

Review

Physical Activity and Postmenopausal Breast Cancer: Proposed Biologic Mechanisms and Areas for Future Research

Heather K. Neilson,¹ Christine M. Friedenreich,¹ Nigel T. Brockton,¹ and Robert C. Millikan²

¹Division of Population Health and Information, Alberta Cancer Board, Calgary, Canada; and ²Department of Epidemiology and Linberger Comprehensive Cancer Center, University of North Carolina, Chapel Hill, North Carolina

Abstract

Convincing evidence now supports a probable preventive role for physical activity in postmenopausal breast cancer. The mechanisms by which long-term physical activity affect risk, however, remain unclear. The aims of this review were to propose a biological model whereby long-term physical activity lowers postmenopausal breast cancer risk and to highlight gaps in the epidemiologic literature. To address the second aim, we summarized epidemiologic literature on 10 proposed biomarkers, namely, body mass index (BMI), estrogens, androgens, sex hormone binding globulin, leptin, adiponectin, markers of insulin resistance, tumor necrosis factor- α , interleukin-6, and C-reactive protein, in relation to postmenopausal breast cancer risk and physical activity, respectively. Associations were deemed "convincing," "probable," "possible," or "hypothesized" using set criteria. Our proposed biological model illustrated the co-occurrence of overweight/obesity, insulin resistance, and chronic inflammation influencing can-

cer risk through interrelated mechanisms. The most convincing epidemiologic evidence supported associations between postmenopausal breast cancer risk and BMI, estrogens, and androgens, respectively. In relation to physical activity, associations were most convincing for BMI, estrone, insulin resistance, and C-reactive protein. Only BMI and estrone were convincingly (or probably) associated with both postmenopausal breast cancer risk and physical activity. There is a need for prospective cohort studies relating the proposed biomarkers to cancer risk and for long-term exercise randomized controlled trials comparing biomarker changes over time, specifically in postmenopausal women. Future etiologic studies should consider interactions among biomarkers, whereas exercise trials should explore exercise effects independently of weight loss, different exercise prescriptions, and effects on central adiposity. (Cancer Epidemiol Biomarkers Prev 2009;18(1):11–27)

Introduction

In 2004 the lifetime probability of developing invasive breast cancer for women in Canada was 11% or 1 in 9, more than any other cancer in women aside from nonmelanoma skin cancer (1). In 2004 in the United States, invasive breast cancer was the most commonly diagnosed cancer in women (age-adjusted incidence rate at 118 per 100,000 women; ref. 2). Although several risk factors are proposed for breast cancer, low levels of physical activity may be one of the most modifiable. Excess body weight and low physical activity together may account for one quarter to one third of all breast cancer cases (3). From the substantial epidemiologic literature on physical activity and breast cancer (3–6), convincing evidence now supports a "probable" preventive role for physical activity in postmenopausal women

(4), whereby habitual activity may lower risk by approximately 20% (7). The evidence in premenopausal women has been generally weaker (4, 5). The mechanisms by which long-term physical activity lowers postmenopausal breast cancer risk, however, remain unclear. Mechanistic insight, ideally from biomarker studies (8), would add biological plausibility to the association, guide future epidemiologic research, identify new targets for interventions, and inform public health recommendations for lowering breast cancer risk.

The aims of our review were to propose a biological model whereby long-term physical activity lowers postmenopausal breast cancer risk and to highlight gaps in the epidemiologic literature. To address the second aim, we summarized the existing epidemiologic literature on the following 10 biologically plausible, candidate biomarkers (anthropometric and blood), in relation to postmenopausal breast cancer risk and physical activity, respectively: body weight or body mass index (BMI), estrogens, androgens, sex hormone binding globulin (SHBG), leptin, adiponectin, markers of insulin resistance (i.e. insulin, glucose, C-peptide, and glycosylated hemoglobin), tumor necrosis factor α (TNF- α), interleukin-6 (IL-6), and C-reactive protein (CRP). Although many

Received 8/15/08; revised 10/5/08; accepted 10/27/08.

Grant support: CMF is supported by an Alberta Heritage Foundation for Medical Research Health Senior Scholar Award. R.C. Millikan is supported by the Specialized Program of Research Excellence in Breast Cancer NIH Grant P50-CA58223.

Requests for reprints: Christine Friedenreich, Division of Population Health, Alberta Cancer Board, 1331-29 Street NW, Calgary, Alberta, Canada T2N 4N2. Phone: 403-521-3841; Fax: 403-270-8003. E-mail: chrisf@cancerboard.ab.ca

Copyright © 2009 American Association for Cancer Research.

doi:10.1158/1055-9965.EPI-08-0756

candidate biomarkers exist (9-14), the plausibility of our chosen biomarkers has been discussed in recent literature (9, 13, 15) and their responses to exercise have been tested (16-20) or are currently under investigation (9) in various exercise intervention trials for postmenopausal breast cancer prevention. Insulin-like growth factor-1 was not included in our review because most previous studies in older women have not shown decreased insulin-like growth factor-1 with increasing physical activity (20, 21). Moreover, at least two reviews of the epidemiologic literature failed to show significantly altered breast cancer risk in postmenopausal women with higher insulin-like growth factor-1 levels (22, 23). Likewise, we excluded mammographic density from our biological model because recent research has generally shown no association between physical activity and breast density (24-30).

Materials and Methods

In February 2008 we searched the published literature using PubMed (NIH). To identify studies we queried medical subject headings (MeSH) for each hypothesized biomarker combined with terms for breast cancer (MeSH term "Breast Neoplasms") and physical activity ("physical activity" or MeSH terms "Motor Activity" or "Exercise"), respectively. Hypothesized biomarker information was retrieved using the following MeSH terms: "Estrogens," "Androgens," "Sex-hormone binding globulin," "C-reactive protein," "Leptin," "Adiponectin," "Interleukin-6," "Tumor necrosis factor-alpha," "Insulin," "Insulin resistance," "Glucose," "C-peptide," "Hemoglobin A, Glycosylated," "Hyperinsulinism," "Body weights and measures," "Body composition," and "Body weight." We limited our electronic search to English language publications in humans. Due to the extensive literature on physical activity and insulin resistance, physical activity and body weight, and breast cancer and body weight, we restricted the latter three searches to English language review articles and meta-analyses in adults. With respect to the remaining topics, we reviewed original articles reporting results explicitly for postmenopausal women. We excluded intervention trials in cancer survivors and studies in diseased women (unless they were type 2 diabetics, whom we included), trained athletes, or the severely obese. Studies presenting results only for women on hormone replacement therapy were also excluded. Furthermore, if postmenopausal status was ambiguous or results were not stratified by menopausal status, then the article was excluded. If menopausal status was not stated but the minimum age of participants was ≥ 55 y, however, the study was included. Acute exercise trials (<4 wk duration) or trials intervening in both exercise and diet were similarly excluded from our review in order to isolate the effects of long-term physical activity from weight loss. The review was not restricted to any particular type or intensity of physical activity.

We classified the epidemiologic evidence using a scheme adapted from the American Institute of Cancer Research/World Cancer Research Fund's recent comprehensive report on physical activity and cancer prevention (4). Unlike the report, however, we did not assess study quality or study heterogeneity within or among study

types and we believed all of the associations to be biologically plausible. Furthermore, we did not assess physical activity methods or adjustment for potential confounders in the individual studies as these assessments were beyond the scope of this review. An association was deemed "convincing" if it was supported by at least two cohort studies or trials (i.e., randomized or nonrandomized) and the expected association was found consistently across all analyses, or if the association was generally supported by a large body of epidemiologic literature and/or public health guidelines. An association was considered "probable" if two or more cohort studies or trials were conducted and most of the analyses supported the same expected association. "Possible" described associations based on two or more cohort studies or trials or five or more case-control, nested case-control, or case-cohort studies and/or 50% of the analyses or less supporting the same expected association. An association was "hypothesized" if supported by a limited number of studies and/or very few analyses, if any, showed the expected association. We summarized each study's findings very simply in terms of "positive" or "negative" based on the direction of the association and whether or not the adjusted results reached statistical significance ($P \leq 0.05$).

Results

Body Weight Measures. A strong biological rationale and wealth of epidemiologic evidence now support a role for elevated body weight in increasing postmenopausal breast cancer risk. Overweight and obesity could lead to cancer through a number of pathways, including higher levels of circulating sex hormones, insulin resistance, chronic inflammation, and/or lower levels of SHBG and adiponectin (Table 1), although the exact mechanisms are unknown.

As expected, our search of the review literature supported a convincing positive association between postmenopausal breast cancer risk and elevated BMI (Table 2). A 2007 review of 24 cohort studies and 56 case-control studies in postmenopausal women (4) provided strong evidence of increased risk with increasing BMI based on consistent findings and clear dose-response relations. Similarly, a 2008 meta-analysis of 31 prospective studies found a 12% increase in risk of postmenopausal breast cancer for every 5 kg/m² increase in BMI [relative risk, 1.12; 95% confidence interval (95% CI), 1.08-1.16; ref. 31]. In general, earlier reviews reported higher risk with adult weight gain (32, 33) and increasing weight and/or BMI for postmenopausal women (3, 33-36). A 2001 meta-analysis of 13 studies in postmenopausal women showed a significantly increased risk of breast cancer by 2% per 1 kg/m² increase in BMI (37). Subsequently, a 2003 pooled analysis of eight prospective studies of postmenopausal women (38) found the relative risk of breast cancer to be 1.19 (95% CI, 1.05-1.34) for every 5 kg/m² increase in BMI. However, when adjusted for free estradiol, the risk approached unity (relative risk, 1.02; 95% CI, 0.89-1.17). In the same analysis, the mean concentration of estrogen metabolites in obese women (BMI ≥ 30.0) was between 60% and 219% higher than in thin women (BMI <22.5), showing the important, presumably causal path from body weight to circulating estrogen levels (Table 1).

Table 1. Possible mechanisms relating commonly proposed biomarkers to postmenopausal breast cancer risk and physical activity

Proposed biomarker	Possible role in postmenopausal breast cancer	Possible impact of physical activity in postmenopausal women
Estrogens	Adipose tissue contains higher levels of aromatase which converts androgens to estrogens (57). Estrogens are mitogens in the breast, stimulating mammary cell proliferation through estrogen receptor-mediated transcriptional activity and by activation of intracellular signaling pathways (48, 57).	Reduces adiposity thereby lowering the capacity for conversion of androgens to estrogens by aromatase and lowering circulating estrogen levels (10, 221). Reduces insulin levels thereby increasing SHBG levels (73-75) which may decrease estradiol bioavailability.
Androgens	Testosterone and androstenedione may increase risk upon conversion to estradiol and estrone, respectively, in adipose tissue (58, 222). Androgens also act directly on breast cells by binding to the androgen receptor (65), a ligand-dependent transcription factor expressed in the majority of breast cancers (223, 224). There may be synergy between estrogens and androgens in increasing breast cancer risk (69). The exact mechanism in postmenopausal breast cancer is unclear (69, 225). <i>In vitro</i> , androgens can directly stimulate or inhibit breast cancer cell proliferation depending on the cell line (69, 226, 227). Some clinical data support a protective role for androgens in breast cancer (227).	Decreases testosterone levels through loss of body fat. Decreased adiposity lowers levels of 17 β -hydroxysteroid dehydrogenase enzyme which converts androstenedione to testosterone in s.c. and intra-abdominal fat (228), thereby lowering testosterone levels. Reduces adiposity thereby lowering the capacity for conversion of androgens to estrogens by aromatase and lowering circulating estrogen levels (10, 221). Reduces insulin levels thereby increasing SHBG levels (73-75) which may decrease testosterone bioavailability.
SHBG	SHBG binds to estradiol and testosterone (55) thereby reducing their bioavailabilities; may act as a negative modulator of estradiol (54).	Reduces insulin levels which increases circulating SHBG (73, 229), thereby decreasing the bioavailabilities of estradiol and testosterone.
Insulin Resistance	Hyperinsulinemia is associated with decreased plasma SHBG (73, 229), thereby increasing sex hormone bioavailability. Insulin exerts mitogenic effects in breast cancer cells <i>in vitro</i> ; (230, 231) may synergize with estrogen (57, 232). Insulin resistance and hyperinsulinemia are strongly related to obesity,(233) and particularly intra-abdominal fat (73, 234). Insulin resistance has been associated with increased leptin, TNF- α , adipose tissue-derived IL-6, and decreased adiponectin, respectively (15, 117).	An acute bout of exercise enhances insulin sensitivity and glucose uptake mainly on account of skeletal muscle activity; however, the effects tend to dissipate within days (78). Prolonged high intensity exercise training can sustain insulin sensitivity (76, 78) and protect against development of type 2 diabetes (77) perhaps by reducing abdominal fat, increasing skeletal muscle mass, increasing glucose transport into the muscle,(235) or decreasing fatty acid synthesis (76, 78).
Body weight, BMI	Overweight and obesity generally results in: higher levels of sex hormones (71, 236, 237); higher levels of aromatase, which converts androgens to estrogens in adipose tissue, and therefore higher levels of total estradiol (57, 75, 238); more abdominal fat and thus more 17 β -hydroxysteroid dehydrogenase, which regulates the conversion of androstenedione to testosterone (228); chronic release of free fatty acids from adipose tissue, resulting in reduced uptake of glucose by the tissues and consequently, increased circulating insulin (57, 238); lower levels of SHBG in response to hyperinsulinemia and thus, higher circulating levels of bioavailable estradiol and testosterone (57, 238); greater release of leptin, IL-6, and TNF- α from adipose tissue and decreased adiponectin (57, 111, 141, 238).	Reduces body weight (41) and decreases central adiposity in some populations (239) including postmenopausal women (17, 107, 108, 216, 240).

Not surprisingly, clinical practice guidelines for treatment of overweight and obesity in the United States (39) and Canada (40) support a convincing association between long-term physical activity and weight loss (Table 3). Although calorie restriction induces more weight loss than exercise alone (41), both countries advocate long-term regular exercise to induce modest weight loss in overweight and obese adults. Exercise is also recommended to maintain weight loss (40, 42, 43). A meta-analysis of 25 years of lifestyle weight loss programs specifically showed that aerobic exercise alone (mean, 15.6 weeks) decreased BMI by 0.8 and initial

weight by 3.6% on average (41). In a meta-analysis of nine pedometer-based interventions, there was a strong dose-response relation whereby longer intervention duration was associated with greater weight loss (44). Another review of eight recent (2000-2006) exercise randomized control trials (RCT) suggested that longer duration of physical activity is optimal for decreasing body weight and adiposity (45).

Estrogens and SHBG. Endogenous estrogen status has become a well-established risk factor for postmenopausal breast cancer (46) regardless of individual risk (predicted

Table 2. Results from epidemiologic studies of proposed biomarkers and breast cancer risk in postmenopausal women

Proposed biomarker in blood	Type of study design, study results (\pm /NA) and number of analyses					
	Cross-sectional			Case-control		
	+	-	NA	+	-	NA
Estrone				2 (241, 242)		
Estradiol						2 (241, 242)
Testosterone				1 (242)		1 (241)
Androstenedione						1 (241)
SHBG						2 (241, 242)
Leptin				2 (125, 126)		4 (245-248)
Adiponectin					4 (125, 204, 245, 250)	2 (219, 251)
TNF- α						
Interleukin-6						
CRP						
Insulin	1 (85)			2 (86, 87)		3 (88, 92, 93)
Glucose				1 (88)		1 (93)
C-peptide				2 (89, 90)		3 (87, 92, 98)
Body weight, BMI	Reviews of the epidemiologic literature support positive associations between postmenopausal breast cancer risk and BMI and/or body weight (3, 4, 31, 33-38).					

NOTE: +, positive association and $P \leq 0.05$; -, negative association and $P \leq 0.05$; NA, no association and $P > 0.05$.

*Original studies included in pooled analyses were exclusive from studies referenced elsewhere in the table. The pooled analysis by Key et al. (59) included only prospective studies in postmenopausal women: six studies on estrone, nine on estradiol, seven on SHBG, seven on testosterone, and five on androstenedione. An update on one cohort included the pooled analysis (253) was subsequently published (254) but was not included in the table to avoid double counting.

by a modified Gail model, the Rosner and Colditz model, or family history of breast cancer; ref. 47) with possible influence on the initiation, promotion, and progression of breast cancer (48). Furthermore, the successful use of antiestrogenic drugs in reducing breast cancer incidence rates supports a role for estrogens in breast cancer etiology (49-51). Several estrogen metabolites may influence breast cancer risk (52) including estrone, estrone sulfate, and 17 β -estradiol (estradiol), the most biologically active endogenous estrogen (53). Higher SHBG might lower risk by acting as a negative modulator of estradiol (54) and by reducing estrogen bioavailability (55). The most common hypothesis linking postmenopausal breast cancer to estrogens relates to adiposity (Table 1). The main source of circulating estrogens, for postmenopausal women who do not use hormone replacement therapy, derives from androgen aromatization in the peripheral tissues such as bone, muscle, brain, and most notably, adipose tissue (56-58).

As expected, our review of the literature revealed a convincing association between postmenopausal breast cancer and estrone and a probable association with estradiol (Table 2). Some of the strongest evidence stemmed from a pooled analysis of prospective studies in postmenopausal women (59), in which the odds ratio for breast cancer was 2.00 (95% CI, 1.47-2.71) for the highest versus the lowest quintiles of total estradiol; for estrone the odds ratio was 2.19 (95% CI, 1.48-3.22). The evidence for SHBG was less convincing, deemed possible using our criteria, with the pooled analysis resulting in an odds ratio of 0.66 (95% CI, 0.43-1.00) reaching only borderline statistical significance (59).

In relation to physical activity, associations were rated probable for estrone but only possible for estradiol and SHBG (Table 3). Two RCTs examined estrone and estradiol in relation to physical activity with one RCT showing no association (60) and the other showing significant inverse associations with both metabolites (18).

It is worth noting that most physical activity studies were cross-sectional in nature (5 of 8 studies on estrone, 7 of 10 studies on estradiol, 5 of 9 studies on SHBG), which is a limited study design because the temporal sequence of cause and effect cannot be shown.

It is unclear whether or not physical activity lowers estrogen levels independently of weight loss. Some cross-sectional studies have found significant inverse associations between physical activity and estrogen levels even after controlling for BMI or adiposity (61-64), suggesting an independent role for physical activity. Yet in a 12-month RCT (18), postmenopausal women assigned to an exercise group who lost $\geq 0.5\%$ body fat experienced decreased estrone, estradiol, free estradiol and increased SHBG, whereas women who did not lose body fat experienced increased estrogen levels.

Androgens. As for estrogens, a growing body of epidemiologic evidence supports a positive association between androgen levels and postmenopausal breast cancer risk (46). Androgens are the most abundant sex steroid hormones in postmenopausal women, with testosterone being one of the most powerful natural forms (65). Before and after menopause, adrenal- or ovarian-derived androstenedione gives rise to testosterone (and its derivative, dihydrotestosterone) in the ovaries and in other tissues such as adipose and breast (65-68). As with estradiol, testosterone binds reversibly to SHBG rendering it biologically inactive (55). The aromatase enzyme converts testosterone to estradiol, and androstenedione to estrone, within the adipose tissue of postmenopausal women (56).

It remains unclear whether increased risk results from androgens increasing breast cell growth directly or indirectly via estrogen production (Table 1; ref. 69). For example, one prospective study in postmenopausal women found little association between androgens and breast cancer risk after controlling for estrogen levels

Table 2. Results from epidemiologic studies of proposed biomarkers and breast cancer risk in postmenopausal women (Cont'd)

Type of study design, study results (\pm /NA) and number of analyses									Classification
Nested case-control or case-cohort			Cohort			Pooled analysis*			
+	-	NA	+	-	NA	+	-	NA	
2 (70, 243)						1 (59)			Convincing
3 (70, 72, 243)		1 (244)				1 (59)			Probable
3 (70, 72, 243)		1 (244)				1 (59)			Probable
2 (70, 72)		1 (243)				1 (59)			Probable
	2 (70, 72)	2 (243, 244)					1 (59)		Possible
	1 (252)	1 (249)							Possible
					1 (156)				Possible
					1 (156)				Hypothesized
					2 (156, 205)				Hypothesized
					1 (94)				Possible
					2 (94, 95)				Possible
1 (91)		2 (96, 97)							Possible
									Convincing

(70). Likewise, respective adjustments for estrone, estradiol, and free estradiol (but not testosterone or androstenedione) substantially weakened associations between BMI and postmenopausal breast cancer risk in one nested case-control study (71). However, adjustment for estradiol levels only slightly attenuated the relative risk associated with testosterone in a pooled analysis of prospective studies (59) and at least one other cohort study (72), thus supporting an independent mechanism for androgens.

Our review found probable associations between postmenopausal breast cancer risk and androstenedione and testosterone, respectively, because most studies supported positive relations (Table 2). In a pooled analysis of prospective studies, postmenopausal women in the highest quintiles of serum testosterone and androstenedione concentrations, respectively, had more than double the risk of developing breast cancer compared with women in the lowest quintiles (relative risk, 2.22; 95% CI, 1.59-3.10 for testosterone; relative risk, 2.15; 95% CI, 1.44-3.21 for androstenedione; ref. 59). In the same analysis, a doubling of androgen levels produced an estimated 20% to 40% increase in breast cancer risk.

Physical activity could lower testosterone levels by decreasing adiposity or by increasing circulating SHBG (refs. 73-75; Table 1). However, our literature review suggested only possible inverse associations between physical activity and serum testosterone and serum androstenedione (Table 3). Similar to the estrogen literature, much of this evidence was derived from cross-sectional studies (4 of 8 studies on testosterone, 4 of 6 studies on androstenedione) and hence, causal inference is limited.

Insulin Resistance. Insulin resistance describes the reduced effectiveness of insulin to regulate blood glucose, primarily via skeletal muscle (76, 77). When tissues cease to respond effectively to insulin, glucose uptake is reduced while the liver increases glucose biosynthesis, resulting in hyperglycemia. The pancreatic response to high blood glucose is increased insulin secretion, resulting in hyperinsulinemia (78-80). Although

insulin resistance can coincide with normal or impaired glucose tolerance, it also increases risk of type 2 diabetes and is a key component of the metabolic syndrome, two conditions that are modifiable by physical activity (77, 81). Genetic and environmental factors contribute to both insulin resistance and the metabolic syndrome; obesity, however, and particularly intra-abdominal adiposity are also considered important determinants of risk (77, 82).

A causal link between insulin resistance and postmenopausal breast cancer risk is biologically plausible (Table 1). Furthermore, in one meta-analysis, diabetes mellitus (largely type 2) was associated with a significant (16%) increase in postmenopausal breast cancer risk (83). Our review suggested a possible increased risk of postmenopausal breast cancer with higher levels of serum insulin, glucose, and C-peptide (a marker of pancreatic insulin secretion; ref. 84), respectively (Table 2). Of the 14 studies measuring at least one of these markers in postmenopausal women, seven found at least one significant positive association (85-91), whereas the remainder showed no association (92-98). Also one large cohort study (99) revealed a weak inverse association between postmenopausal breast cancer risk and Hb_{A1C}, a measure of long-term blood glucose levels (100).

In contrast to its possible association with breast cancer risk, we classified the link between physical activity and insulin resistance as convincing (Table 3) based on the scientific consensus (7, 76, 101-104). Specifically, a statement by the American Diabetes Association, the North American Association for the Study of Obesity, and the American Society for Clinical Nutrition (102), and others promote moderate weight loss via regular aerobic, and possibly resistance (103), exercise to improve insulin sensitivity (105) and prevent diabetes (7, 76, 102, 104). The effect of exercise may be strongest for those with impaired (versus normal) glucose tolerance (76) and when followed at higher doses (106), higher intensity (107), or when as combined aerobic/resistance exercise versus aerobic exercise alone (108).

Adipokines and Inflammatory Markers. Adipokines (adipocytokines) are a group of biologically active

Table 3. Results from epidemiologic studies of proposed biomarkers and physical activity in postmenopausal women

Proposed biomarker in blood	Type of study design, study results (\pm /NA) and number of analyses					
	Cross-sectional			Case-control		
	+	-	NA	+	-	NA
Estrone	3 (61, 63, 255)		2 (62, 256)		1 (257)	
Estradiol	2 (62, 255)		5 (61, 63, 64, 258, 259)			1 (257)
Testosterone	1 (62)		3 (61, 255, 256)			1 (257)
Androstenedione	2 (63, 255)		2 (61, 62)			
SHBG	1 (261)		4 (63, 258, 259, 262)			
Leptin						
Adiponectin						
TNF- α			2 (157, 158)			
Interleukin-6	2 (157, 173)		1 (158)			
CRP	2 (157, 214)					
Insulin, glucose, C-peptide	Reviews generally promote moderate weight loss with exercise to improve insulin sensitivity (101, 105) and prevent diabetes (7, 76, 102, 104).					
Body weight, BMI	Clinical guidelines for weight loss in the United States (39) and Canada (40) advocate long-term regular exercise to induce modest weight loss in overweight and obese adults.					

NOTE: +, -, and NA $P > 0.05$.

*The associations deemed "+" or "-" were based on the change observed in exercisers versus the change observed in controls with the exception of the RCT described by Giannopoulou et al. (129), which did not include a control arm; rather, the results at baseline were compared with the end of study within the exercise only arm.

polypeptides produced by adipocytes or adipose tissue; they include leptin, adiponectin, TNF- α , and IL-6 (109-111). CRP is not an adipokine but is produced in the liver in response to TNF- α and IL-6 levels (112), which are all considered common indicators of inflammation (113). Chronic inflammation is an acknowledged risk factor for several cancers (114), and obesity (112) and the metabolic syndrome (115) represent low-grade, systemic inflammatory states with elevated levels of inflammatory markers. The sustained proliferative activity, microenvironmental changes, and oxidative stress associated with chronic inflammation could act together to deregulate normal cell growth and development, thereby promoting initiated cells toward malignancy (116).

Postmenopausal breast cancer risk is positively associated with increased BMI (Table 2), and adipokines similarly exhibit strong positive correlations with BMI (15), hyperinsulinemia, insulin resistance, and type 2 diabetes (conditions related to overweight and obesity; refs. 15, 117). Several biological mechanisms implicate adipokines in breast cancer etiology (Table 4), but the extent to which adipokines influence breast cancer risk directly or act as biomarkers of increased adiposity, insulin resistance, and chronic inflammation, is unresolved.

Regular physical activity may reduce inflammation independently of fat loss (118), but the mechanisms are unknown. One RCT in postmenopausal women (118) found that a 6-month weight loss intervention, comprising a hypocaloric diet and exercise, significantly decreased plasma TNF- α , IL-6, and CRP whereas significant changes were not observed in the group receiving only the diet intervention. Interestingly, the two groups experienced similar losses in body weight and adipose tissue. Furthermore, a recent review article concluded that physical fitness generally decreases inflammatory markers even after adjusting for adiposity (119).

Leptin. Leptin is both a neurohormone and a member of the cytokine superfamily (120). Although first discov-

ered in 1994 (121), its role in cancer etiology is only recently appreciated. Adipose tissue is quantitatively the most important source of leptin and the primary determinant of circulating leptin levels (122-124). Leptin is widely known for its ability to counteract obesity by inducing satiety and limiting caloric intake (123), but paradoxically, human obesity is associated with higher levels of circulating leptin, possibly signifying leptin resistance (57, 122). Considerable *in vitro* evidence implicates leptin as a risk factor for breast cancer (Table 4), either by direct mitogenic action on breast cells or perhaps indirectly, for example, by increasing estrogen production or by promoting insulin resistance.

Despite biological plausibility, we classified the epidemiologic evidence relating leptin to postmenopausal breast cancer risk as possible (Table 2). Of the seven studies we reviewed, only two case-control studies (125, 126) found significant positive associations. We did not identify any cohort studies in postmenopausal women. In contrast, the relation between higher physical activity and decreased leptin was deemed probable in our review (Table 3). Although only five studies were identified, all five were prospective trials and four (16, 127-129) produced significant decreases in leptin. Moreover, reviews have suggested that the greatest impact on leptin is achieved by exercise training of longer duration, extending beyond 12 weeks, and at higher intensities (130, 131).

Adiponectin. Adiponectin was first described in 1995 as the most abundant gene product of human adipocytes (132). It is now gaining recognition as a predictive indicator of abdominal fat and obesity-related sequelae such as the metabolic syndrome (133) and now, possibly, breast cancer. Although adiponectin is produced only by adipocytes (57, 134), unlike other adipokines, it has a strong inverse correlation with adiposity (135); consequently, weight loss increases adiponectin levels (136, 137). The relation may occur in part because IL-6 and TNF- α , which increase in obesity (138), are potent

Table 3. Results from epidemiologic studies of proposed biomarkers and physical activity in postmenopausal women (Cont'd)

Type of study design, study results (\pm /NA) and number of analyses						Classification
Trial			RCT*			
+	-	NA	+	-	NA	
				1 (18)	1 (60)	Probable
				1 (18)	1 (60)	Possible
	2 (240, 260)			1 (19)		Possible
	2 (240, 263)	1 (260)			2 (19, 60)	Possible
	2 (127, 128)	1 (264)		2 (16, 129)	1 (18)	Possible
		1 (147)				Probable
		1 (128)			1 (129)	Hypothesized
	1 (265)			1 (129)	1 (129)	Hypothesized
						Hypothesized
						Convincing
						Convincing (general population)
						Convincing (general population)

inhibitors of adiponectin expression and secretion (ref. 139). Generally, adiponectin is acknowledged to be anti-inflammatory and antiatherogenic (140-143), and lower levels of adiponectin are strongly associated with insulin resistance (144, 145), perhaps more strongly than obesity or adiposity (146).

Although lower levels of adiponectin may imply increased risk for postmenopausal breast cancer, we found only a possible association based on our literature review (Table 2). Five of seven studies in postmenopausal women showed negative associations, but none were prospective in nature and therefore these data are limited for assessing causality. In relation to physical activity, only a hypothesized association exists at this time (Table 3). We identified only two studies of exercise and adiponectin in postmenopausal women (129, 147); both were prospective trials, but neither found any statistically significant effect.

The effect of exercise on adiponectin in other populations (i.e., not exclusively postmenopausal women) has been reviewed previously (148, 149). Despite three of eight exercise trials producing significantly increased adiponectin levels in one review (148), the overall evidence on chronic exercise was inconclusive because many factors were uncontrolled for, namely, weight change, diet, and the effects of cytokines. Furthermore, regression analysis of 15 exercise trials found no significant relation between changes in body weight and adiponectin (148). According to another review (149), chronic exercise that improves fitness, increases insulin sensitivity, and reduces body weight will increase adiponectin levels if the training volume is sufficiently high and lasts longer than 2 months. Whether exercise influenced adiponectin through weight loss alone was unclear from these studies (149).

TNF- α . TNF- α is a cytokine produced mainly by macrophages that infiltrate adipose tissue in obesity (109) but also by a variety of tumor cells, including breast carcinoma (114, 150). TNF- α is well known for its critical roles in host defense, inflammation, and organogenesis (151). The relation of TNF- α to cancer, however, is less straightforward. Although TNF- α can induce apoptosis and necrosis, as its name implies, chronic moderately

elevated levels of TNF- α seem to promote nearly all steps leading to cancer, from cellular transformation to metastasis (Table 4; refs. 114, 151, 152). Because TNF- α mRNA and TNF- α protein are released by adipose tissue (153-155), weight loss may decrease circulating TNF- α .

To our knowledge, only one epidemiologic study has evaluated TNF- α in postmenopausal breast cancer (ref. 156; Table 2). This cohort study followed 2,438 older adults (ages 70-79 years) for an average of 5.5 years. Despite biological plausibility, no association was found between circulating TNF- α and breast cancer incidence. Similarly, of four studies investigating physical activity and TNF- α (128, 129, 157, 158), none found a statistically significant association, although lower TNF- α levels corresponded with increased physical activity in one study (cross-sectional; ref. 157) and TNF- α levels decreased after 14 weeks of exercise in another (RCT; ref. 129). Consequently, we classified both associations as hypothesized (Tables 2 and 3).

IL-6. IL-6 is a cytokine that occurs predominantly in circulating form (159) originating from a number of sources (160) including fibroblasts, macrophages, lymphocytes (145), skeletal muscle (161), and adipose tissue (162). Obesity is strongly associated with elevated circulating IL-6 (161), although it is estimated that adipocytes *per se* account for only 10% of IL-6 released from adipose tissue (163). Because TNF- α stimulates the release of IL-6, it is suggested that moderate increases in systemic IL-6 and CRP may actually reflect chronic TNF- α production (164, 165). IL-6 has a broad range of regulatory functions involving inflammation and immune responses (15) but might also increase breast cancer risk by IL-6-induced insulin resistance (145) and aromatase activity (Table 4; ref. 166).

Our review identified only one study examining IL-6 levels relative to postmenopausal breast cancer risk; no significant association was found (156) and so we deemed this association hypothesized (Table 2). Interestingly, other studies not exclusive to postmenopausal women have showed higher IL-6 levels in breast cancer cases relative to controls (167-171), and some have implicated IL-6 as a negative prognosticator in breast

Table 4. Possible mechanisms relating recently proposed biomarkers to postmenopausal breast cancer risk and physical activity

Proposed Biomarker	Possible role in postmenopausal breast cancer	Possible impact of physical activity in postmenopausal women
Leptin	Induces aromatase and stabilizes estrogen receptor- α (57, 122, 124, 266-268). Although leptin can improve insulin sensitivity (145), elevated leptin levels are associated with insulin resistance (117). Hypothalamic actions of leptin could theoretically decrease systemic insulin sensitivity and adiponectin production (269). Expression is induced by high levels of estrogens and insulin (124, 141, 268). Mitogen in breast cancer cells (15); inhibits apoptosis; pro-angiogenic (111, 122, 124, 141).	Weight loss decreases body fat, which is the main source of circulating leptin (122-124).
Adiponectin	Gene expression and secretion from adipocytes are reduced by TNF- α and IL-6 (57, 142); production might also be reduced partially by leptin (269). Promotes and enhances insulin sensitivity (140, 142, 144, 270); reduced adiponectin leads to insulin resistance and compensatory hyperinsulinemia (271). Antiangiogenic (272); antimitogenic, and anti-inflammatory (111). In one breast cancer cell line adiponectin had no effect on apoptosis but did inhibit cell proliferation (273).	Fat loss decreases IL-6 and TNF- α (138), which are potent inhibitors of adiponectin expression and secretion (139). Hence, weight loss may increase circulating adiponectin levels. Chronic physical activity may lower inflammation (e.g., circulating IL-6, TNF- α) independently of fat loss (118); however, the mechanisms for this effect are unknown.
TNF- α	A key regulator of IL-6 synthesis (57). Stimulates estrogen biosynthesis via aromatase induction (166). Induces insulin resistance (109, 164). Paradoxical action: inhibits tumor cell proliferation (274) but also acts as a tumor promoter (151, 152, 275). Can cause direct DNA damage; antiapoptotic and mitogenic (151); promotes invasion, angiogenesis and metastasis of tumor cells (114, 150, 152).	Fat loss may decrease TNF- α levels given that TNF- α mRNA and TNF- α protein are released from adipose tissue in obesity (153-155). Chronic physical activity may reduce the number of mononuclear cells in the blood thereby depleting a source of TNF- α (138).
IL-6	Release is stimulated by TNF- α ; has been speculated that systemic IL-6 reflects ongoing production of TNF- α (164, 165); IL-6 in turn, exerts inhibitory effects on TNF- α (165). Plays a primary role in stimulating hepatic production of CRP (159). Produces insulin resistance in adipocytes (276, 277); possible role in type 2 diabetes (160). Stimulates estrogen biosynthesis by the induction of aromatase activity (166). Promotes breast cancer cell motility suggesting a role in metastasis (167). Complex role of IL-6 in breast cancer cells <i>in vitro</i> . Up-regulates antiapoptotic and angiogenic proteins in tumor cells but also induces apoptosis in estrogen receptor-positive mammary carcinoma cell lines (172).	Although the acute effects of exercise on IL-6 levels have been studied widely (160), the mechanisms whereby chronic physical activity alter IL-6 levels are unclear (161). Reduced adiposity may decrease IL-6 levels given that IL-6 originates from adipose tissue (162), among other sources. Chronic physical activity may reduce the number of mononuclear cells in the blood thereby depleting a source of IL-6 (138).
CRP	A prototypical marker of inflammation (174). Production is promoted by TNF- α and IL-6 (112). Independently associated with leptin in healthy individuals, possibly via induction of IL-6 by leptin (278). CRP production is strongly, positively related to insulin resistance and can change with insulin levels independently of changes in obesity (279).	Long-term physical activity may decrease CRP by reducing adiposity, by reducing cytokine production (i.e., IL-6 and TNF- α) in muscle and mononuclear cells, or by other means (179).

cancer patients (172). Therefore, a similar relation in postmenopausal women remains plausible.

We classified the relation between physical activity and IL-6 in postmenopausal women as hypothesized (Table 3). Only cross-sectional studies were identified, and two (157, 173) of the three showed significant inverse associations. In both studies, statistical significance was maintained even after adjusting for BMI. Thus, although

weight loss can reduce IL-6 (138), exercise might modify IL-6 levels through an independent mechanism.

CRP. CRP is a hepatocyte-derived, acute phase protein considered to be the prototypical marker of inflammation in humans (174) and might also affect postmenopausal breast cancer risk. CRP levels correlate positively with weight (175, 176) and weight gain (177) and have been

similarly related to type 2 diabetes risk (178). Because physical activity confers benefits in each of these conditions, one might expect it similarly to decrease CRP levels.

We identified only two studies of CRP levels and postmenopausal breast cancer risk. Due to the limited epidemiologic literature, we deemed the association to be hypothesized (Table 2). It is noteworthy, however, that both were cohort studies and neither found a statistically significant association. In contrast, an association between CRP and physical activity has been more widely studied and is quite persuasive (Table 3). We identified four studies in postmenopausal women, including one prospective trial and one RCT; all four found that higher levels of physical activity corresponded to lower levels of circulating CRP. We regarded this association as convincing in postmenopausal women. Reviews in other populations support this conclusion. One review of 17 cross-sectional studies of regular physical activity showed consistent evidence of lowered serum CRP in

the highest versus the lowest physical activity levels (179). In another review, longitudinal studies showed reduced CRP levels with exercise training (180).

In postmenopausal women it remains unclear whether exercise or weight loss modifies CRP levels (119, 138). One review concluded that CRP levels decline whether weight loss is achieved through exercise or diet (181), implying weight loss is most important. A meta-analysis of five RCTs of long-term exercise training in men and women found statistically significant decreases in body weight and adiposity but only a nonsignificant reduction in CRP levels, suggesting neither exercise nor weight loss is effective (182); however, not all subjects had high CRP levels at baseline and only one study focused on postmenopausal women (183). In the latter RCT of breast cancer survivors, the mean CRP level decreased in the exercise group, but relative to controls this decrease was not statistically significant ($P = 0.066$).

Proposed Biological Model. An overview of our proposed mechanisms for breast cancer (Tables 1 and 4) and

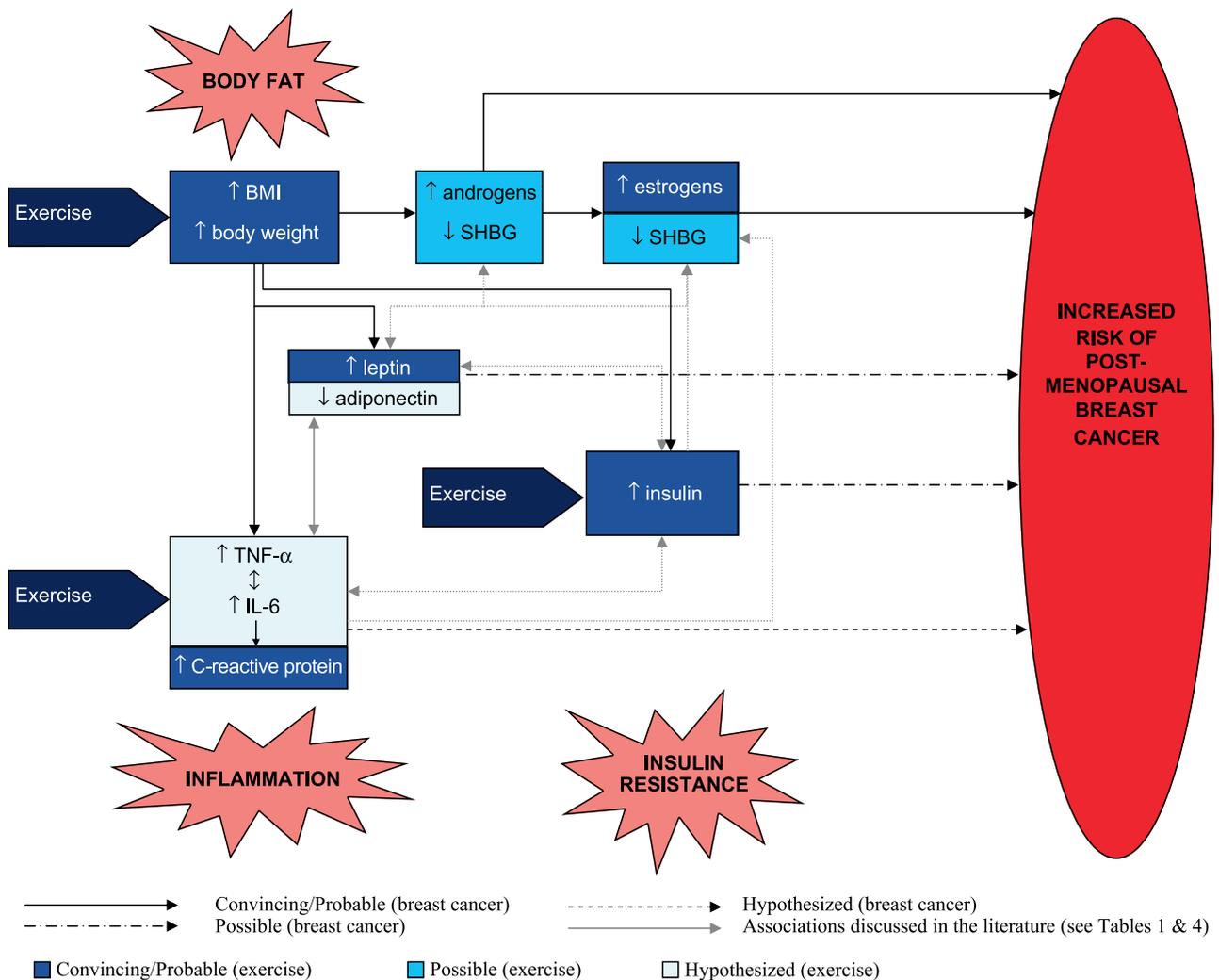


Figure 1. Biological model relating proposed biomarkers to long-term exercise (*shading*) and postmenopausal breast cancer risk (*arrows*).

a summary of findings from our literature review (Tables 2 and 3) are illustrated in Fig. 1. This figure suggests that the co-occurrence of overweight/obesity, insulin resistance, and chronic inflammation may increase postmenopausal breast cancer risk through several complex biological mechanisms. As shown, the most convincing epidemiologic evidence supports associations between breast cancer risk and body weight, estrogens, and androgens, respectively. In relation to physical activity, associations were most convincing for body weight, estrone, leptin, insulin resistance, and CRP levels. Only body weight and estrone were convincingly (or probably) associated with breast cancer risk and physical activity. The least amount of evidence supported roles for TNF- α and IL-6 in this model based on the epidemiologic evidence to date.

This review focused on postmenopausal women and thus, we can only speculate on the effects of these influences over the life course. Hypothetically, cumulative lifetime levels of some biomarkers (e.g., related to insulin resistance or inflammation) might affect postmenopausal breast cancer risk, and sustained physical activity could modify lifetime exposure. However, the epidemiologic evidence relating physical activity and body size to breast cancer is less convincing for premenopausal women (4, 5, 184) and hence, the proposed biological model probably becomes more important after menopause. This finding might reflect a mechanism involving adiposity and sex hormones, for example, because only after menopause, when ovarian production of estrogens has ceased, does adipose tissue become the primary source of circulating estrogens.

Discussion and Future Research Directions

This review provides an overview of three commonly proposed mechanisms relating physical activity to postmenopausal breast cancer risk; namely overweight and obesity, insulin resistance, and chronic inflammation. As illustrated in our model, several mechanisms potentially act simultaneously to increase postmenopausal breast cancer risk with opposing, promoting, and possibly synergistic pathways at play. Considering the complexity of the model, it is not surprising that epidemiologic findings have been inconsistent for some candidate biomarkers. Several questions remain regarding the biological mechanisms that mediate the association between physical activity and breast cancer. Which biomarkers are most predictive of risk? Which are direct versus indirect consequences of exercise? The proposed model also has important implications for the design and analysis of future etiologic studies. It is conceivable, for instance, that the combined effects of elevated sex hormones, insulin resistance, and elevated inflammatory markers present greater risk than any one factor individually; future etiologic studies should explore potential interactions among biomarkers. The notion that interacting pathways connect physical activity to breast cancer risk was proposed by Hoffman-Goetz et al. ten years ago (10), but since then most epidemiologic researchers have continued to study candidate biomarkers in isolation.

As shown in our model, all of the proposed biomarkers are, at least indirectly, related to body size.

Therefore, the effect of exercise could be entirely or only partially dependent on weight loss. The available literature does not adequately address whether fitness or decreased fatness is more important for lowering postmenopausal breast cancer risk. However, the Nutrition and Exercise in Women (NEW) Trial led by Dr. Anne McTiernan will address this issue in an ongoing RCT involving over 500 postmenopausal women (13). Additionally, Holt et al. (185) recently explored the effect of fit versus fat on insulin sensitivity in a study of 25 men. The independent effects of adiposity, physical fitness, and physical activity energy expenditure were compared. Multiple regression analysis revealed significant relations between adiposity and whole body insulin sensitivity, and between physical activity energy expenditure and liver insulin sensitivity. We encourage similar studies in the future, designed specifically to compare the effects of physical activity and weight loss and to measure biomarkers prospectively during interventions.

The generalizability of our model is tempered by potential effect modification. It is possible, for instance, that the proposed mechanisms contribute only in certain high-risk subgroups of postmenopausal women (e.g., obese women with BMI ≥ 30 and/or insulin resistance) for whom, according to our model, exercise might confer the greatest benefits. Additional factors such as genetic polymorphisms, family history, race, diet, and medications might further modify the roles of the proposed biomarkers on breast cancer risk. For example, weight gain and BMI only increase postmenopausal breast cancer risk in never users of hormone replacement therapy (186-191). The effect of exercise on breast cancer risk may be similarly modified by hormone replacement therapy use. Two (192, 193) of at least four (192-195) observational studies that examined effect modification of this kind found a stronger protective effect of physical activity among never hormone replacement therapy users as compared with ever users. Furthermore, one cross-sectional study of postmenopausal women showed a two-fold increased median CRP level in hormone replacement therapy users compared with nonusers with control for potential confounders (196). Besides hormone replacement therapy, several other CRP-altering medications have been identified (197). Finally, dietary composition and smoking could hypothetically modify the benefits of exercise by influencing insulin resistance and inflammation (198-200). Given the multitude of potential effect modifiers in our model, the notion that physical activity decreases breast cancer risk generally in postmenopausal women cannot be practically applied to individuals.

It is also conceivable that the proposed biomarkers increase risk for only certain tumor types; namely, hormone receptor-positive or hormone receptor-negative tumors. Interactions with estrogen receptor status have been explored previously, for instance, in relation to breast cancer risk and body weight (186, 191, 201), adiposity (202), sex hormone levels (203), impaired glucose metabolism (99), serum adiponectin (204), and IL-6 levels (171). Furthermore, a recent review provided preliminary evidence that physical activity produces a somewhat greater risk reduction for hormone receptor-negative tumors (estrogen receptor negative/progesterone receptor negative) than for hormone receptor-positive tumors (estrogen receptor positive/progesterone receptor positive; ref. 6). Future etiologic

studies should similarly consider tumor characteristics to advance the interpretation of our model.

Despite the potential importance of frequency, duration, or intensity, surprisingly little evidence exists for defining an effective exercise prescription to reduce postmenopausal breast cancer risk (4). One might expect particular types of physical activity to target abdominal fat and perhaps greater levels of energy expenditure to achieve greater weight loss. Some authors propose that exercise of greater duration, intensity, or volume may also be most effective for decreasing leptin (130, 131) and increasing adiponectin (149). Hence, it would be very useful for future exercise intervention trials to compare the effects of different exercise prescriptions in the context of our biological model.

Our review revealed several noteworthy gaps in past epidemiologic research. First, there is clearly a lack of prospective exercise trials examining sex hormone changes in postmenopausal women undergoing physical activity modification. Thus, it is unclear whether or not exercise is causal in reducing androgen and estrogen levels. This is important to note because sex hormones are currently the most compelling candidate biomarkers, given their probable/convincing associations with breast cancer risk (Table 2). Second, the biomarkers designated as hypothesized in this review are hypothetical due largely to a lack of research. Only one study on TNF- α and IL-6 (156) and two on CRP (156, 205) related specifically to postmenopausal breast cancer risk. Furthermore, the soluble receptor for IL-6 (sIL-6R) is a known agonist of IL-6 activity whereas higher levels of TNF soluble receptors (sTNFR1, sTNFR2) inhibit TNF- α activity (206); these could additionally contribute to our proposed model. Therefore, in the context of postmenopausal breast cancer, more epidemiologic research into inflammatory markers and their soluble receptors is warranted.

Despite biological plausibility, our review of four independent studies (128, 129, 157, 158) suggested no relation between TNF- α and physical activity level (128, 129, 157, 158). This finding may indicate that greater levels of concurrent weight loss (or weight disparities in observational studies) are required before differences in TNF- α levels are detectable. Alternatively, changes in TNF- α might be confounded by dietary composition, which is also associated with chronic inflammation (200). Finally, it is possible that compensatory mechanisms to proinflammatory factors or perhaps soluble TNF receptors varied with exercise, but these factors were not studied and/or were excluded from our review. Given these possibilities, excluding TNF- α from our model at this stage might be premature.

To maximize the pool of literature for review, we selected BMI and body weight as our surrogate measures of adiposity. BMI predicts total body fat with varying accuracy depending on the study population (207, 208) but is generally comparable with alternative measures (BMI - % body fat, $r = 0.69-0.75$ (208, 209) and $r = 0.81$ for postmenopausal women (210); BMI - abdominal fat assessed by computer tomography, $r = 0.8$ ref. 207). Therefore, BMI is an acceptable measure for guiding public health recommendations. Abdominal fat, however, may be more etiologically relevant to breast cancer mechanistic research given its inverse association with SHBG levels (211) and insulin sensitivity (212) and

positive associations with circulating insulin (213) and CRP levels (214) in postmenopausal women.

Positive associations have been found in many, but not all, studies of waist-hip ratio, waist circumference, or other measures of central adiposity and breast cancer risk in postmenopausal women (3, 32, 33, 36, 75). Abdominal fatness was deemed a probable risk factor for postmenopausal breast cancer in the World Cancer Research Fund/American Institute for Cancer Research 2007 report (4). However, current North American guidelines claim there is only limited evidence supporting the effectiveness of physical activity for abdominal fat loss (ref. 40; also concluded by ref. 215) and only modest reductions in abdominal fat to be expected, if at all (39). It remains plausible, however, that postmenopausal women could be amenable to significant abdominal fat loss given the most effective exercise prescription (e.g., evidenced by refs. 17, 108, 216). Thus, further exercise trials and etiologic studies are needed, ideally using more accurate measures of abdominal fat such as dual energy X-ray absorptiometry. Perhaps more accurate, targeted anthropometric measures will clarify the role of central adiposity.

Our model focuses primarily on mechanisms related to the promotion of postmenopausal breast cancer, but physical activity could influence risk at several points along the cancer continuum (8). Markers associated with detoxification pathways, DNA repair mechanisms, oxidative stress, and various aspects of immune function could all be relevant to postmenopausal breast cancer and also modifiable by exercise (6, 8-10). Moreover, recently proposed biomarkers may be integrated into our model as new research is conducted. For example, resistin is an adipose tissue-derived polypeptide associated with (and named for) insulin resistance in rodents, but its role in humans remains controversial; it might induce insulin resistance or could play a proinflammatory role (217, 218). Resistin is an emerging risk factor for breast cancer with significantly increased serum levels in postmenopausal cases relative to controls in two recent studies (125, 219). However, two recent studies suggested resistin is not modifiable by exercise (129, 220). Still, based on the limited evidence to date, more research is warranted to elucidate resistin's response to exercise and its possible role in postmenopausal breast cancer.

In summary, the current review provides a conceptual framework for future research into the biological mechanisms surrounding physical activity and postmenopausal breast cancer risk. BMI and sex hormones have so far been the most commonly cited biomarkers relating physical activity to decreased risk, but emerging evidence now suggests that insulin resistance and chronic inflammation could play pivotal roles. The important interrelations between these mechanisms must be considered when analyzing data or planning future studies. Two general types of prospective studies are required to validate our model: cohort studies relating the proposed biomarkers to cancer risk, and exercise RCTs comparing biomarker changes at several time points over the long term, specifically in postmenopausal women. Convincing findings from both fields of study, with account for effect modification, would strengthen the existing epidemiologic evidence, and would ultimately guide breast cancer prevention strategies for postmenopausal women.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

Acknowledgments

We thank Colleen Lachance for her conscientious administrative support throughout this review.

References

- Canadian Cancer Society, National Cancer Institute of Canada. Canadian cancer statistics 2008. Toronto (Canada). 2008.
- United States Cancer Statistics: 1999–2004 Incidence and Mortality Web-based Report. U.S. Cancer Statistics Working Group 2007. Atlanta: U.S. Department of Health and Human Services, Centers for Disease Control and Prevention and National Cancer Institute.
- IARC Working Group. IARC handbook of cancer prevention, volume 6: weight control and physical activity. Lyon: IARC; 2002.
- World Cancer Research Fund and the American Institute for Cancer research. Food, nutrition, physical activity, and the prevention of cancer: a global perspective. American Institute for Cancer Research; 2007.
- Monninkhof EM, Elias SG, Vlems FA, et al. Physical activity and breast cancer: a systematic review. *Epidemiology* 2007;18:137–57.
- Friedenreich CM, Cust AE. Physical activity and breast cancer risk: impact of timing, type and dose of activity and population subgroup effects. *Br J Sports Med* 2008;42:636–47.
- Warburton DE, Katzmarzyk PT, Rhodes RE, Shephard RJ. Evidence-informed physical activity guidelines for Canadian adults. *Can J Public Health* 2007;98 Suppl 2:S16–68.
- Rundle A. Molecular epidemiology of physical activity and cancer. *Cancer Epidemiol Biomarkers Prev* 2005;14:227–36.
- Campbell KL, McTiernan A. Exercise and biomarkers for cancer prevention studies. *J Nutr* 2007;137:161–95.
- Hoffman-Goetz L, Apter D, Demark-Wahnefried W, Goran MI, McTiernan A, Reichman ME. Possible mechanisms mediating an association between physical activity and breast cancer. *Cancer* 1998; 83:621–8.
- Shephard RJ, Shek PN. Associations between physical activity and susceptibility to cancer: possible mechanisms. *Sports Med* 1998;26: 293–315.
- Kruk J, Boul-Enein HY. Physical activity in the prevention of cancer. *Asian Pac J Cancer Prev* 2006;7:11–21.
- McTiernan A. Mechanisms linking physical activity with cancer. *Nat Rev Cancer* 2008;8:205–11.
- Westerlind KC. Physical activity and cancer prevention-mechanisms. *Med Sci Sports Exerc* 2003;35:1834–40.
- Rose DP, Komninou D, Stephenson GD. Obesity, adipocytokines, and insulin resistance in breast cancer. *Obes Rev* 2004;5:153–65.
- Frank LL, Sorensen BE, Yasui Y, et al. Effects of exercise on metabolic risk variables in overweight postmenopausal women: a randomized clinical trial. *Obes Res* 2005;13:615–25.
- Irwin ML, Yasui Y, Ulrich CM, et al. Effect of exercise on total and intra-abdominal body fat in postmenopausal women: a randomized controlled trial. *JAMA* 2003;289:323–30.
- McTiernan A, Tworoger SS, Ulrich CM, et al. Effect of exercise on serum estrogens in postmenopausal women: a 12-month randomized clinical trial. *Cancer Res* 2004;64:2923–8.
- McTiernan A, Tworoger SS, Rajan KB, et al. Effect of exercise on serum androgens in postmenopausal women: a 12-month randomized clinical trial. *Cancer Epidemiol Biomarkers Prev* 2004;13: 1099–105.
- McTiernan A, Sorensen B, Yasui Y, et al. No effect of exercise on insulin-like growth factor 1 and insulin-like growth factor binding protein 3 in postmenopausal women: a 12-month randomized clinical trial. *Cancer Epidemiol Biomarkers Prev* 2005;14:1020–1.
- Orenstein MR, Friedenreich CM. Review of physical activity and the IGF family. *J Physic Act Health* 2004;1:291–320.
- Fletcher O, Gibson L, Johnson N, et al. Polymorphisms and circulating levels in the insulin-like growth factor system and risk of breast cancer: a systematic review. *Cancer Epidemiol Biomarkers Prev* 2005;14:2–19.
- Rehnan AG, Egger M, Minder C, O'Dwyer ST, Shalet SM, Zwahlen M. IGF-I, IGF binding protein-3 and breast cancer risk: comparison of 3 meta-analyses. *Int J Cancer* 2005;115:1006–7.
- Irwin ML, Aiello EJ, McTiernan A, et al. Physical activity, body mass index, and mammographic density in postmenopausal breast cancer survivors. *J Clin Oncol* 2007;25:1061–6.
- Maskarinec G, Pagano I, Chen Z, Nagata C, Gram IT. Ethnic and geographic differences in mammographic density and their association with breast cancer incidence. *Breast Cancer Res Treat* 2007;104: 47–56.
- Reeves KW, Gierach GL, Modugno F. Recreational physical activity and mammographic breast density characteristics. *Cancer Epidemiol Biomarkers Prev* 2007;16:934–42.
- Peters TM, Ekelund U, Leitzmann M, et al. Physical activity and mammographic breast density in the EPIC-Norfolk cohort study. *Am J Epidemiol* 2008;167:579–85.
- Sellers TA, Vachon CM, Pankratz VS, et al. Association of childhood and adolescent anthropometric factors, physical activity, and diet with adult mammographic breast density. *Am J Epidemiol* 2007;166: 456–64.
- Samimi G, Colditz GA, Baer HJ, Tamimi RM. Measures of energy balance and mammographic density in the Nurses' Health Study. *Breast Cancer Res Treat* 2008;109:113–22.
- Oestreich N, Capra A, Bromberger J, et al. Physical activity and mammographic density in a cohort of midlife women. *Med Sci Sports Exerc* 2008;40:451–6.
- Rehnan AG, Tyson M, Egger M, Heller RF, Zwahlen M. Body-mass index and incidence of cancer: a systematic review and meta-analysis of prospective observational studies. *Lancet* 2008;371:569–78.
- Ziegler RG. Anthropometry and breast cancer. *J Nutr* 1997;127: 924–85.
- Friedenreich CM. Review of anthropometric factors and breast cancer risk. *Eur J Cancer Prev* 2001;10:15–32.
- van den Brandt PA, Spiegelman D, Yaun SS, et al. Pooled analysis of prospective cohort studies on height, weight, and breast cancer risk. *Am J Epidemiol* 2000;152:514–27.
- Cleary MP, Maihle NJ. The role of body mass index in the relative risk of developing premenopausal versus postmenopausal breast cancer. *Proc Soc Exp Biol Med* 1997;216:28–43.
- Ballard-Barbash R, Swanson CA. Body weight: estimation of risk for breast and endometrial cancers. *Am J Clin Nutr* 1996;63:437–15.
- Bergstrom A, Pisani P, Tenet V, Wolk A, Adami HO. Overweight as an avoidable cause of cancer in Europe. *Int J Cancer* 2001;91:421–30.
- Key TJ, Appleby PN, Reeves GK, et al. Body mass index, serum sex hormones, and breast cancer risk in postmenopausal women. *J Natl Cancer Inst* 2003;95:1218–26.
- NIH. Clinical Guidelines on the Identification, Evaluation, and Treatment of Overweight and Obesity in Adults - The Evidence Report. *Obes Res* 1998;6 Suppl 2:51–209S.
- Lau DC, Douketis JD, Morrison KM, Hramiak IM, Sharma AM, Ur E. 2006 Canadian clinical practice guidelines on the management and prevention of obesity in adults and children [summary]. *CMAJ* 2007; 176:S1–13.
- Miller WC, Kocaja DM, Hamilton EJ. A meta-analysis of the past 25 years of weight loss research using diet, exercise or diet plus exercise intervention. *Int J Obes Relat Metab Disord* 1997;21:941–7.
- Curioni CC, Lourenco PM. Long-term weight loss after diet and exercise: a systematic review. *Int J Obes (Lond)* 2005;29:1168–74.
- Hill J, Wing R. The national weight control registry. *The Permanente Journal* 2003;7:34–7.
- Richardson CR, Newton TL, Abraham JJ, Sen A, Jimbo M, Swartz AM. A meta-analysis of pedometer-based walking interventions and weight loss. *Ann Fam Med* 2008;6:69–77.
- Janiszewski PM, Ross R. Physical activity in the treatment of obesity: beyond body weight reduction. *Appl Physiol Nutr Metab* 2007;32: 512–22.
- Hankinson SE, Eliassen AH. Endogenous estrogen, testosterone and progesterone levels in relation to breast cancer risk. *J Steroid Biochem Mol Biol* 2007;106:24–30.
- Eliassen AH, Missmer SA, Tworoger SS, Hankinson SE. Endogenous steroid hormone concentrations and risk of breast cancer: does the association vary by a woman's predicted breast cancer risk? *J Clin Oncol* 2006;24:1823–30.
- Yager JD, Davidson NE. Estrogen carcinogenesis in breast cancer. *N Engl J Med* 2006;354:270–82.
- Fisher B, Costantino JP, Wickerham DL, et al. Tamoxifen for prevention of breast cancer: report of the National Surgical Adjuvant Breast and Bowel Project P-1 Study. *J Natl Cancer Inst* 1998;90:1371–88.
- Cauley JA, Norton L, Lippman ME, et al. Continued breast cancer risk reduction in postmenopausal women treated with raloxifene: 4-year results from the MORE trial. Multiple outcomes of raloxifene evaluation. *Breast Cancer Res Treat* 2001;65:125–34.
- Uray IP, Brown PH. Prevention of breast cancer: current state of the science and future opportunities. *Expert Opin Investig Drugs* 2006; 15:1583–600.
- Muti P, Rogan E, Cavalieri E. Androgens and estrogens in the etiology and prevention of breast cancer. *Nutr Cancer* 2006;56:247–52.

53. Colditz G, Baer HJ, Tamimi R. Breast cancer. In: Schottenfeld D, Fraumeni JF, editors. *Cancer epidemiology and prevention*. Oxford University Press; 2006. p. 995–1012.
54. Fortunati N, Catalano MG. Sex hormone-binding globulin (SHBG) and estradiol cross-talk in breast cancer cells. *Horm Metab Res* 2006; 38:236–40.
55. Anderson DC. Sex-hormone-binding globulin. *Clin Endocrinol (Oxf)* 1974;3:69–96.
56. Kendall A, Folkler EJ, Dowsett M. Influences on circulating oestrogens in postmenopausal women: relationship with breast cancer. *J Steroid Biochem Mol Biol* 2007;103:99–109.
57. Lorincz AM, Sukumar S. Molecular links between obesity and breast cancer. *Endocr Relat Cancer* 2006;13:279–92.
58. Siiteri PK. Adipose tissue as a source of hormones. *Am J Clin Nutr* 1987;45:277–82.
59. Key T, Appleby P, Barnes J, Reeves G. Endogenous sex hormones and breast cancer in postmenopausal women: reanalysis of nine prospective studies. *J Natl Cancer Inst* 2002;94:606–16.
60. Figueroa A, Going SB, Milliken LA, et al. Effects of exercise training and hormone replacement therapy on lean and fat mass in postmenopausal women. *J Gerontol A Biol Sci Med Sci* 2003;58: M266–70.
61. Cauley JA, Gutai JP, Kuller LH, LeDonne D, Powell JG. The epidemiology of serum sex hormones in postmenopausal women. *Am J Epidemiol* 1989;129:1120–31.
62. Chan MF, Dowsett M, Folkler E, et al. Usual physical activity and endogenous sex hormones in postmenopausal women: the European Prospective Investigation into Cancer - Norfolk population study. *Cancer Epidemiol Biomarkers Prev* 2007;16:900–5.
63. Madigan MP, Troisi R, Potischman N, Dorgan JF, Brinton LA, Hoover RN. Serum hormone levels in relation to reproductive and lifestyle factors in postmenopausal women (United States). *Cancer Causes Control* 1998;9:199–207.
64. Verkasalo PK, Thomas HV, Appleby PN, Davey GK, Key TJ. Circulating levels of sex hormones and their relation to risk factors for breast cancer: a cross-sectional study in 1092 pre- and postmenopausal women (United Kingdom). *Cancer Causes Control* 2001;12:47–59.
65. Nicolas Diaz-Chico B, German RF, Gonzalez A, et al. Androgens and androgen receptors in breast cancer. *J Steroid Biochem Mol Biol* 2007; 105:1–15.
66. Lillie EO, Bernstein L, Ursin G. The role of androgens and polymorphisms in the androgen receptor in the epidemiology of breast cancer. *Breast Cancer Res* 2003;5:164–73.
67. Recchione C, Venturelli E, Manzari A, Cavalleri A, Martinetti A, Secreto G. Testosterone, dihydrotestosterone and oestradiol levels in postmenopausal breast cancer tissues. *J Steroid Biochem Mol Biol* 1995;52:541–6.
68. Suzuki T, Miki Y, Moriya T, et al. In situ production of sex steroids in human breast carcinoma. *Med Mol Morphol* 2007;40:121–7.
69. Liao DJ, Dickson RB. Roles of androgens in the development, growth, and carcinogenesis of the mammary gland. *J Steroid Biochem Mol Biol* 2002;80:175–89.
70. Zeleniuch-Jacquotte A, Shore RE, Koenig KL, et al. Postmenopausal levels of oestrogen, androgen, and SHBG and breast cancer: long-term results of a prospective study. *Br J Cancer* 2004;90:153–9.
71. Rinaldi S, Key TJ, Peeters PH, et al. Anthropometric measures, endogenous sex steroids and breast cancer risk in postmenopausal women: a study within the EPIC cohort. *Int J Cancer* 2006; 118:2832–9.
72. Kaaks R, Rinaldi S, Key TJ, et al. Postmenopausal serum androgens, oestrogens and breast cancer risk: the European prospective investigation into cancer and nutrition. *Endocr Relat Cancer* 2005; 12:1071–82.
73. Kaaks R. Nutrition, hormones, and breast cancer: is insulin the missing link? *Cancer Causes Control* 1996;7:605–25.
74. Kaaks R, Lukanova A. Energy balance and cancer: the role of insulin and insulin-like growth factor-I. *Proc Nutr Soc* 2001;60:91–106.
75. Stephenson GD, Rose DP. Breast cancer and obesity: an update. *Nutr Cancer* 2003;45:1–16.
76. Ivy JL. Role of exercise training in the prevention and treatment of insulin resistance and non-insulin-dependent diabetes mellitus. *Sports Med* 1997;24:321–36.
77. Perez-Martin A, Raynaud E, Mercier J. Insulin resistance and associated metabolic abnormalities in muscle: effects of exercise. *Obes Rev* 2001;2:47–59.
78. Dishman RK, Washburn RA, Heath GW. Physical activity and diabetes. *Physical activity epidemiology*. Champaign (IL): Human Kinetics; 2004. p.191–207.
79. Reaven GM. Metabolic syndrome: definition, relationship to insulin resistance, and clinical utility. In: Shils ME, Shike M, Ross AC, Caballero B, Cousins RJ, editors. *Modern nutrition in health and disease*. Philadelphia: Lippincott Williams & Wilkins; 2006. p.1004–12.
80. Harris NS, Winter WE. The chemical pathology of insulin resistance and the metabolic syndrome. *MLO Med Lab Obs* 2004;36:20, 22–5.
81. Roberts CK, Barnard RJ. Effects of exercise and diet on chronic disease. *J Appl Physiol* 2005;98:3–30.
82. Despres JP, Lemieux I. Abdominal obesity and metabolic syndrome. *Nature* 2006;444:881–7.
83. Larsson SC, Mantzoros CS, Wolk A. Diabetes mellitus and risk of breast cancer: a meta-analysis. *Int J Cancer* 2007;121:856–62.
84. Bonser AM, Garcia-Webb P. C-peptide measurement: methods and clinical utility. *Crit Rev Clin Lab Sci* 1984;19:297–352.
85. Lawlor DA, Smith GD, Ebrahim S. Hyperinsulinaemia and increased risk of breast cancer: findings from the British Women's Heart and Health Study. *Cancer Causes Control* 2004;15:267–75.
86. Han C, Zhang HT, Du L, et al. Serum levels of leptin, insulin, and lipids in relation to breast cancer in China. *Endocrine* 2005;26: 19–24.
87. Hirose K, Toyama T, Iwata H, Takezaki T, Hamajima N, Tajima K. Insulin, insulin-like growth factor-I and breast cancer risk in Japanese women. *Asian Pac J Cancer Prev* 2003;4:239–46.
88. Garmendia ML, Pereira A, Alvarado ME, Atalah E. Relation between insulin resistance and breast cancer among Chilean women. *Ann Epidemiol* 2007;17:403–9.
89. Yang G, Lu G, Jin F, et al. Population-based, case-control study of blood C-peptide level and breast cancer risk. *Cancer Epidemiol Biomarkers Prev* 2001;10:1207–11.
90. Bruning PF, Bonfrer JM, van Noord PA, Hart AA, Jong-Bakker M, Nooijen WJ. Insulin resistance and breast-cancer risk. *Int J Cancer* 1992;52:511–6.
91. Verheus M, Peeters PH, Rinaldi S, et al. Serum C-peptide levels and breast cancer risk: results from the European Prospective Investigation into Cancer and Nutrition (EPIC). *Int J Cancer* 2006;119:659–67.
92. Jernstrom H, Barrett-Connor E. Obesity, weight change, fasting insulin, proinsulin, C-peptide, and insulin-like growth factor-1 levels in women with and without breast cancer: the Rancho Bernardo Study. *J Womens Health Genet Based Med* 1999;8:1265–72.
93. Gamayunova VB, Bobrov Y, Tsyrlina EV, Evtushenko TP, Bernstein LM. Comparative study of blood insulin levels in breast and endometrial cancer patients. *Neoplasma* 1997;44:123–6.
94. Muti P, Quattrin T, Grant BJ, et al. Fasting glucose is a risk factor for breast cancer: a prospective study. *Cancer Epidemiol Biomarkers Prev* 2002;11:1361–8.
95. Manjer J, Kaaks R, Riboli E, Berglund G. Risk of breast cancer in relation to anthropometry, blood pressure, blood lipids and glucose metabolism: a prospective study within the Malmo Preventive Project. *Eur J Cancer Prev* 2001;10:33–42.
96. Keinan-Boker L, Bueno de Mesquita HB, Kaaks R, et al. Circulating levels of insulin-like growth factor I, its binding proteins -1,-2, -3, C-peptide and risk of postmenopausal breast cancer. *Int J Cancer* 2003;106:90–5.
97. Toniolo P, Bruning PF, Akhmedkhanov A, et al. Serum insulin-like growth factor-I and breast cancer. *Int J Cancer* 2000;88:828–32.
98. Schairer C, Hill D, Sturgeon SR, et al. Serum concentrations of IGF-I, IGFBP-3 and c-peptide and risk of hyperplasia and cancer of the breast in postmenopausal women. *Int J Cancer* 2004;108:773–9.
99. Lin J, Ridker PM, Rifai N, et al. A prospective study of hemoglobin A1c concentrations and risk of breast cancer in women. *Cancer Res* 2006;66:2869–75.
100. Gabbay KH. Glycosylated hemoglobin and diabetes mellitus. *Med Clin North Am* 1982;66:1309–15.
101. Albright A, Franz M, Hornsby G, et al. American College of Sports Medicine position stand. Exercise and type 2 diabetes. *Med Sci Sports Exerc* 2000;32:1345–60.
102. Klein S, Sheard NF, Pi-Sunyer X, et al. Weight management through lifestyle modification for the prevention and management of type 2 diabetes: rationale and strategies: a statement of the American Diabetes Association, the North American Association for the Study of Obesity, and the American Society for Clinical Nutrition. *Diabetes Care* 2004;27:2067–73.
103. Dela F, Kjaer M. Resistance training, insulin sensitivity and muscle function in the elderly. *Essays Biochem* 2006;42:75–88.
104. Clark DO. Physical activity efficacy and effectiveness among older adults and minorities. *Diabetes Care* 1997;20:1176–82.
105. Ryan AS. Insulin resistance with aging: effects of diet and exercise. *Sports Med* 2000;30:327–46.
106. Asikainen TM, Miilunpalo S, Kukkonen-Harjula K, et al. Walking trials in postmenopausal women: effect of low doses of exercise and exercise fractionation on coronary risk factors. *Scand J Med Sci Sports* 2003;13:284–92.

107. DiPietro L, Dziura J, Yeckel CW, Neuffer PD. Exercise and improved insulin sensitivity in older women: evidence of the enduring benefits of higher intensity training. *J Appl Physiol* 2006;100:142–9.
108. Cuff DJ, Meneilly GS, Martin A, Ignaszewski A, Tildesley HD, Frohlich JJ. Effective exercise modality to reduce insulin resistance in women with type 2 diabetes. *Diabetes Care* 2003;26:2977–82.
109. Waki H, Tontonoz P. Endocrine functions of adipose tissue. *Annu Rev Pathol* 2007;2:31–56.
110. Trayhurn P, Bing C, Wood IS. Adipose tissue and adipokines - energy regulation from the human perspective. *J Nutr* 2006;136:1935–9S.
111. Vona-Davis L, Rose DP. Adipokines as endocrine, paracrine, and autocrine factors in breast cancer risk and progression. *Endocr Relat Cancer* 2007;14:189–206.
112. Lee YH, Pratley RE. The evolving role of inflammation in obesity and the metabolic syndrome. *Curr Diab Rep* 2005;5:70–5.
113. Priest EL, Church TS. Obesity, cytokines, and other inflammatory markers. In: McTieman A, editor. *Cancer prevention and management through exercise and weight control*. CRC Taylor & Francis; 2006. p. 317–27.
114. Aggarwal BB, Shishodia S, Sandur SK, Pandey MK, Sethi G. Inflammation and cancer: how hot is the link? *Biochem Pharmacol* 2006;72:1605–21.
115. Das UN. Metabolic syndrome X: an inflammatory condition? *Curr Hypertens Rep* 2004;6:66–73.
116. Coussens LM, Werb Z. Inflammation and cancer. *Nature* 2002;420:860–7.
117. Vona-Davis L, Howard-McNatt M, Rose DP. Adiposity, type 2 diabetes and the metabolic syndrome in breast cancer. *Obes Rev* 2007;8:395–408.
118. You T, Berman DM, Ryan AS, Nicklas BJ. Effects of hypocaloric diet and exercise training on inflammation and adipocyte lipolysis in obese postmenopausal women. *J Clin Endocrinol Metab* 2004;89:1739–46.
119. Hamer M. The relative influences of fitness and fatness on inflammatory factors. *Prev Med* 2007;44:3–11.
120. Finck BN, Johnson RW. Tumor necrosis factor- α regulates secretion of the adipocyte-derived cytokine, leptin. *Microsc Res Tech* 2000;50:209–15.
121. Zhang Y, Proenca R, Maffei M, Barone M, Leopold L, Friedman JM. Positional cloning of the mouse obese gene and its human homologue. *Nature* 1994;372:425–32.
122. Garofalo C, Surmacz E. Leptin and cancer. *J Cell Physiol* 2006;207:12–22.
123. Sulikowska M, Golaszewska J, Wincewicz A, Koda M, Baltaziak M, Sulikowski S. Leptin – from regulation of fat metabolism to stimulation of breast cancer growth. *Pathol Oncol Res* 2006;12:69–72.
124. Surmacz E. Obesity hormone leptin: a new target in breast cancer? *Breast Cancer Res* 2007;9:301.
125. Hou WK, Xu YX, Yu T, et al. Adipocytokines and breast cancer risk. *Chin Med J (Engl)* 2007;120:1592–6.
126. Ozet A, Arpacı F, Yılmaz MI, et al. Effects of tamoxifen on the serum leptin level in patients with breast cancer. *Jpn J Clin Oncol* 2001;31:424–7.
127. Kohrt WM, Landt M, Birge SJ, Jr. Serum leptin levels are reduced in response to exercise training, but not hormone replacement therapy, in older women. *J Clin Endocrinol Metab* 1996;81:3980–5.
128. Hayase H, Nomura S, Abe T, Izawa T. Relation between fat distributions and several plasma adipocytokines after exercise training in premenopausal and postmenopausal women. *J Physiol Anthropol Appl Human Sci* 2002;21:105–13.
129. Giannopoulou I, Fernhall B, Carhart R, et al. Effects of diet and/or exercise on the adipocytokine and inflammatory cytokine levels of postmenopausal women with type 2 diabetes. *Metabolism* 2005;54:866–75.
130. Hulver MW, Houmard JA. Plasma leptin and exercise: recent findings. *Sports Med* 2003;33:473–82.
131. Kraemer RR, Chu H, Castracane VD. Leptin and exercise. *Exp Biol Med (Maywood)* 2002;227:701–8.
132. Scherer PE, Williams S, Fogliano M, Baldini G, Lodish HF. A novel serum protein similar to C1q, produced exclusively in adipocytes. *J Biol Chem* 1995;270:26746–9.
133. Trujillo ME, Scherer PE. Adiponectin – journey from an adipocyte secretory protein to biomarker of the metabolic syndrome. *J Intern Med* 2005;257:167–75.
134. Trayhurn P, Bing C. Appetite and energy balance signals from adipocytes. *Philos Trans R Soc Lond B Biol Sci* 2006;361:1237–49.
135. Arita Y, Kihara S, Ouchi N, et al. Paradoxical decrease of an adipose-specific protein, adiponectin, in obesity. *Biochem Biophys Res Commun* 1999;257:79–83.
136. Hotta K, Funahashi T, Arita Y, et al. Plasma concentrations of a novel, adipose-specific protein, adiponectin, in type 2 diabetic patients. *Arterioscler Thromb Vasc Biol* 2000;20:1595–9.
137. Matsuzawa Y. Adiponectin: Identification, physiology and clinical relevance in metabolic and vascular disease. *Atheroscler Suppl* 2005;6:7–14.
138. Nicklas BJ, You T, Pahor M. Behavioural treatments for chronic systemic inflammation: effects of dietary weight loss and exercise training. *CMAJ* 2005;172:1199–209.
139. Bruun JM, Lihn AS, Verdich C, et al. Regulation of adiponectin by adipose tissue-derived cytokines: *in vivo* and *in vitro* investigations in humans. *Am J Physiol Endocrinol Metab* 2003;285:E527–33.
140. Chandran M, Phillips SA, Ciaraldi T, Henry RR. Adiponectin: more than just another fat cell hormone? *Diabetes Care* 2003;26:2442–50.
141. Kaur T, Zhang ZF. Obesity, breast cancer and the role of adipocytokines. *Asian Pac J Cancer Prev* 2005;6:547–52.
142. Ronti T, Lupattelli G, Mannarino E. The endocrine function of adipose tissue: an update. *Clin Endocrinol (Oxf)* 2006;64:355–65.
143. Wang Y, Lam KS, Xu A. Adiponectin as a negative regulator in obesity-related mammary carcinogenesis. *Cell Res* 2007;17:280–2.
144. Frayn KN. Obesity and metabolic disease: is adipose tissue the culprit? *Proc Nutr Soc* 2005;64:7–13.
145. Pittas AG, Joseph NA, Greenberg AS. Adipocytokines and insulin resistance. *J Clin Endocrinol Metab* 2004;89:447–52.
146. Weyer C, Funahashi T, Tanaka S, et al. Hypoadiponectinemia in obesity and type 2 diabetes: close association with insulin resistance and hyperinsulinemia. *J Clin Endocrinol Metab* 2001;86:1930–5.
147. Ring-Dimitriou S, Paulweber B, von Duvillard SP, et al. The effect of physical activity and physical fitness on plasma adiponectin in adults with predisposition to metabolic syndrome. *Eur J Appl Physiol* 2006;98:472–81.
148. Simpson KA, Singh MA. Effects of exercise on adiponectin: a systematic review. *Obesity (Silver Spring)* 2008;16:241–56.
149. Kraemer RR, Castracane VD. Exercise and humoral mediators of peripheral energy balance: ghrelin and adiponectin. *Exp Biol Med (Maywood)* 2007;232:184–94.
150. Montesano R, Soulie P, Eble JA, Carrozzino F. Tumor necrosis factor α confers an invasive, transformed phenotype on mammary epithelial cells. *J Cell Sci* 2005;118:3487–500.
151. Balkwill F. TNF- α in promotion and progression of cancer. *Cancer Metastasis Rev* 2006;25:409–16.
152. Szlosarek P, Charles KA, Balkwill FR. Tumor necrosis factor- α as a tumour promoter. *Eur J Cancer* 2006;42:745–50.
153. Hotamisligil GS, Arner P, Caro JF, Atkinson RL, Spiegelman BM. Increased adipose tissue expression of tumor necrosis factor- α in human obesity and insulin resistance. *J Clin Invest* 1995;95:2409–15.
154. Kern PA, Saghizadeh M, Ong JM, Bosch RJ, Deem R, Simsolo RB. The expression of tumor necrosis factor in human adipose tissue. Regulation by obesity, weight loss, and relationship to lipoprotein lipase. *J Clin Invest* 1995;95:2111–9.
155. Weisberg SP, McCann D, Desai M, Rosenbaum M, Leibel RL, Ferrante AW, Jr. Obesity is associated with macrophage accumulation in adipose tissue. *J Clin Invest* 2003;112:1796–808.
156. Il'yasova D, Colbert LH, Harris TB, et al. Circulating levels of inflammatory markers and cancer risk in the health aging and body composition cohort. *Cancer Epidemiol Biomarkers Prev* 2005;14:2413–8.
157. Elosua R, Bartali B, Ordovas JM, Corsi AM, Lauretani F, Ferrucci L. Association between physical activity, physical performance, and inflammatory biomarkers in an elderly population: the InCHIANTI study. *J Gerontol A Biol Sci Med Sci* 2005;60:760–7.
158. McFarlin BK, Flynn MG, Campbell WW, Stewart LK, Timmerman KL. TLR4 is lower in resistance-trained older women and related to inflammatory cytokines. *Med Sci Sports Exerc* 2004;36:1876–83.
159. Yudkin JS, Kumari M, Humphries SE, Mohamed-Ali V. Inflammation, obesity, stress and coronary heart disease: is interleukin-6 the link? *Atherosclerosis* 2000;148:209–14.
160. Shephard RJ. Cytokine responses to physical activity, with particular reference to IL-6: sources, actions, and clinical implications. *Crit Rev Immunol* 2002;22:165–82.
161. Robinson LE, Graham TE. Metabolic syndrome, a cardiovascular disease risk factor: role of adipocytokines and impact of diet and physical activity. *Can J Appl Physiol* 2004;29:808–29.
162. Kern PA, Ranganathan S, Li C, Wood L, Ranganathan G. Adipose tissue tumor necrosis factor and interleukin-6 expression in human obesity and insulin resistance. *Am J Physiol Endocrinol Metab* 2001;280:E745–51.
163. Fried SK, Bunkin DA, Greenberg AS. Omental and subcutaneous adipose tissues of obese subjects release interleukin-6: depot difference and regulation by glucocorticoid. *J Clin Endocrinol Metab* 1998;83:847–50.

164. Bruunsgaard H. Physical activity and modulation of systemic low-level inflammation. *J Leukoc Biol* 2005;78:819–35.
165. Petersen AM, Pedersen BK. The anti-inflammatory effect of exercise. *J Appl Physiol* 2005;98:1154–62.
166. Purohit A, Reed MJ. Regulation of estrogen synthesis in postmenopausal women. *Steroids* 2002;67:979–83.
167. Asgeirsson KS, Olafsdottir K, Jonasson JG, Ogmundsdottir HM. The effects of IL-6 on cell adhesion and e-cadherin expression in breast cancer. *Cytokine* 1998;10:720–8.
168. Jiang XP, Yang DC, Elliott RL, Head JF. Reduction in serum IL-6 after vaccination of breast cancer patients with tumour-associated antigens is related to estrogen receptor status. *Cytokine* 2000;12:458–65.
169. Kozlowski L, Zakrzewska I, Tokajuk P, Wojtukiewicz MZ. Concentration of interleukin-6 (IL-6), interleukin-8 (IL-8) and interleukin-10 (IL-10) in blood serum of breast cancer patients. *Rocz Akad Med Białymst* 2003;48:82–4.
170. Jablonska E, Kiluk M, Markiewicz W, Piotrowski L, Grabowska Z, Jablonski J. TNF- α , IL-6 and their soluble receptor serum levels and secretion by neutrophils in cancer patients. *Arch Immunol Ther Exp (Warsz)* 2001;49:63–9.
171. Hussein MZ, Al FA, Abdel BI, Attia O. Serum IL-6 and IL-12 levels in breast cancer patients. *Egypt J Immunol* 2004;11:165–70.
172. Knupfer H, Preiss R. Significance of interleukin-6 (IL-6) in breast cancer (review). *Breast Cancer Res Treat* 2007;102:129–35.
173. Taaffe DR, Harris TB, Ferrucci L, Rowe J, Seeman TE. Cross-sectional and prospective relationships of interleukin-6 and C-reactive protein with physical performance in elderly persons: MacArthur studies of successful aging. *J Gerontol A Biol Sci Med Sci* 2000;55:M709–15.
174. Devaraj S, Kasim-Karakas S, Jialal I. The effect of weight loss and dietary fatty acids on inflammation. *Curr Atheroscler Rep* 2006;8:477–86.
175. Forouhi NG, Sattar N, McKeigue PM. Relation of C-reactive protein to body fat distribution and features of the metabolic syndrome in Europeans and South Asians. *Int J Obes Relat Metab Disord* 2001;25:1327–31.
176. Aronson D, Bartha P, Zinder O, et al. Obesity is the major determinant of elevated C-reactive protein in subjects with the metabolic syndrome. *Int J Obes Relat Metab Disord* 2004;28:674–9.
177. Fogarty AW, Glancy C, Jones S, Lewis SA, McKeever TM, Britton JR. A prospective study of weight change and systemic inflammation over 9 y. *Am J Clin Nutr* 2008;87:30–5.
178. Haffner SM. Insulin resistance, inflammation, and the prediabetic state. *Am J Cardiol* 2003;92:18–26J.
179. Kasapis C, Thompson PD. The effects of physical activity on serum C-reactive protein and inflammatory markers: a systematic review. *J Am Coll Cardiol* 2005;45:1563–9.
180. Plaisance EP, Grandjean PW. Physical activity and high-sensitivity C-reactive protein. *Sports Med* 2006;36:443–58.
181. Selvin E, Paynter NP, Erlinger TP. The effect of weight loss on C-reactive protein: a systematic review. *Arch Intern Med* 2007;167:31–9.
182. Kelley GA, Kelley KS. Effects of aerobic exercise on C-reactive protein, body composition, and maximum oxygen consumption in adults: a meta-analysis of randomized controlled trials. *Metabolism* 2006;55:1500–7.
183. Fairey AS, Courneya KS, Field CJ, et al. Effect of exercise training on C-reactive protein in postmenopausal breast cancer survivors: a randomized controlled trial. *Brain Behav Immun* 2005;19:381–8.
184. Ballard-Barbash R. Obesity, weight change, and breast cancer incidence. In: McTiernan A, editor. *Cancer prevention and management through exercise and weight control*. CRC Press; 2006. p. 219–32.
185. Holt HB, Wild SH, Wareham N, et al. Differential effects of fatness, fitness and physical activity energy expenditure on whole-body, liver and fat insulin sensitivity. *Diabetologia* 2007;50:1698–706.
186. Eng SM, Gammon MD, Terry MB, et al. Body size changes in relation to postmenopausal breast cancer among women on Long Island, New York. *Am J Epidemiol* 2005;162:229–37.
187. Feigelson HS, Jonas CR, Teras LR, Thun MJ, Calle EE. Weight gain, body mass index, hormone replacement therapy, and postmenopausal breast cancer in a large prospective study. *Cancer Epidemiol Biomarkers Prev* 2004;13:220–4.
188. Lahmann PH, Hoffmann K, Allen N, et al. Body size and breast cancer risk: findings from the European Prospective Investigation into Cancer and Nutrition (EPIC). *Int J Cancer* 2004;111:762–71.
189. Morimoto LM, White E, Chen Z, et al. Obesity, body size, and risk of postmenopausal breast cancer: the Women's Health Initiative (United States). *Cancer Causes Control* 2002;13:741–51.
190. Friedenreich CM, Courneya KS, Bryant HE. Case-control study of anthropometric measures and breast cancer risk. *Int J Cancer* 2002;99:445–52.
191. Suzuki R, Rylander-Rudqvist T, Ye W, Saji S, Wolk A. Body weight and postmenopausal breast cancer risk defined by estrogen and progesterone receptor status among Swedish women: a prospective cohort study. *Int J Cancer* 2006;119:1683–9.
192. Patel AV, Calle EE, Bernstein L, Wu AH, Thun MJ. Recreational physical activity and risk of postmenopausal breast cancer in a large cohort of US women. *Cancer Causes Control* 2003;14:519–29.
193. Slattery ML, Edwards S, Murtaugh MA, et al. Physical activity and breast cancer risk among women in the southwestern United States. *Ann Epidemiol* 2007;17:342–53.
194. Tehard B, Friedenreich CM, Oppert JM, Clavel-Chapelon F. Effect of physical activity on women at increased risk of breast cancer: results from the E3N cohort study. *Cancer Epidemiol Biomarkers Prev* 2006;15:57–64.
195. Friedenreich CM, Bryant HE, Courneya KS. Case-control study of lifetime physical activity and breast cancer risk. *Am J Epidemiol* 2001;154:336–47.
196. Ridker PM, Hennekens CH, Rifai N, Buring JE, Manson JE. Hormone replacement therapy and increased plasma concentration of C-reactive protein. *Circulation* 1999;100:713–6.
197. Prasad K. C-reactive protein (CRP)-lowering agents. *Cardiovasc Drug Rev* 2006;24:33–50.
198. Chioloro A, Faeh D, Paccaud F, Cornuz J. Consequences of smoking for body weight, body fat distribution, and insulin resistance. *Am J Clin Nutr* 2008;87:801–9.
199. O'Keefe JH, Gheewala NM, O'Keefe JO. Dietary strategies for improving post-prandial glucose, lipids, inflammation, and cardiovascular health. *J Am Coll Cardiol* 2008;51:249–55.
200. Bullo M, Casas-Agustench P, migo-Correig P, Aranceta J, Salas-Salvado J. Inflammation, obesity and comorbidities: the role of diet. *Public Health Nutr* 2007;10:1164–72.
201. Althuis MD, Fergenbaum JH, Garcia-Closas M, Brinton LA, Madigan MP, Sherman ME. Etiology of hormone receptor-defined breast cancer: a systematic review of the literature. *Cancer Epidemiol Biomarkers Prev* 2004;13:1558–68.
202. Macinnis RJ, English DR, Gertig DM, Hopper JL, Giles GG. Body size and composition and risk of postmenopausal breast cancer. *Cancer Epidemiol Biomarkers Prev* 2004;13:2117–25.
203. Missmer SA, Eliassen AH, Barbieri RL, Hankinson SE. Endogenous estrogen, androgen, and progesterone concentrations and breast cancer risk among postmenopausal women. *J Natl Cancer Inst* 2004;96:1856–65.
204. Tian YF, Chu CH, Wu MH, et al. Anthropometric measures, plasma adiponectin, and breast cancer risk. *Endocr Relat Cancer* 2007;14:669–77.
205. Siemes C, Visser LE, Coebergh JW, et al. C-reactive protein levels, variation in the C-reactive protein gene, and cancer risk: the Rotterdam Study. *J Clin Oncol* 2006;24:5216–22.
206. Heaney ML, Golde DW. Soluble receptors in human disease. *J Leukoc Biol* 1998;64:135–46.
207. Lear SA, Humphries KH, Kohli S, Birmingham CL. The use of BMI and waist circumference as surrogates of body fat differs by ethnicity. *Obesity (Silver Spring)* 2007;15:2817–24.
208. Ode JJ, Pivarnik JM, Reeves MJ, Knous JL. Body mass index as a predictor of percent fat in college athletes and nonathletes. *Med Sci Sports Exerc* 2007;39:403–9.
209. Heinrich KM, Jitnarin N, Suminski RR, et al. Obesity classification in military personnel: a comparison of body fat, waist circumference, and body mass index measurements. *Mil Med* 2008;173:67–73.
210. Blew RM, Sardinha LB, Milliken LA, et al. Assessing the validity of body mass index standards in early postmenopausal women. *Obes Res* 2002;10:799–808.
211. Kaye SA, Folsom AR, Soler JT, Prineas RJ, Potter JD. Associations of body mass and fat distribution with sex hormone concentrations in postmenopausal women. *Int J Epidemiol* 1991;20:151–6.
212. Rendell M, Hulthén UL, Tornquist C, Groop L, Mattiasson I. Relationship between abdominal fat compartments and glucose and lipid metabolism in early postmenopausal women. *J Clin Endocrinol Metab* 2001;86:744–9.
213. Van Pelt RE, Evans EM, Schechtman KB, Ehsani AA, Kohrt WM. Waist circumference vs body mass index for prediction of disease risk in postmenopausal women. *Int J Obes Relat Metab Disord* 2001;25:1183–8.
214. Manns PJ, Williams DP, Snow CM, Wander RC. Physical activity, body fat, and serum C-reactive protein in postmenopausal women with and without hormone replacement. *Am J Hum Biol* 2003;15:91–100.
215. Ross R, Janssen I. Is abdominal fat preferentially reduced in response to exercise-induced weight loss? *Med Sci Sports Exerc* 1999;31:5568–72.
216. Giannopoulou I, Ploutz-Snyder LL, Carhart R, et al. Exercise is

- required for visceral fat loss in postmenopausal women with type 2 diabetes. *J Clin Endocrinol Metab* 2005;90:1511–8.
217. McTiernan PG, Kusminski CM, Kumar S. Resistin. *Curr Opin Lipidol* 2006;17:170–5.
 218. Lazar MA. Resistin- and Obesity-associated metabolic diseases. *Horm Metab Res* 2007;39:710–6.
 219. Kang JH, Yu BY, Youn DS. Relationship of serum adiponectin and resistin levels with breast cancer risk. *J Korean Med Sci* 2007;22:117–21.
 220. Rokling-Andersen MH, Reseland JE, Veierod MB, et al. Effects of long-term exercise and diet intervention on plasma adipokine concentrations. *Am J Clin Nutr* 2007;86:1293–301.
 221. Patel AV, Bernstein L. Physical activity and cancer incidence: breast cancer. In: McTiernan A, editor. *Cancer prevention and management through exercise and weight control*. Taylor & Francis Group; 2006. p. 49–74.
 222. Grodin JM, Siiteri PK, MacDonald PC. Source of estrogen production in postmenopausal women. *J Clin Endocrinol Metab* 1973;36:207–14.
 223. Kuonen-Boumeester V, Van der Kwast TH, van Putten WL, Claassen C, van OB, Henzen-Logmans SC. Immunohistochemical determination of androgen receptors in relation to oestrogen and progesterone receptors in female breast cancer. *Int J Cancer* 1992;52:581–4.
 224. Moinfar F, Okcu M, Tsybrovskyy O, et al. Androgen receptors frequently are expressed in breast carcinomas: potential relevance to new therapeutic strategies. *Cancer* 2003;98:703–11.
 225. von Schoultz B. Androgens and the breast. *Maturitas* 2007;57:47–9.
 226. Birrell SN, Bentel JM, Hickey TE, et al. Androgens induce divergent proliferative responses in human breast cancer cell lines. *J Steroid Biochem Mol Biol* 1995;52:459–67.
 227. Dimitrakakis C, Zhou J, Bondy CA. Androgens and mammary growth and neoplasia. *Fertil Steril* 2002;77 Suppl 4:S26–33.
 228. Corbould AM, Judd SJ, Rodgers RJ. Expression of types 1, 2, and 3 17 β -hydroxysteroid dehydrogenase in subcutaneous abdominal and intra-abdominal adipose tissue of women. *J Clin Endocrinol Metab* 1998;83:187–94.
 229. Pugeat M, Crave JC, Elmidi M, et al. Pathophysiology of sex hormone binding globulin (SHBG): relation to insulin. *J Steroid Biochem Mol Biol* 1991;40:841–9.
 230. Chappell J, Leitner JW, Solomon S, Golovchenko I, Goalstone ML, Draznin B. Effect of insulin on cell cycle progression in MCF-7 breast cancer cells. Direct and potentiating influence. *J Biol Chem* 2001;276:38023–8.
 231. Osborne CK, Bolan G, Monaco ME, Lippman ME. Hormone responsive human breast cancer in long-term tissue culture: effect of insulin. *Proc Natl Acad Sci U S A* 1976;73:4536–40.
 232. van der Burg B, Rutteman GR, Blankenstein MA, de Laat SW, van Zoelen EJ. Mitogenic stimulation of human breast cancer cells in a growth factor-defined medium: synergistic action of insulin and estrogen. *J Cell Physiol* 1988;134:101–8.
 233. Haslam DW, James WP. Obesity. *Lancet* 2005;366:1197–209.
 234. Stoll BA. Nutrition and breast cancer risk: can an effect via insulin resistance be demonstrated? *Breast Cancer Res Treat* 1996;38:239–46.
 235. Sigal RJ, Kenny GP, Wasserman DH, Castaneda-Sceppa C. Physical activity/exercise and type 2 diabetes. *Diabetes Care* 2004;27:2518–39.
 236. Boyapati SM, Shu XO, Gao YT, et al. Correlation of blood sex steroid hormones with body size, body fat distribution, and other known risk factors for breast cancer in post-menopausal Chinese women. *Cancer Causes Control* 2004;15:305–11.
 237. Lamar CA, Dorgan JF, Longcope C, Stanczyk FZ, Falk RT, Stephenson HE, Jr. Serum sex hormones and breast cancer risk factors in postmenopausal women. *Cancer Epidemiol Biomarkers Prev* 2003;12:380–3.
 238. Calle EE, Kaaks R. Overweight, obesity and cancer: epidemiological evidence and proposed mechanisms. *Nat Rev Cancer* 2004;4:579–91.
 239. Kay SJ, Fatarone Singh MA. The influence of physical activity on abdominal fat: a systematic review of the literature. *Obes Rev* 2006;7:183–200.
 240. Walker KZ, Piers LS, Putt RS, Jones JA, O'Dea K. Effects of regular walking on cardiovascular risk factors and body composition in normoglycemic women and women with type 2 diabetes. *Diabetes Care* 1999;22:555–61.
 241. Adly L, Hill D, Sherman ME, et al. Serum concentrations of estrogens, sex hormone-binding globulin, and androgens and risk of breast cancer in postmenopausal women. *Int J Cancer* 2006;119:2402–7.
 242. Yu H, Shu XO, Shi R, et al. Plasma sex steroid hormones and breast cancer risk in Chinese women. *Int J Cancer* 2003;105:92–7.
 243. Manjer J, Johansson R, Berglund G, et al. Postmenopausal breast cancer risk in relation to sex steroid hormones, prolactin and SHBG (Sweden). *Cancer Causes Control* 2003;14:599–607.
 244. Beattie MS, Costantino JP, Cummings SR, et al. Endogenous sex hormones, breast cancer risk, and tamoxifen response: an ancillary study in the NSABP Breast Cancer Prevention Trial (P-1). *J Natl Cancer Inst* 2006;98:110–5.
 245. Mantzoros C, Petridou E, Dessypris N, et al. Adiponectin and breast cancer risk. *J Clin Endocrinol Metab* 2004;89:1102–7.
 246. Petridou E, Papadiamantis Y, Markopoulos C, Spanos E, Dessypris N, Trichopoulos D. Leptin and insulin growth factor I in relation to breast cancer (Greece). *Cancer Causes Control* 2000;11:383–8.
 247. Woo HY, Park H, Ki CS, Park YL, Bae WG. Relationships among serum leptin, leptin receptor gene polymorphisms, and breast cancer in Korea. *Cancer Lett* 2005;237:137–42.
 248. Sauter ER, Garofalo C, Hewett J, Hewett JE, Morelli C, Surmacz E. Leptin expression in breast nipple aspirate fluid (NAF) and serum is influenced by body mass index (BMI) but not by the presence of breast cancer. *Horm Metab Res* 2004;36:336–40.
 249. Stattin P, Soderberg S, Biessy C, et al. Plasma leptin and breast cancer risk: a prospective study in northern Sweden. *Breast Cancer Res Treat* 2004;86:191–6.
 250. Miyoshi Y, Funahashi T, Kihara S, et al. Association of serum adiponectin levels with breast cancer risk. *Clin Cancer Res* 2003;9:5699–704.
 251. Korner A, Pazaitou-Panayiotou K, Kelesidis T, et al. Total and high-molecular-weight adiponectin in breast cancer: *in vitro* and *in vivo* studies. *J Clin Endocrinol Metab* 2007;92:1041–8.
 252. Tworoger SS, Eliassen AH, Kelesidis T, et al. Plasma adiponectin concentrations and risk of incident breast cancer. *J Clin Endocrinol Metab* 2007;92:1510–6.
 253. Hankinson SE, Willett WC, Manson JE, et al. Plasma sex steroid hormone levels and risk of breast cancer in postmenopausal women. *J Natl Cancer Inst* 1998;90:1292–9.
 254. Tamimi RM, Byrne C, Colditz GA, Hankinson SE. Endogenous hormone levels, mammographic density, and subsequent risk of breast cancer in postmenopausal women. *J Natl Cancer Inst* 2007;99:1178–87.
 255. McTiernan A, Wu L, Chen C, et al. Relation of BMI and physical activity to sex hormones in postmenopausal women. *Obesity (Silver Spring)* 2006;14:1662–77.
 256. Newcomb PA, Klein R, Klein BE, et al. Association of dietary and life-style factors with sex hormones in postmenopausal women. *Epidemiology* 1995;6:318–21.
 257. Nelson ME, Meredith CN, Dawson-Hughes B, Evans WJ. Hormone and bone mineral status in endurance-trained and sedentary postmenopausal women. *J Clin Endocrinol Metab* 1988;66:927–33.
 258. Nagata C, Shimizu H, Takami R, Hayashi M, Takeda N, Yasuda K. Relations of insulin resistance and serum concentrations of estradiol and sex hormone-binding globulin to potential breast cancer risk factors. *Jpn J Cancer Res* 2000;91:948–53.
 259. Nagata C, Kabuto M, Takatsuka N, Shimizu H. Associations of alcohol, height, and reproductive factors with serum hormone concentrations in postmenopausal Japanese women. Steroid hormones in Japanese postmenopausal women. *Breast Cancer Res Treat* 1997;44:235–41.
 260. Hakkinen K, Pakarinen A. Serum hormones and strength development during strength training in middle-aged and elderly males and females. *Acta Physiol Scand* 1994;150:211–9.
 261. Wu F, Ames R, Evans MC, France JT, Reid IR. Determinants of sex hormone-binding globulin in normal postmenopausal women. *Clin Endocrinol (Oxf)* 2001;54:81–7.
 262. Goodman-Gruen D, Barrett-Connor E. Sex hormone-binding globulin and glucose tolerance in postmenopausal women. The Rancho Bernardo Study. *Diabetes Care* 1997;20:645–9.
 263. Caballero MJ, Maynar M. Effects of physical exercise on sex hormone binding globulin, high density lipoprotein cholesterol, total cholesterol and triglycerides in postmenopausal women. *Endocr Res* 1992;18:261–79.
 264. Ryan AS, Pratley RE, Elahi D, Goldberg AP. Changes in plasma leptin and insulin action with resistive training in postmenopausal women. *Int J Obes Relat Metab Disord* 2000;24:27–32.
 265. Okita K, Nishijima H, Murakami T, et al. Can exercise training with weight loss lower serum C-reactive protein levels? *Arterioscler Thromb Vasc Biol* 2004;24:1868–73.
 266. Catalano S, Marsico S, Giordano C, et al. Leptin enhances, via AP-1, expression of aromatase in the MCF-7 cell line. *J Biol Chem* 2003;278:28668–76.
 267. Geisler J, Haynes B, Ekse D, Dowsett M, Lonning PE. Total body aromatization in postmenopausal breast cancer patients is strongly correlated to plasma leptin levels. *J Steroid Biochem Mol Biol* 2007;104:27–34.
 268. Schaffler A, Scholmerich J, Buechler C. Mechanisms of disease: adipokines and breast cancer - endocrine and paracrine mechanisms

- that connect adiposity and breast cancer. *Nat Clin Pract Endocrinol Metab* 2007;3:345–54.
269. Huypens P. Leptin controls adiponectin production via the hypothalamus. *Med Hypotheses* 2007;68:87–90.
270. Yamauchi T, Kamon J, Waki H, et al. The fat-derived hormone adiponectin reverses insulin resistance associated with both lipotrophy and obesity. *Nat Med* 2001;7:941–6.
271. Kelesidis I, Kelesidis T, Mantzoros CS. Adiponectin and cancer: a systematic review. *Br J Cancer* 2006;94:1221–5.
272. Brakenhielm E, Veitonmaki N, Cao R, et al. Adiponectin-induced antiangiogenesis and antitumor activity involve caspase-mediated endothelial cell apoptosis. *Proc Natl Acad Sci U S A* 2004;101:2476–81.
273. Arditi JD, Venihaki M, Karalis KP, Chrousos GP. Antiproliferative effect of adiponectin on MCF7 breast cancer cells: a potential hormonal link between obesity and cancer. *Horm Metab Res* 2007;39:9–13.
274. Rozen F, Zhang J, Pollak M. Antiproliferative action of tumor necrosis factor- α on MCF-7 breast cancer cells is associated with increased insulin-like growth factor binding protein-3 accumulation. *Int J Oncol* 1998;13:865–9.
275. Balkwill F. Tumor necrosis factor or tumor promoting factor? *Cytokine Growth Factor Rev* 2002;13:135–41.
276. Lagathu C, Bastard JP, Auclair M, Maachi M, Capeau J, Caron M. Chronic interleukin-6 (IL-6) treatment increased IL-6 secretion and induced insulin resistance in adipocyte: prevention by rosiglitazone. *Biochem Biophys Res Commun* 2003;311:372–9.
277. Rotter V, Nagaev I, Smith U. Interleukin-6 (IL-6) induces insulin resistance in 3T3-1 adipocytes and is, like IL-8 and tumor necrosis factor- α , overexpressed in human fat cells from insulin-resistant subjects. *J Biol Chem* 2003;278:45777–84.
278. Shamsuzzaman AS, Winnicki M, Wolk R, et al. Independent association between plasma leptin and C-reactive protein in healthy humans. *Circulation* 2004;109:2181–5.
279. McLaughlin T, Abbasi F, Lamendola C, et al. Differentiation between obesity and insulin resistance in the association with C-reactive protein. *Circulation* 2002;106:2908–12.

Physical Activity and Postmenopausal Breast Cancer: Proposed Biologic Mechanisms and Areas for Future Research

Heather K. Neilson, Christine M. Friedenreich, Nigel T. Brockton, et al.

Cancer Epidemiol Biomarkers Prev 2009;18:11-27.

Updated version Access the most recent version of this article at:
<http://cebp.aacrjournals.org/content/18/1/11>

Cited articles This article cites 269 articles, 51 of which you can access for free at:
<http://cebp.aacrjournals.org/content/18/1/11.full#ref-list-1>

Citing articles This article has been cited by 23 HighWire-hosted articles. Access the articles at:
<http://cebp.aacrjournals.org/content/18/1/11.full#related-urls>

E-mail alerts [Sign up to receive free email-alerts](#) related to this article or journal.

Reprints and Subscriptions To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions To request permission to re-use all or part of this article, use this link
<http://cebp.aacrjournals.org/content/18/1/11>.
Click on "Request Permissions" which will take you to the Copyright Clearance Center's (CCC) Rightslink site.