

Null Results in Brief

No Association between *Matrix Metalloproteinase (MMP)-1, MMP-3, and MMP-7* SNPs and Endometrial Cancer Risk

Alicia Beeghly-Fadiel,¹ Yong-Bing Xiang,² Sandra L. Deming,¹ Ji-Rong Long,¹ Wang-Hong Xu,³ Qiuyin Cai,¹ Wei Zheng,¹ and Xiao Ou Shu¹

¹Vanderbilt University Medical Center, Division of Epidemiology, Nashville, Tennessee; and ²Shanghai Cancer Institute, Department of Epidemiology; and ³Fudan University School of Public Health, Department of Epidemiology, Shanghai, China

Introduction

The matrix metalloproteinases (MMP) function as regulators of the dynamic tissue remodeling that occurs in the endometrial lining of the uterus during the normal human menstrual cycle; dysregulation of the MMPs is thought to contribute to the development of both endometriosis and endometrial cancer (1). MMP-1 expression was found to be significantly higher in endometriotic lesions than in surrounding endometrium (2, 3), whereas MMP-3 levels have been reported to be both lower (4) and higher (5) in women with endometriosis compared with women without. MMP-7 expression was found to be significantly higher in endometrial hyperplasia and adenocarcinomas than normal endometrium (6) and, further, was associated with higher grade endometrial tumors (7), and both myometrial (6) and lymph node invasion (8). These three MMP genes are located on the negative strand of chromosome 11, and functional polymorphisms that influence their respective transcription levels have been identified for each (9-12). However, previous studies on MMP SNPs and endometrial cancer are sparse; two studies were found to have evaluated a single MMP-1 single nucleotide polymorphism (SNP) and results were inconsistent. Therefore, this comprehensive study of individual genetic variation across MMP-1, MMP-3, and MMP-7 was undertaken to evaluate associations with endometrial cancer susceptibility.

Materials and Methods

The Shanghai Endometrial Cancer Study (SECS) is a large, population-based case-control study that has been previously described (13, 14). Briefly, cases were women

diagnosed with endometrial cancer between January 1997 and December 2003, ages 30 to 69 y, identified from the Shanghai Cancer Registry. Controls were randomly selected from the Shanghai Resident Registry and frequency matched to cases in 5-year intervals. Of 1,458 identified eligible cases, in-person interviews were completed for 1,204 (82.6%). Reasons for nonparticipation included refusal ($n = 137$, 9.4%), death before interview ($n = 66$, 4.5%), inability to be located ($n = 37$, 2.5%), and health or communication problems ($n = 14$, 1.0%). Of eligible controls identified (1,629), in-person interviews were completed for 1,212 (74.4%). Reasons for nonparticipation included refusal ($n = 340$, 20.9%), absence during the study period ($n = 61$, 3.7%), and health or communication problems ($n = 16$, 1.1%). Institutional review board approval was granted by relevant institutions in both China and the United States. Informed consent was obtained from each included participant. DNA samples were provided and available for 87.3% of cases ($n = 1,052$) and 87.3% ($n = 1,058$) of controls.

Haplotype-tagging SNPs were selected from Han Chinese data from the HapMap Project (15) using the Tagger program (16) to capture SNPs with a minimum minor allele frequency (MAF) of 0.05 in either MMP-1, MMP-3, or MMP-7 (± 5 kb) with an r^2 of 0.90 or greater. Known or potentially functional SNPs were forced into the haplotype-tagging SNP selection process. For MMP-1, 17 SNPs were selected, with 14 successfully genotyped. For MMP-3, seven SNPs were selected, with six successfully genotyped. For MMP-7, 12 SNPs were selected, with 11 successfully genotyped. Genotyping was conducted using the Affymetrix Targeted Genotyping System (Affymetrix; ref. 17) for 1,037 cases (98.6%) and 1,018 controls (96.2%).

Hardy-Weinberg equilibrium was applied to test the observed and expected genotype frequencies for cases and controls (χ^2 test). Associations between SNPs and covariates were evaluated with the χ^2 test or t test when appropriate. Covariates considered included age at diagnosis, education, age at menarche, age at menopause among postmenopausal women, menopausal status, number of pregnancies, oral contraceptive use, body mass index, waist-to-hip ratio (WHR), physical activity in the preceding decade, and first-degree family history of breast, colorectal, or endometrial cancer. Odds ratios and

Cancer Epidemiol Biomarkers Prev 2009;18(6):1925-8

Received 3/16/09; accepted 3/27/09; published OnlineFirst 5/12/09.

Grant support: The Vanderbilt Microarray Shared Resource is supported by the Vanderbilt-Ingram Cancer Center (P30 CA68485), the Vanderbilt Diabetes Research and Training Center (P60 DK20593), the Vanderbilt Digestive Disease Center (P30 DK58404), and the Vanderbilt Vision Center (P30 EY08126). This study would not have been possible without the support of the study participants and research staff of the Shanghai Endometrial Cancer Study. This work was supported by United States Public Health Service grant R01 CA92585 from the National Cancer Institute.

Requests for reprints: Xiao-Ou Shu, Vanderbilt Epidemiology Center, Vanderbilt University Medical Center, 2525 West End Avenue, Suite 600, IMPH, Nashville, TN 37203-1738. Phone: 615-936-0713; Fax: 615-936-8291. E-mail: Xiao-ou.shu@vanderbilt.edu

Copyright © 2009 American Association for Cancer Research.

doi:10.1158/1055-9965.EPI-09-0244

corresponding 95% confidence intervals were determined by logistic regression using additive models that included adjustment for age and education. Dominant and recessive models were additionally used when appropriate. Linkage disequilibrium was assessed by Haploview (18). All statistical tests were two-tailed, and *P* values were considered to be statistically significant when ≤ 0.05 .

Results

Consistent with previous SECS analyses (13, 14) and other epidemiologic studies, cases, and controls included in the current study differed with regard to age at menarche, age at menopause, menopausal status, number of pregnancies, use of oral contraceptives, body mass index and WHR, physical activity, and first-degree family history of cancer (data not shown). SNPs included in this study are listed in Table 1; their order corresponds to the open reading frames of the genes on the negative strand of chromosome 11. Of the 31 polymorphisms genotyped, one was found not to be polymorphic in this study population (*MMP-7 rs11568819*) and thus was not included in our analyses. No SNPs were found to deviate from Hardy-Weinberg equilibrium. Associations with endometrial cancer risk

were calculated in additive effect models that included adjustment for age and education; further adjustment for body mass index, number of pregnancies, menopausal status, or family history of cancer did not appreciably alter the effect estimates. No significant associations were observed. Two *MMP-7* SNPs, *rs17098318* and *rs11568818*, both tended to confer an increased, but nonsignificant, risk of endometrial cancer for homozygotes, in both additive and recessive models. Furthermore, no SNPs were found to have effects that significantly differed by menopausal status. The linkage disequilibrium structure of these 30 SNPs is shown in Fig. 1, and includes six haplotype blocks. Similar to single SNP analysis, no significant effects were observed in haplotype analysis of these MMP polymorphisms (data not shown).

Discussion

Promoter polymorphisms in the *MMP-1*, *MMP-3*, and *MMP-7* genes have been associated with altered susceptibility to cancer in human populations (17, 19-30), although studies on endometrial cancer risk and MMP SNPs have been lacking. Two small studies evaluating *MMP-1 -1607 1G/2G (rs1799750)* and *MMP-3 -1171*

Table 1. MMP-3, MMP-1, and MMP-7 SNPs and endometrial cancer risk, evaluated among 1,037 cases and 1,018 controls, the SECS

Gene, SNP	Region	Alleles*	MAF [†]	HWE <i>p</i> [§]	Endometrial cancer risk, OR (95% CI) [‡]		
					AB	BB	<i>P</i>
MMP-3							
rs645419	Promoter	G/A	32.6%	0.573	1.1 (0.9-1.3)	1.0 (0.8-1.4)	0.628
rs632478	Promoter	C/A	33.0%	0.766	1.1 (0.9-1.3)	1.0 (0.8-1.4)	0.717
rs522616	Promoter	A/G	36.2%	0.700	1.0 (0.8-1.2)	1.2 (0.9-1.6)	0.321
rs679620	Exon 2	G/A	32.9%	0.754	1.1 (0.9-1.3)	1.0 (0.8-1.4)	0.765
rs650108	Intron 8	A/G	40.2%	0.905	1.0 (0.9-1.3)	1.1 (0.8-1.4)	0.624
rs655403	Intron 8	C/T	7.2%	0.552	1.0 (0.8-1.3)	1.5 (0.4-5.3)	0.832
MMP-1							
rs484915	Promoter	A/T	34.2%	0.311	1.0 (0.8-1.2)	0.9 (0.7-1.3)	0.777
rs1155764	Promoter	T/G	22.0%	0.765	0.9 (0.7-1.0)	1.1 (0.7-1.7)	0.397
rs509332	Promoter	A/G	13.4%	0.124	1.1 (0.9-1.4)	0.8 (0.4-1.5)	0.639
rs470206	Promoter	G/A	13.4%	0.114	1.1 (0.9-1.4)	0.8 (0.5-1.5)	0.672
rs2075847	Promoter	T/C	24.0%	0.253	1.1 (0.9-1.3)	1.2 (0.8-1.6)	0.369
rs498186	Promoter	A/C	44.0%	0.966	1.0 (0.8-1.2)	0.9 (0.7-1.2)	0.507
rs475007	Promoter	T/A	34.0%	0.702	0.9 (0.8-1.1)	1.1 (0.9-1.5)	0.626
rs996999	Intron 4	C/T	49.1%	0.291	1.1 (0.9-1.3)	1.0 (0.7-1.2)	0.731
rs470558	Exon 5	G/A	12.4%	0.642	0.9 (0.7-1.1)	1.0 (0.5-2.2)	0.421
rs7125062	Intron 6	C/T	30.6%	0.478	1.0 (0.8-1.2)	0.8 (0.6-1.1)	0.472
rs1938901	Intron 8	T/C	42.4%	0.691	1.1 (0.9-1.3)	1.1 (0.9-1.4)	0.440
rs2071231	Intron 9	T/G	20.4%	0.607	1.1 (0.9-1.3)	1.2 (0.8-1.8)	0.173
rs7945189	3' FR	C/T	8.8%	0.250	0.9 (0.7-1.1)	1.3 (0.4-4.3)	0.411
rs1470504	3' FR	G/A	13.9%	0.663	0.9 (0.7-1.1)	0.9 (0.4-1.7)	0.328
MMP-7							
rs880197	Promoter	A/T	38.7%	0.755	0.9 (0.7-1.1)	0.9 (0.7-1.1)	0.230
rs17098318	Promoter	G/A	7.9%	0.309	1.0 (0.8-1.2)	2.2 (0.7-7.2)	0.782
rs11568818	Promoter	A/G	8.0%	0.281	1.0 (0.7-1.2)	2.0 (0.6-6.5)	0.970
rs11225307	Intron 3	A/G	26.5%	0.461	1.1 (0.9-1.3)	1.1 (0.8-1.6)	0.325
rs17352054	Intron 5	A/C	12.0%	0.682	1.2 (1.0-1.5)	1.1 (0.6-2.3)	0.112
rs495041	3' FR	C/T	49.5%	0.508	0.9 (0.8-1.2)	1.0 (0.8-1.2)	0.741
rs10895304	3' FR	A/G	24.5%	0.853	1.0 (0.8-1.2)	1.0 (0.7-1.5)	0.865
rs7935378	3' FR	T/C	23.0%	0.941	0.9 (0.8-1.1)	1.1 (0.7-1.6)	0.655
rs12184413	3' FR	C/T	29.5%	0.577	0.9 (0.8-1.1)	1.0 (0.7-1.4)	0.636
rs11225297	3' FR	A/T	20.7%	0.629	1.0 (0.8-1.2)	0.9 (0.5-1.4)	0.572

Abbreviations: OR, odds ratio; CI, confidence interval.

*Major and minor alleles as determined by the distribution among SECS controls.

[†]Minor Allele Frequency (MAF) among SECS controls.

[‡]Odds ratio and 95% confidence interval for the risk of endometrial cancer, age and education adjusted; AA, major allele homozygous; BB, minor allele homozygous, AB heterozygous; *P* value for trend.

[§]Hardy-Weinberg equilibrium test, *P* value among SECS controls.

^{||}3' Flanking region, downstream of the coding region.

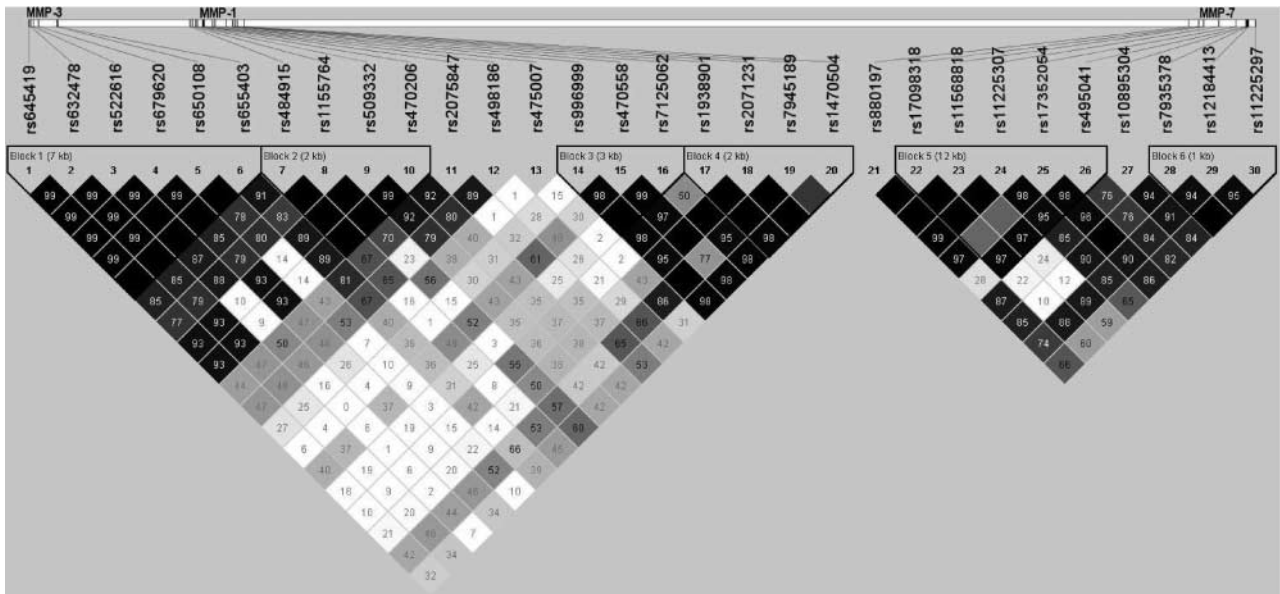


Figure 1. Linkage disequilibrium structure of 30 MMP-3, MMP-1, and MMP-7 SNPs, on chromosome 11 among 1,018 controls from the SECS; value shown in D' .

5A/6A (*rs35068180* and *rs3025059*) and endometriosis found mixed results. No association for either SNP was seen among 56 cases and 71 controls (31), whereas the *MMP-1* 2G allele was found to confer an increased risk of endometriosis among 100 cases and 150 controls (32). Similarly, the *MMP-1* -1607 2G allele was found to confer an increased risk of endometrial adenocarcinoma among 100 cases and 150 controls (33), whereas no difference was seen between 107 cases and 213 controls (34). Unfortunately, neither of these functional SNPs were genotyped in the current study. However, *MMP-3* *rs679620* was genotyped and shares moderate linkage disequilibrium with *MMP-1* *rs1799750* ($D = 0.79$; $r^2 = 0.60$; ref. 35); no association with endometrial cancer risk was observed. To our knowledge, no previous studies of *MMP-3* or *MMP-7* SNPs and endometrial cancer risk have been conducted. In this study, both of the functional promoter *MMP-7* SNPs were genotyped. Although *MMP-7* -153 C/T (*rs11568819*) was not found to be polymorphic in this population, *MMP-7* -181 A/G (*rs11568818*) and another promoter SNP in high linkage disequilibrium (*rs17098318*; $D = 1.0$; $r^2 = 0.99$) both seemed to confer an increased risk of endometrial cancer in homozygote carriers of the rare allele. This is similar to our findings for breast cancer risk among premenopausal women (17), and may indicate a real, but low prevalence association that the current study lacked adequate power to detect under recessive models. Given the size of our study population, this analysis had only 31% power to detect a recessive effect of an odds ratio of 2.0 for a gene with a MAF of only 8%. However, for additive associations, we had >92% power to detect an odds ratio of 1.4 for a SNP with a MAF of 10%, >93% power to detect an odds ratio of 1.3 for a SNP with a MAF of 20%, and >77% power to detect an odds ratio of 1.2 for a SNP with a MAF of 30%. In summary, 30 haplotype tagging polymorphisms in *MMP-1*, *MMP-3*, and *MMP-7* were evaluated among 1,037 endometrial cancer cases and 1,018 controls; none were found to be significantly associated with endometrial cancer risk.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

Acknowledgments

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked *advertisement* in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

We thank Dr. Fan Jin for her contributions to implementing the study in Shanghai; Regina Courtney, Qing Wang, Dr. Shawn Levy, and the Vanderbilt Microarray Shared Resource for their contributions to the genotyping; and Brandy Venuti for her assistance in the preparation of this manuscript.

References

- Curry TE, Jr., Osteen KG. The matrix metalloproteinase system: changes, regulation, and impact throughout the ovarian and uterine reproductive cycle. *Endocr Rev* 2003;24:428–65.
- Hudelist G, Lass H, Keckstein J, et al. Interleukin 1 α and tissue-lytic matrix metalloproteinase-1 are elevated in ectopic endometrium of patients with endometriosis. *Hum Reprod* 2005;20:1695–701.
- Hudelist G, Keckstein J, Czerwenka K, et al. Estrogen receptor β and matrix metalloproteinase 1 are coexpressed in uterine endometrium and endometriotic lesions of patients with endometriosis. *Fertil Steril* 2005;84:1249–56.
- Uzan C, Cortez A, Dufournet C, Fauvet R, Siffroi JP, Darai E. Eutopic endometrium and peritoneal, ovarian and bowel endometriotic tissues express a different profile of matrix metalloproteinases-2, -3 and -11, and of tissue inhibitor metalloproteinases-1 and -2. *Virchows Arch* 2004;445:603–9.
- Gilbert-Estelles J, Estelles A, Gilbert J, et al. Expression of several components of the plasminogen activator and matrix metalloproteinase systems in endometriosis. *Hum Reprod* 2003;18:1516–22.
- Obokata A, Watanabe J, Nishimura Y, Arai T, Kawaguchi M, Kuramoto H. Significance of matrix metalloproteinase-7 [correction of matrix metalloproteinase-2], -11 and tissue inhibitor of metalloproteinase-1 expression in normal, hyperplastic and neoplastic endometrium. *Anticancer Res* 2007;27:95–105.
- Misugi F, Sumi T, Okamoto E, et al. Expression of matrix metalloproteinases and tissue inhibitors of metalloproteinase in uterine endometrial carcinoma and a correlation between expression of matrix metalloproteinase-7 and prognosis. *Int J Mol Med* 2005;16:541–6.
- Ueno H, Yamashita K, Azumano I, Inoue M, Okada Y. Enhanced

- production and activation of matrix metalloproteinase-7 (matrilysin) in human endometrial carcinomas. *Int J Cancer* 1999;84:470-7.
9. Ye S, Watts GF, Mandalia S, Humphries SE, Henney AM. Preliminary report: genetic variation in the human stromelysin promoter is associated with progression of coronary atherosclerosis. *Br Heart J* 1995;73:209-15.
 10. Ye S, Eriksson P, Hamsten A, Kurkinen M, Humphries SE, Henney AM. Progression of coronary atherosclerosis is associated with a common genetic variant of the human stromelysin-1 promoter which results in reduced gene expression. *J Biol Chem* 1996;271:13055-60.
 11. Rutter JL, Mitchell TI, Buttice G, et al. A single nucleotide polymorphism in the matrix metalloproteinase-1 promoter creates an Ets binding site and augments transcription. *Cancer Res* 1998;58:5321-5.
 12. Jormsjo S, Whatling C, Walter DH, Zeiher AM, Hamsten A, Eriksson P. Allele-specific regulation of matrix metalloproteinase-7 promoter activity is associated with coronary artery luminal dimensions among hypercholesterolemic patients. *Arterioscler Thromb Vasc Biol* 2001;21:1834-9.
 13. Xu WH, Dai Q, Xiang YB, et al. Interaction of soy food and tea consumption with CYP19A1 genetic polymorphisms in the development of endometrial cancer. *Am J Epidemiol* 2007;166:1420-30.
 14. Deming SL, Zheng W, Xu WH, et al. UGT1A1 genetic polymorphisms, endogenous estrogen exposure, soy food intake, and endometrial cancer risk. *Cancer Epidemiol Biomarkers Prev* 2008;17:563-70.
 15. The International HapMap Project. *Nature* 2003;426:789-96.
 16. de Bakker PIW, McVean G, Sabeti PC, et al. A high-resolution HLA and SNP haplotype map for disease association studies in the extended human MHC. *Nat Genet* 2006;38:1166-72.
 17. Beeghly-Fadiel A, Long JR, Gao YT, et al. Common MMP-7 polymorphisms and breast cancer susceptibility: a multistage study of association and functionality. *Cancer Res* 2008;68:6453-9.
 18. Barrett JC, Fry B, Maller J, Daly MJ. Haploview: analysis and visualization of LD and haplotype maps. *Bioinformatics* 2005;21:263-5.
 19. Hashimoto T, Uchida K, Okayama N, et al. Association of matrix metalloproteinase (MMP)-1 promoter polymorphism with head and neck squamous cell carcinoma. *Cancer Lett* 2004;211:19-24.
 20. Hirata H, Okayama N, Naito K, et al. Association of a haplotype of matrix metalloproteinase (MMP)-1 and MMP-3 polymorphisms with renal cell carcinoma. *Carcinogenesis* 2004;25:2379-84.
 21. Zinzindohoue F, Blons H, Hans S, et al. Single nucleotide polymorphisms in MMP1 and MMP3 gene promoters as risk factor in head and neck squamous cell carcinoma. *Anticancer Res* 2004;24:2021-6.
 22. Lievre A, Milet J, Carayol J, et al. Genetic polymorphisms of MMP1, MMP3 and MMP7 gene promoter and risk of colorectal adenoma. *BMC Cancer* 2006;6:270.
 23. O-charoenrat P, Leksriskul P, Sangruchi S. A functional polymorphism in the matrix metalloproteinase-1 gene promoter is associated with susceptibility and aggressiveness of head and neck cancer. *Int J Cancer* 2006;118:2548-53.
 24. Lu Z, Cao Y, Wang Y, et al. Polymorphisms in the matrix metalloproteinase-1, 3, and 9 promoters and susceptibility to adult astrocytoma in northern China. *J Neurooncol* 2007;85:65-73.
 25. Nasr HB, Mestiri S, Chahed K, et al. Matrix metalloproteinase-1 (-1607) 1G/2G and -9 (-1562) C/T promoter polymorphisms: susceptibility and prognostic implications in nasopharyngeal carcinomas. *Clin Chim Acta* 2007;384:57-63.
 26. Nishizawa R, Nagata M, Noman A, et al. The 2G allele of promoter region of Matrix metalloproteinase-1 as an essential pre-condition for the early onset of oral squamous cell carcinoma. *BMC Cancer* 2007;7:187.
 27. Woo M, Park K, Nam J, Kim JC. Clinical implications of matrix metalloproteinase-1, -3, -7, -9, -12, and plasminogen activator inhibitor-1 gene polymorphisms in colorectal cancer. *J Gastroenterol Hepatol* 2007;22:1064-70.
 28. Sauter W, Rosenberger A, Beckmann L, et al. Matrix metalloproteinase 1 (MMP1) is associated with early-onset lung cancer. *Cancer Epidemiol Biomarkers Prev* 2008;17:1127-35.
 29. Singh H, Jain M, Mittal B. MMP-7 (-181A>G) promoter polymorphisms and risk for cervical cancer. *Gynecol Oncol* 2008;110:71-5.
 30. Sugimoto M, Furuta T, Kodaira C, et al. Polymorphisms of matrix metalloproteinase-7 and chymase are associated with susceptibility to and progression of gastric cancer in Japan. *J Gastroenterol* 2008;43:751-61.
 31. Ferrari MM, Biondi ML, Rossi G, et al. Analysis of two polymorphisms in the promoter region of matrix metalloproteinase 1 and 3 genes in women with endometriosis. *Acta Obstet Gynecol Scand* 2006;85:212-7.
 32. Shan K, Ying W, Jian-Hui Z, Wei G, Na W, Yan L. The function of the SNP in the MMP1 and MMP3 promoter in susceptibility to endometriosis in China. *Mol Hum Reprod* 2005;11:423-7.
 33. Nishioka Y, Kobayashi K, Sagae S, et al. A single nucleotide polymorphism in the matrix metalloproteinase-1 promoter in endometrial carcinomas. *Jpn J Cancer Res* 2000;91:612-5.
 34. Sugimoto M, Yoshida S, Kennedy S, Deguchi M, Ohara N, Maruo T. Matrix metalloproteinase-1 and -9 promoter polymorphisms and endometrial carcinoma risk in a Japanese population. *J Soc Gynecol Investig* 2006;13:523-9.
 35. Beeghly-Fadiel A, Cai Q, Lu W, et al. No Association between MMP-1 or MMP-3 Polymorphisms and Breast Cancer Susceptibility: A report from the Shanghai Breast Cancer Study. *Cancer Epidemiology Biomarkers Prev* 2009;18:1324-7.

Cancer Epidemiology, Biomarkers & Prevention

AACR American Association
for Cancer Research

No Association between *Matrix Metalloproteinase (MMP)-1, MMP-3* , and *MMP-7* SNPs and Endometrial Cancer Risk

Alicia Beeghly-Fadiel, Yong-Bing Xiang, Sandra L. Deming, et al.

Cancer Epidemiol Biomarkers Prev 2009;18:1925-1928.

Updated version Access the most recent version of this article at:
<http://cebp.aacrjournals.org/content/18/6/1925>

Cited articles This article cites 35 articles, 10 of which you can access for free at:
<http://cebp.aacrjournals.org/content/18/6/1925.full#ref-list-1>

Citing articles This article has been cited by 1 HighWire-hosted articles. Access the articles at:
<http://cebp.aacrjournals.org/content/18/6/1925.full#related-urls>

E-mail alerts [Sign up to receive free email-alerts](#) related to this article or journal.

Reprints and Subscriptions To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions To request permission to re-use all or part of this article, use this link
<http://cebp.aacrjournals.org/content/18/6/1925>.
Click on "Request Permissions" which will take you to the Copyright Clearance Center's (CCC) Rightslink site.