male and female newborns.

Number of significantly correlating genes

Correlation analyses of gene expression levels with biomarkers of exposure and with MN measured showed variable numbers of significantly correlating genes (Table 2). Because of missing values, the numbers of available samples for correlation analysis vary per biomarker and were significantly lower than the 111 mentioned in Table 1. Overall, separate analyses for male and female newborns resulted in higher numbers of significantly correlating genes per gender with low overlap of similarly expressed genes between the 2 sexes, thus indicating a clear gender-specific toxicogenomic response.

Biologic evaluation

DR CALUX. Correlation analysis showed that male infants show a higher number of significantly correlating transcripts in relation to dioxin and dioxin-like exposure, assessed by DR CALUX assay, when compared with females. For males, functional interpretation revealed significantly affected processes relevant to carcinogenicity, such as programmed cell death/apoptosis, spindle/nuclear division, and the ubiquitination–proteasome pathway (Table 3). In females, mainly epigenetic responses were found, such as nucleotide and chromatin assembly (Table 4). The mitotic cell cycle appeared to be affected in both males and females.

Tox-Profiler results showed more processes to be significantly affected in female neonates compared with males (Table 5). Female-specific processes included translation and posttranslational modification. Tox-Profiler also confirmed the epigenetic responses in females, although these were also (not-significantly) positively correlated in males. T- and B-cell receptor signaling pathways were found to be suppressed in females. Translational processes were also found for male newborns. Interestingly, these were oppositely regulated when compared with females. Also, proteasome degradation was found to be oppositely regulated between the sexes as well as the TNF-alpha-NF-kB signaling pathway.

ERα CALUX. Female newborns showed a higher number of significantly correlating transcripts in relation to estrogen and estrogen-like exposure when compared with males. Accordingly, more processes were found to be significantly affected in females, of which the majority was immune related. Other processes included more general posttranslation events, whereas estrogen (-like)

Table 4. The GO terms, pathways, and networks enriched for males among the significantly correlating genes for the various biomarkers for females

GO term/Pathway/Network	# Significant Processes (CC $\geq$ 0.60/CC $\geq$ 0.75)	P-Value (# Genes Affected/Total # in Pathway)	Database
DR CALUX	18/0		
Nucleosome assembly		1.22E-05 (4/69)	GO
Cell cycle_Initiation of mitosis		6.37E-03 (2/18)	Pathways
ERα CALUX	74/88		
Immune system process		8.49E-06 (58/863)	GO
Regulation of lymphocyte activation		1.99E-05 (18/155)	GO
AR CALUX	39/33		
Translational elongation		8.72E-19 (28/104)	GO
Ubiquitin-dependent protein catabolic process		4.70E-06 (21/214)	GO
AA Hb-adducts	0/0		
GA Hb-adducts	0/0		
‰MNBN	3/6		
Nucleotide-excision repair, DNA damage removal		1.30E-04 (3/18)	GO
Cell cycle_Initiation of mitosis		6.37E-03 (2/18)	Pathways

Only processes mentioned in the text are presented; for complete lists refer to Supplementary data Table S2.

Table 5. GO terms and pathways enriched for males among all correlating genes using Tox-Profiler

Biomarker	Process	# Significant Processes Males/Females	T-Value Males	P-Value Males	T-Value Females	P-Value Females	Database
DR CALUX		5/29					
	Nucleosome assembly		4.1	0.154	6.4	<0.001	GO
	T-cell receptor signaling pathway		−2.7	1.000	−4.5	0.001	KEGG
	B-cell receptor signaling pathway		−0.6	1.000	−4.1	0.005	KEGG
	TNF-alpha-NF-kB Signaling Pathway		2.9	0.551	−4.2	0.010	WikiPathways
ERα CALUX		2/41					
	Immune response		−1.4	1.000	6.8	<0.001	GO
	T-cell receptor signaling pathway		−1.5	1.000	6.8	<0.001	WikiPathways
	B-cell receptor signaling pathway		−1.9	1.000	4.9	<0.001	WikiPathways
AR CALUX		12/24					
	Immune response		−2.5	1.000	−6.6	<0.001	GO
	Ubiquitin-dependent protein catabolic process		0.9	1.000	5.9	<0.001	GO
	Nucleosome		−4.7	0.020	−1.2	1.000	GO
AA Hb-adducts		4/17					
	Translational elongation		−1.7	1.000	8.8	<0.001	GO
	RNA splicing		−5.6	<0.001	1.7	1.0000	GO
GA Hb-adducts		8/12					
	Wnt signaling pathway		4.2	0.032	0.2	1.000	KEGG
‰MNBN		30/13					
	Translational elongation		8.5	<0.001	−5.7	<0.001	GO
	Spliceosome		4.6	0.002	−5.4	<0.001	KEGG
	mRNA processing		4.0	0.012	−4.1	0.002	WikiPathways
	Pathways in cancer		4.8	0.001	1.8	1.000	KEGG
	Translational elongation		8.5	<0.001	−5.7	<0.001	GO

Only processes mentioned in the text are presented; for complete lists refer to Supplementary data Table S3.

exposure also appeared to affect apoptosis Table 4). Tox-Profiler-based analysis in particular showed the immune response to be positively correlated in females. Also, more general cellular processes mostly related to transcription and posttranslational processes were identified. Interestingly, all processes found using Tox-Profiler were oppositely regulated between the sexes, although mainly reaching significance in females only (Table 5).

AR CALUX. Male newborns showed a higher number of significantly correlating genes with AR CALUX involved in immune response, cell-cycle regulation, ubiquitin–proteasomal proteolysis and cytoskeletal-related processes (Table 3). Functional enrichment analyses on significantly correlating genes for females resulted in an extensive list comprising general cellular processes like posttranslation, metabolic, and biosynthesis processes (Table 4). This finding is confirmed by the Tox-Profiler analysis, which also shows general cellular processes to be significantly affected in female newborns (Table 5).

Hemoglobin-adduct level. Analyses revealed only weak correlation of GA Hb-adducts with gene expression, leaving too few significantly correlating genes to conduct biological interpretation analyses. For AA Hb-adducts, in particular for males, regulation of developmental process, exocytosis, and regulation of T-cell activation and neuron differentiation was found (Tables 3 and 4). Tox-Profiler results revealed general processes oppositely regulated between the 2 sexes, although significance was generally found for females, except for mRNA splicing, which was significantly (negatively) correlated with AA Hb-adducts in males (Table 5).

Micronuclei frequency. Male newborns showed a higher number of genes significantly correlating with MN, when compared with female newborns (1397 vs. 95; Table 2). Functional enrichment analyses using a correlation coefficient cut-off of 0.60 identified general cellular processes like translation and RNA processing, next to a number of immune-related processes. A higher cut-off of 0.75, decreasing the number of genes to 270 and thus increasing specificity, resulted in more specific processes such as nuclear fragmentation during apoptosis and cell-cycle regulation (Table 3). Despite the limited number of significantly correlating genes for female newborns, cell-cycle regulation and DNA damage removal

	Gene symbol	Agilent gene ID	CC Males	CC Females	Regulation of progression through cell cycle**	Proliferation/replication**	Apoptosis**	Regulation of transcription**	Cromatin/histone**	Immune response**
DR										
	JAK2	Janus kinase 2 a protein tyrosine kinase	0.77*	-0.18						
	EWSR1	Ewing sarcoma breakpoint region 1	0.69*	-0.20						
	HSP90AB1	Heat shock protein 90kDa alpha cytosolic, class B member 1	0.62*	-0.26						
	ZNF384	Zinc finger protein 384	0.71*	-0.12						
Era										
	FUS	Fusion involved in t 12;16 in maligt liposarcoma	-0.66*	0.70*						
	CD79B	CD79b molecule, immunoglobulin-associated beta	-0.50	0.70*						
	LCK	Lymphocyte-specific protein tyrosine kinase	-0.25	0.80*						
	MLLT6	Myeloid/lymphoid or mixed-lineage leukemia trithorax homolog, Drosophila; translocated to, 6	0.39	-0.63*						
AR										
	DEK	DEK oncogene DNAbinding	-0.43	0.72*						
	MLLT3	Myeloid/lymphoid or mixed-lineage leukemia trithorax homolog, Drosophila; translocated to, 3	-0.50	0.65*						
	KRAS	v-Ki-ras2 Kirsten rat sarcoma viral oncogene homolog	-0.49	0.63*						
	IGH@	Immunoglobulin heavylocus	0.62*	-0.41						
‰ MNBN										
	BCL2	B-cell CLL/lymphoma 2	-0.77*	0.49						
	RUNX1	Runt-related transcription factor 1 acute myeloid leukemia 1; aml1 oncogene	-0.80*	0.33						
	TPM3	Tropomyosin 3	0.72*	-0.34						
	EIF4A2	Eukaryotic translation initiation factor 4A, isoform 2	0.69*	-0.35						
AA Hb-adducts										
	NIN	Ninein GSK3B interacting protein	-0.64*	0.12						
	PBX1	Pre B-cell leukemia homeobox1	0.66*	-0.09						
GA Hb-adducts										

Figure 1. Human leukemia and lymphoma-associated genes with opposite correlations with biomarkers between male and female newborns. Only the top 4, based on deltaCC are presented; for complete lists refer to Supplementary Table S4. * indicates a significant correlation, with an unadjusted P value less than 0.05. ** indicates the query terms used to search the GO term annotations for each gene.

appeared to be enriched (Table 4). Tox-Profiler analysis confirmed the significant correlation of immune-related processes in males. Also, the translational and posttranslational processes found by using significantly correlating transcripts were confirmed, showing negatively correlation with MN in males, where a significant positive correlation was found in females.

No significant correlations were observed between MN and the exposure markers when the subjects were grouped together as well as for females separately. Only in males, acrylamide and glycidamide hemoglobin adducts correlated significantly with MN (CC = 0.75 with $P = 0.019$ and CC = 0.73 with $P = 0.025$, respectively).

Inversely correlating effects on leukemia- and lymphoma-associated genes. Finally, to evaluate differences at the transcriptomic level between male and female newborns in relation to blood cancer risk, we investigated the

effects on human cancer genes. Figure 1 presents the top 4 (based on deltaCC) oppositely correlating cancer genes between male and female newborns in association with the biomarkers. In total, 4 transcripts correlated inversely in relation to DR CALUX, 13 in relation to ERα CALUX, 7 in relation to AR CALUX, 18 in relation to MN, 2 in relation to acrylamide, and none in relation to glycidamide (Supplementary data Table S4).

Discussion

Because a gender effect in the incidence of childhood cancers is known to exist, we hypothesized differences at the transcriptomic level in cord blood between male and female newborns as a consequence of fetal carcinogenic exposure. Concordantly, different transcriptomic responses to environmental carcinogens between the

sexes were observed. Taking into account that the largest differences between sexes in childhood cancer are found for leukemias and lymphomas (2), and that we measured transcriptomics, exposure biomarkers and effect biomarkers in blood of newborns, this discussion will focus on the relation between transscirptome data and childhood risks for acute lymphoblastic leukemia (ALL).

DR CALUX

The nongenotoxic carcinogen exposures determined by means of DR CALUX, comprise compounds that activate the Ah receptor, such as PCDDs, PCDFs, and dioxin-like PCBs (14). In our study, far more genes correlated with DR CALUX in males than in females (Table 2). In addition, functional interpretation has shown large gender differences in dose-dependent transcriptomics responses.

An epigenetic response to Ah-receptor-mediated exposure was found, showing nucleosome-related processes positively correlating with DR CALUX mainly in females. Effects on the nucleosome are in concordance with the known mechanism-of-action of dioxins; activation of transcription by dioxin is accompanied by changes in chromatin structure, which depend upon a functional aromatic hydrocarbon (Ah) receptor (34). It is, however, unclear why this is sex-specific and how this could affect ALL susceptibility. The (mitotic) cell cycle was mainly affected in males. Although it is generally accepted that Ah receptor activation causes toxic and carcinogenic effects through its role in detoxification by modulating expression of metabolizing enzymes, activation of the Ah receptor possibly also participates in pathways such as cell-cycle regulation, apoptosis, and immune response (35). Possibly, the activation of cell-cycle regulating genes via the Ah-receptor in male neonates is stronger than in females, and therefore males are more susceptible to develop ALL because of exposure to PCB and other Ah-receptor activators.

T- and B-cell receptor signaling pathways are negatively correlated with dioxin exposure in female neonates. Fetal TCDD exposure in animals has been linked to inhibition of cellular differentiation and maturation, primarily targeting T lymphocytes, leading to thymic atrophy and immunosuppression in offspring (36). This may suggest that the immune system is more suppressed in females exposed to Ah-activators, than in males. That would imply a higher risk in females, which is not the case in childhood ALL. However, a higher incidence has been reported in female infants (37) possibly suggesting T- and B-cell receptor signaling pathways to mechanistically underlie the increased risk of developing of infant ALL.

Interestingly, the TNF-alpha-NF-kB signaling pathway is negatively correlated with dioxin exposure in females, whereas (not significantly) positively correlated in male newborns. The association of inflammation and cancer has been studied extensively (38) showing TNF-α acts as a key mediator of inflammation and cancer (39). Although this pathway is involved in general cancer susceptibility, NF-kB activation has been consistently associated with childhood ALL (40, 41). The involvement of the NF-kB pathway has been suggested to play a role in the sex-specific character of childhood leukemia based on the association of the *IRF4* gene with higher male-specific risk with childhood ALL (42). *IRF4* was, however, not found to correlate with dioxin exposure, although we found several other genes showing differential gender-specific correlation within the TNF-alpha-NF-kB signaling pathway, including, for example, *TRAF1*, *TRAF3*, *MAP3k3*, *MAP3k14*, and *CASP8*. This might further indicate the TNF-alpha-NF-kB signaling pathway as a possible contributing factor in relation to the male predominance in childhood ALL. Moreover, the unaffected or even nonsignificantly increased signaling by tumor necrosis factor (TNF)-α in males might also result in a failure of the lymphocytes to undergo apoptosis (43), possibly contributing to the increased risk of childhood ALL in males. This is underlined by the male-specific negative correlation of *CASP8* with dioxin exposure. Accordingly, apoptosis was found to be exclusively affected in males. Nine human blood cancer-associated genes were found to be inversely correlating strongly indicating a gender-specific response to dioxins in relation to tumorigenesis. The human antiapoptotic cancer genes *JAK2* and *VHL* are positively correlated with dioxin exposure in males, whereas negatively in females contributing to apoptosis blockade, possibly predisposing males to develop childhood ALL. These cancer genes have been associated with childhood ALL (44–45).

In conclusion, the female-specic suppression of the TNF-alpha-NF-kB signaling pathway upon dioxin exposure might protect female neonates from developing childhood ALL, whereas their male counterparts are at higher risk, possibly because of consequent blockade of apoptosis.

ERα CALUX

Estrogens have been linked to B- and T-cell activation, enhanced antigen presentation by dendritic cells, and cytokine production (46). Accordingly, we found immune-related processes to be affected, with a higher response in females compared with males. In fact, results using unfiltered correlation data suggest an inverse regulation upon estrogen receptor activation with a significant stimulation of the GO term "immune system" in females and (nonsignificant) suppression in males. GO terms like B- and T-cell signaling show similar inverse trends. The male-specific immune suppression may contribute to the higher incidence of childhood ALL because leukemia is associated with immune dysfunction (47). Further confirming gender-specificity, we found leukemia- and lymphoma-related genes differentially correlating with estrogen exposure (Figure 1). Not all genes conclusively point to a male predisposition for developing ALL, in fact we found a female-specific-positive correlation of *MllT6*. Mixed lineage leukemias are histone methyl-transferases that regulate gene activation and are also well known to be rearranged in acute myeloid and

lymphoid leukemias (48). Interestingly, the female-specific correlation might be in line with the literature. Unlike childhood leukemia diagnosed in childhood, where there is a predominance in males, high frequencies of MLL rearrangements in female Infant leukemia have been reported, as well as a higher risk for females to develop infant ALL (37, 49).

AR CALUX

Next to androgens, certain environmental pollutants may influence AR signaling pathways by acting as or interfering with endogenous androgens, regulating the expression of various genes involved in cell-cycle control, apoptosis, cell growth, and differentiation (50). We found the cell-cycle GO term to correlate significantly to AR CALUX in males. Possibly, the activation of cell-cycle genes via the AR-receptor in male neonates is stronger than in females, and leading to a higher males-specific susceptibility. In contrast, the immune response was significantly suppressed in only females implying a higher cancer risk in females. Concordantly, differentially correlating leukemia and lymphoma genes with AR receptor activation do not conclusively support a male predisposition. In fact, next to the *MLLT6* gene correlating with the ERα receptor, a female-specific positive correlation of *MllT3* with AR receptor activation was found which might be related to female predisposition to develop infant leukemia (37, 48, 49).

Hb-adducts

Tox-Profiler revealed the Wnt signaling pathway and a number of general processes oppositely regulated between the 2 sexes upon acrylamide exposure. Aberrations of the Wnt pathway have been associated with various types of cancer including ALL. A male-specific activation of the Wnt pathway might thus contribute to the higher incidence of cancer in the blood system.

Micronuclei frequencies

Micronuclei formation in peripheral blood T-lymphocytes may be regarded as a phenotypic endpoint that reflects the deleterious effects on chromosomes because of the (combined) exposure to genotoxic carcinogens (51). Previous studies report that the baseline MN frequency is relatively low in newborns and no differences in gender have been identified (29, 52), which is confirmed in our study. Despite this, the sex-specific transcriptomic profiles associated with MN differ dramatically. Overall, a higher number of significantly correlating genes in male newborns in relation to MN was observed. Correlations were frequently opposite between males and females. The male-specific profile contains many more genes relevant to ALL and carcinogenicity in general. This might contribute to the higher childhood cancer risk in males. Interestingly, 6 genes of our MN correlating genes are also present in the recently published MN gene network (53). They were found to be oppositely correlating between male and female neonates, that is, *BCL2*, *CDC20*,

PTTG1, *BAX*, *PCNA*, and *LMNA*, suggesting sex-specific contribution to the network. Finally, MN-associated sex-specific differences in general cellular processes such as transcription, posttranslational processes, RNA processing, and splicing might be of relevance with regard to the reported gender differences in the incidence of childhood cancers, however, interpreting these findings is difficult.

Conclusions

While exposure to acrylamide and endocrine disruptors did not differ significantly between sexes at birth, distinct gender-specific dose- and response-related transcriptomic profiles were identified. In particular, male-specific TNF-alpha-NF-kB signaling upon dioxin exposure and subsequent failure of the lymphocytes to undergo apoptosis, and activation of the Wnt pathway in boys upon acrylamide exposure might represent possible mechanistic explanations for the gender-specificity in the incidence of childhood ALL. Interestingly, oppositely correlating leukemia and lymphoma genes between male and female newborns were identified in relation to the different biomarkers of exposure which might be relevant to male-specific predisposition to develop childhood ALL and female-specific predisposition to develop infant ALL.

Our study has the strength of considering multiple parameters in relation to gene expression, although we recognize that the correlation analyses in the current study of gene expression with the various biomarkers are based on a relatively small number of samples making multivariate correlation analyses impractical. This is because of the limited availability of samples with biomarker measurements and gender-specific analyses, which reduces statistical power. Nevertheless, because the gender-specific responses occur for multiple biomarkers, we are confident about our conclusions. To confirm the generated hypotheses, future research based on higher number of samples is needed.

Authors' Contributions

Conception and design: D. van Leeuwen, M. Løvik, B. Granum, M. Kirsch-Volders, J. C.S. Kleinjans, H. van Loveren, J. H.M. van Delft
Development of methodology: D. van Leeuwen, H. Besselink, P. Rydberg, J. C.S. Kleinjans, H. van Loveren, J. H.M. van Delft
Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): K. Hochstenbach, M. Løvik, B. Granum, U. C. Nygaard, E. Namork, K. vande Loock, H. Besselink, M. Törnqvist, P. Rydberg
Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): K. Hochstenbach, D. van Leeuwen, H. Gmuender, M. Kirsch-Volders, M. Törnqvist, H. von Stedingk, H. van Loveren
Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases): R. W. Gottschalk, B. Granum
Writing, review, and/or revision of the manuscript: K. Hochstenbach, M. Løvik, B. Granum, U. C. Nygaard, E. Namork, M., J. H.M. van Delft Kirsch-Volders, I. Decordier, H. Besselink, M. Törnqvist, H. von Stedingk, J. C.S. Kleinjans, H. van Loveren
Study supervision: D. van Leeuwen, M. Løvik, H. van Loveren, J. H.M. van Delft

Grant Support

References

1. Cartwright RA, Gurney KA, Moorman AV. Sex ratios and the risks of haematological malignancies. Br J Haematol 2002;118:1071–7.
2. Linet MS, Wacholder S, Zahm SH. Interpreting epidemiologic research: lessons from studies of childhood cancer. Pediatrics 2003;112:218–32.
3. Preston-Martin S, Pike MC, Ross RK, Jones PA, Henderson BE. Increased cell division as a cause of human cancer. Cancer Res 1990;50:7415–21.
4. Tower RL, Spector LG. The epidemiology of childhood leukemia with a focus on birth weight and diet. Crit Rev Clin Lab Sci 2007;44:203–42.
5. Pombo-de-Oliveira MS, Koifman S. Infant acute leukemia and maternal exposures during pregnancy. Cancer Epidemiol Biomarkers Prev 2006;15:2336–41.
6. Steliarova-Foucher E, Stiller C, Kaatsch P, Berrino F, Coebergh JW, Lacour B, et al. Geographical patterns and time trends of cancer incidence and survival among children and adolescents in Europe since the 1970s (the ACCISproject): an epidemiological study. Lancet 2004;364:2097–105.
7. Wild CP, Kleinjans J. Children and increased susceptibility to environmental carcinogens: evidence or empathy? Cancer Epidemiol Biomarkers Prev 2003;12:1389–94.
8. Gluckman PD, Hanson MA, Cooper C, Thornburg KL. Effect of in utero and early-life conditions on adult health and disease. N Eng J Med 2008;359:61–73.
9. Mose T, Mathiesen L, Karttunen V, Nielsen JK, Sieppi E, Kummu M, et al. Meta-analysis of data from human ex vivo placental perfusion studies on genotoxic and immunotoxic agents within the integrated European project NewGeneris. Placenta 2012;33:433–9.
10. Annola K, Keski-Rahkonen P, Vahakangas K, Lehtonen M. Simultaneous determination of acrylamide, its metabolite glycidamide and antipyrine in human placental perfusion fluid and placental tissue by liquid chromatography-electrospray tandem mass spectrometry. J Chromatogr B Analyt Technol Biomed Life Sci 2008;876:191–7.
11. Balakrishnan B, Henare K, Thorstensen EB, Ponnampalam AP, Mitchell MD. Transfer of bisphenol A across the human placenta. Am J Obstet Gynecol 2010;202:e1–7.
12. Sala M, Ribas-Fitó N, Cardo E, de Muga ME, Marco E, Mazón C, et al. Levels of hexachlorobenzene and other organochlorine compounds in cord blood: exposure across placenta. Chemosphere 2001;43: 895–901.
13. Wang SL, Lin CY, Guo YL, Lin LY, Chou WL, Chang LW. Infant exposure to polychlorinated dibenzo-p-dioxins, dibenzofurans and biphenyls (PCDD/Fs, PCBs)—correlation between prenatal and postnatal exposure. Chemosphere 2004;54:1459–73.
14. Gizzi G, Vincent U, von Holst C, de Jong J, Genouel C. Validation of an analytical method for the determination of carbadox and olaquindox in feedstuff by liquid chromatography coupled to UV and/or diode array detection. Food Addit Contam 2007;24:1226–35.
15. Choi SM, Yoo SD, Lee BM. Toxicological characteristics of endocrine-disrupting chemicals: developmental toxicity, carcinogenicity, and mutagenicity. J Toxicol Environ Health B Crit Rev 2004;7:1–24.
16. Tareke E, Rydberg P, Karlsson P, Eriksson S, Tornqvist M. Analysis of acrylamide, a carcinogen formed in heated foodstuffs. J Agric Food Chem 2002;50:4998–5006.
17. Tornqvist M. Acrylamide in food: the discovery and its implications: a historical perspective. Adv Exp Med Biol 2005;561:1–19.
18. Ghanayem BI, Witt KL, Kissling GE, Tice RR, Recio L. Absence of acrylamide-induced genotoxicity in CYP2E1-null mice: evidence consistent with a glycidamide-mediated effect. Mutat Res 2005;578: 284–97.
19. Besaratinia A, Pfeifer GP. A review of mechanisms of acrylamide carcinogenicity. Carcinogenesis 2007;28:519–28.
20. von Stedingk H, Rydberg P, Tornqvist M. A new modified Edman procedure for analysis of N-terminal valine adducts in hemoglobin by LC-MS/MS. J Chromatogr B Analyt Technol Biomed Life Sci 2010;878: 2483–90.
21. Fenech M. Cytokinesis-block micronucleus cytome assay. Nat Protoc 2007;2:1084–104.
22. Kirsch-Volders M, Elhajouji A, Cundari E, Van Hummelen P. The in vitro micronucleus test: a multi-endpoint assay to detect simultaneously mitotic delay, apoptosis, chromosome breakage, chromosome loss and non-disjunction. Mutat Res 1997;392:19–30.
23. Kirsch-Volders M, Plas G, Elhajouji A, Lukamowicz M, Gonzalez L, Vande Loock K, et al. The in vitro MN assay in 2011: origin and fate, biological significance, protocols, high throughput methodologies and toxicological relevance. Arch Toxicol 2011;85:873–99.
24. Bonassi S, Znaor A, Ceppi M, Lando C, Chang WP, Holland N, et al. An increased micronucleus frequency in peripheral blood lymphocytes predicts the risk of cancer in humans. Carcinogenesis 2007;28: 625–31.
25. Brouwers MM, Besselink H, Bretveld RW, Anzion R, Scheepers PT, Brouwer A, et al. Estrogenic and androgenic activities in total plasma measured with reporter-gene bioassays: relevant exposure measures for endocrine disruptors in epidemiologic studies? Environ Int 2011;37:557–64.
26. Pedersen M, Halldorsson TI, Mathiesen L, Mose T, Brouwer A, Hedegaard M, et al. Dioxin-like exposures and effects on estrogenic and androgenic exposures and micronuclei frequency in mother-newborn pairs. Environ Int 2010;36:344–51.
27. von Stedingk H, Vikström AC, Rydberg P, Pedersen M, Nielsen JK, Segerbäck D, et al. Analysis of hemoglobin adducts from acrylamide, glycidamide, and ethylene oxide in paired mother/cord blood samples from Denmark. Chem Res Toxicol 2011;24:1957–65.
28. Decordier I, Papine A, Plas G, Roesems S, Vande Loock K, Moreno-Palomo J, et al. Automated image analysis of cytokinesis-blocked micronuclei: an adapted protocol and a validated scoring procedure for biomonitoring. Mutagenesis 2009;24(1):85–93.
29. Vande Loock K, Fthenou E, Decordier I, Chalkiadaki G, Keramarou M, Plas G, et al. Maternal and gestational factors and micronuclei frequencies in umbilical blood: the NewGeneris Rhea Cohort in Crete. Environ Health Perspect 2011;119:1460–5.
30. Edgar R, Domrachev M, Lash AE. Gene Expression Omnibus: NCBI gene expression and hybridization array data repository. Nucleic Acids Res 2002;30:207–10.
31. Available at: http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE31836.
32. Boorsma A, Foat BC, Vis D, Klis F, Bussemaker HJ. T-profiler: scoring the activity of predefined groups of genes using gene expression data. Nucleic Acids Res 2005;33:W592–5.
33. Futreal PA, Coin L, Marshall M, Down T, Hubbard T, Wooster R, et al. A census of human cancer genes. Nat Rev Cancer 2004;4:177–83.
34. Morgan JE, Whitlock JP Jr. Transcription-dependent and transcription-independent nucleosome disruption induced by dioxin. Proc Natl Acad Sci U S A 1992;89:11622–6.
35. Puga A, Ma C, Marlowe JL. The aryl hydrocarbon receptor cross-talks with multiple signal transduction pathways. Biochem Pharmacol 2009;77:713–22.
36. Yoshizawa K, Heatherly A, Malarkey DE, Walker NJ, Nyska A. A critical comparison of murine pathology and epidemiological data of TCDD, PCB126, and PeCDF. Toxicol Pathol 2007;35:865–79.

37. Pieters R. Biology and treatment of infant leukemias. In:Pui ChinHon, editor. Treatment of acute leukemias. New directions for Clinical Research. Totowa. Humana Press; 2003: p. 61–75.

38. Mantovani A, Garlanda C, Allavena P. Molecular pathways and targets in cancer-related inflammation. Ann Med 2010;42:161–70.

39. Balkwill F. Tumour necrosis factor and cancer. Nat Rev Cancer 2009;9:361–71.

40. Jost PJ, Ruland J. Aberrant NF-kappaB signaling in lymphoma: mechanisms, consequences, and therapeutic implications. Blood 2007;109:2700–7.

41. Kordes U, Krappmann D, Heissmeyer V, Ludwig WD, Scheidereit C. Transcription factor NF-kappaB is constitutively activated in acute lymphoblastic leukemia cells. Leukemia 2000;14:399–402.

42. Do TN, Ucisik-Akkaya E, Davis CF, Morrison BA, Dorak MT. An intronic polymorphism of IRF4 gene influences gene transcription *in vitro* and shows a risk association with childhood acute lymphoblastic leukemia in males. Biochim Biophys Acta 2010;1802:292–300.

43. Kast RE, Altschuler EL. Anti-apoptosis function of TNF-alpha in chronic lymphocytic leukemia: lessons from Crohn's disease and the therapeutic potential of bupropion to lower TNF-alpha. Arch Immunol Ther Exp (Warsz) 2005;53:143–7.

44. Mullighan CG, Zhang J, Harvey RC, Collins-Underwood Jr, Schulman BA, Phillips LA, et al. JAK mutations in high-risk childhood acute lymphoblastic ##leukemia. Proc Natl Acad Sci U S A 2009;106: 9414–8.

45. Benetatos L, Dasoula A, Syed N, Hatzimichael E, Crook T, Bourantas KL. Methylation analysis of the von Hippel–Lindau gene in acute myeloid leukaemia and myelodysplastic syndromes. Leukemia 2008;22:1293–5.

46. Pierdominici M, Maselli A, Colasanti T, Giammarioli AM, Delunardo F, Vacirca D, et al. Estrogen receptor profiles in human peripheral blood lymphocytes. Immunol Lett 2010;132:79–85.

47. Ramsay AG, Gribben JG. Immune dysfunction in chronic lymphocytic leukemia T cells and lenalidomide as an immunomodulatory drug. Haematologica 2009;94:1198–202.

48. Ansari KI, Mishra BP, Mandal SS. MLL histone methylases in gene expression, hormone signaling and cell cycle. Front Biosci 2009;14: 3483–95.

49. Ross JA, Robison LL. MLL rearrangements in infant leukemia: is there a higher frequency in females? Leuk Res 1997;21:793–5.

50. Heisler LE, Evangelou A, Lew AM, Trachtenberg J, Elsholtz HP, Brown TJ. Androgen-dependent cell cycle arrest and apoptotic death in PC-3 prostatic cell cultures expressing a full-length human androgen receptor. Mol Cell Endocrinol 1997;126:59–73.

51. Holland N, Fucic A, Merlo DF, Sram R, Kirsch-Volders M. Micronuclei in neonates and children: effects of environmental, genetic, demographic and disease variables. Mutagenesis 2011;26:51–6.

52. Milosevic-Djordjevic O, Grujicic D, Arsenijevic S, Brkic M, Ugrinovic S, Marinkovic D. Micronuclei in cord blood lymphocytes as a biomarker of transplacental exposure to environmental pollutants. Tohoku J Exp Med 2007;213:231–9.

53. van Leeuwen DM, Pedersen M, Knudsen LE, Bonassi S, Fenech M, Kleinjans JC, et al. Transcriptomic network analysis of micronuclei-related genes: a case study. Mutagenesis 2011;26:27–32.

Cancer Epidemiology, Biomarkers & Prevention

AACR American Association for Cancer Research

Global Gene Expression Analysis in Cord Blood Reveals Gender-Specific Differences in Response to Carcinogenic Exposure *In Utero*

Kevin Hochstenbach, Danitsja M. van Leeuwen, Hans Gmuender, et al.

Updated version	Access the most recent version of this article at: doi:10.1158/1055-9965.EPI-12-0304
Supplementary Material	Access the most recent supplemental material at: http://cebp.aacrjournals.org/content/suppl/2012/08/09/1055-9965.EPI-12-0304.DC1

Cited articles	This article cites 51 articles, 8 of which you can access for free at: http://cebp.aacrjournals.org/content/21/10/1756.full#ref-list-1
Citing articles	This article has been cited by 3 HighWire-hosted articles. Access the articles at: http://cebp.aacrjournals.org/content/21/10/1756.full#related-urls

E-mail alerts	Sign up to receive free email-alerts related to this article or journal.
Reprints and Subscriptions	To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.
Permissions	To request permission to re-use all or part of this article, use this link http://cebp.aacrjournals.org/content/21/10/1756. Click on "Request Permissions" which will take you to the Copyright Clearance Center's (CCC) Rightslink site.