

# Vitamin D Receptor Gene Polymorphisms and Epithelial Ovarian Cancer Risk

Galina Lurie,<sup>1</sup> Lynne R. Wilkens,<sup>1</sup> Pamela J. Thompson,<sup>1</sup> Katharine E. McDuffie,<sup>1</sup> Michael E. Carney,<sup>2</sup> Keith Y. Terada,<sup>2</sup> and Marc T. Goodman<sup>1</sup>

<sup>1</sup>Cancer Epidemiology Program, Cancer Research Center of Hawaii and <sup>2</sup>Department of Obstetrics and Gynecology, John A. Burns School of Medicine, University of Hawaii, Honolulu, Hawaii

## Abstract

Epidemiologic and laboratory studies support a role for the vitamin D endocrine system in ovarian carcinogenesis. The association of ovarian cancer risk with polymorphisms in the *vitamin D receptor* (VDR) gene, including rs10735810 (*FokI*), rs11568820 (*Cdx-2*), rs1544410 (*BsmI*), rs7975232 (*ApaI*), rs731236 (*TaqI*), and *BsmI-ApaI-TaqI* combined genotypes, was examined among 313 women with epithelial ovarian carcinoma and 574 controls. Odds ratios (OR) and 95% confidence intervals (95% CI) were estimated using unconditional logistic regression. The associations of VDR polymorphisms with risk were generally inconsistent across ethnic groups. Among Caucasian women (72 cases, 148 controls), heterozygous and homozygous *ApaI A* allele carriers were at increased ovarian carcinoma risk compared with homozygous carriers of the *ApaI a* allele (OR 2.8, 95% CI 1.2-7.0 and OR 3.4, 95%

CI 1.3-9.1;  $P_{\text{trend}} = 0.02$ ). Caucasian heterozygous carriers of *FokI f* allele were also at increased risk of ovarian carcinoma compared with homozygous carriers of the common allele (OR 2.5, 95% CI 1.3-4.8;  $P_{\text{trend}} = 0.04$ ). Among Japanese women (94 cases, 173 controls), ovarian cancer risk was significantly decreased (OR 0.5, 95% CI 0.3-0.9) among *Cdx-2 A* allele heterozygotes compared with homozygote G allele carriers ( $P_{\text{trend}} = 0.03$ ). Compared with the *bbaATT BsmI-ApaI-TaqI* genotype, *bbaATT* and *BBAAtt* genotypes were associated with increased ovarian cancer risk in Caucasian women (OR 4.2, 95% CI 1.3-13.1 and OR 5.2, 95% CI 1.6-17.5), but not in Japanese women (OR 1.1, 95% CI 0.6-1.9 and OR 2.3, 95% CI:0.4-12.3). This investigation provides some evidence that polymorphisms in the VDR gene might influence ovarian cancer susceptibility. (Cancer Epidemiol Biomarkers Prev 2007;16(12):2566-71)

## Introduction

Evidence from epidemiologic and laboratory studies supports a role for the vitamin D endocrine system in ovarian carcinogenesis. Several ecologic investigations have reported an inverse association between sunlight exposure and age-specific ovarian cancer incidence (1) and mortality (2) rates. Because the major source of vitamin D in humans is sunlight synthesis in the skin, these investigations suggest that vitamin D exposure may reduce the risk for ovarian cancer. In addition, an inverse association of dietary vitamin D and ovarian cancer has been shown in one study (3), although not in other studies (4, 5). The observations that vitamin D and its synthetic analogues inhibit growth and induce apoptosis in ovarian cells in culture and in animal models of ovarian cancer (6-9) provide further plausibility to this hypothesis.

The vitamin D receptor (VDR) is a nuclear transcription factor that belongs to the steroid hormone receptor family (10) and mediates most of the actions of vitamin D

(11). The presence of VDR in the normal ovarian epithelium, in human ovarian tumors, and in human ovarian cancer cell lines has been shown (6, 12, 13). A proposed mechanism for the role of vitamin D in carcinogenesis involves regulation of differentiation and proliferation of cancer cells possibly by influencing cell cycle regulatory proteins (14). Down-regulation of telomerase activity by vitamin D might be another component of the ovarian cancer cells growth suppression (7). Vitamin D might influence ovarian carcinogenesis indirectly through an endocrine pathway. VDR is necessary for full ovarian function through direct effects on estrogen biosynthesis and regulation of aromatase gene expression. VDR-null mice exhibit gonadal insufficiency, reduced aromatase gene expression, low aromatase activity, and elevated serum levels of luteinizing and follicle-stimulating hormones (15). The concept of ovarian cancer as a result of hypergonadotropic hypogonadism was first suggested by Cramer et al. (16, 17).

We hypothesize that common genetic variants in the VDR gene might be associated with ovarian cancer risk. In this study, we selected single-nucleotide polymorphisms (SNP) that have been found to have some functional significance *in vitro* (reviewed by ref. 18). Each of the candidate SNPs were previously found associated with the risk of breast (19-21), colon (22, 23), or prostate cancers (24-26) that are malignancies with possible etiologic similarities to ovarian cancer (27). The rs10735810 (*FokI*) is a coding nonsynonymous SNP in the translational initiation codon that has been reported to

Received 8/16/07; revised 9/20/07; accepted 10/2/07.

**Grant support:** Public Health Service grant R01-CA-58598 and NIH Department of Health and Human Services contracts N01-CN-67001 and N01-CN-25403.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked *advertisement* in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

**Requests for reprints:** Galina Lurie, Cancer Research Center of Hawaii, 1236 Lauhala Street, Room 301C, Honolulu, HI 96813. Phone: 808-586-5866; Fax: 808-586-2984. E-mail: glurie@CRCH.hawaii.edu

Copyright © 2007 American Association for Cancer Research.

doi:10.1158/1055-9965.EPI-07-0753

have functional significance in several *in vitro* studies with C (often called "F" allele) more effective than the T (or "f") allele in transactivation of the vitamin D signal (28-30). The *FokI* polymorphism was considered an independent marker of the *VDR* gene as it has not been reported to be in linkage disequilibrium with any of the other *VDR* polymorphisms (18). The *Cdx-2* (rs11568820) SNP is within a functional binding site of the intestinal specific transcription factor *Cdx-2* in the promoter region of the *VDR* gene and is thought to modulate the transcription of the *VDR* gene (18, 30-32). *BsmI* (rs1544410), *ApaI* (rs7975232) in intron 8, and a silent *TaqI* (rs731236) in exon 9 SNPs, located near the 3' end of the *VDR* gene, have been reported to be in strong linkage disequilibrium. These 3' SNPs do not change the amino acid sequence of the encoded protein; however, they may affect gene expression through regulation of mRNA stability (33).

## Materials and Methods

**Study Design and Population.** This population-based case-control study included 313 cases 18 years of age or older diagnosed with primary histologically confirmed epithelial ovarian carcinoma in 1993 to 2006 and identified through the rapid reporting system of the Hawaii Tumor Registry, which is part of the Surveillance, Epidemiology, and End-Results Program of the National Cancer Institute (34). Control subjects ( $n = 574$ ) were randomly selected from participants in an annual survey of representative households in Hawaii (35) and supplemented with women 65 years of age or older through random sampling from lists obtained from the Health Care Finance Administration and were frequency-matched to cases based on ethnicity and 5-year age groups in an approximate 1:1.5 ratio. Eligibility criteria for controls included age 18 years or older, residency in Hawaii for a minimum of 1 year, no prior history of ovarian cancer, and having at least one intact ovary. The response rate was 64% for cases and 67% for controls. The study protocol was approved by the Institutional Review Board of the University of Hawaii. All study participants signed detailed consent forms before participation.

**Data Collection.** Study participants were interviewed using a structured pretested questionnaire that included sociodemographic and health-related information, menstrual, reproductive and gynecologic histories, and exogenous hormone use (36). Interviewers were uniformly trained and supervised to standardize interviewing and coding techniques. Quality control and performance of the interviewers was monitored by the project coordinator through a repeat interview of a random sample of 15% of subjects on a random 5% of the interview questions.

**Genetic Analysis.** DNA was purified from whole blood using Qiagen midi kits (Qiagen). The DNA samples were analyzed by PCR/RFLP using *TaqMan* (Applied Biosystems). Samples from cases and controls were intermixed on each plate. We included 144 (7.4%) randomly placed blinded samples on each plate to evaluate accuracy and reproducibility. In addition, each 384-well plate included eight non-DNA controls. The concordance rates among duplicates were 98% for *BsmI*

and 100% for all other SNPs. Call rates were 98% to 100% for all polymorphisms.

**Statistical Analysis.** Unconditional multiple logistic regression models were used to calculate odds ratios (OR) and 95% confidence intervals (95% CI) for the association of genotype with ovarian carcinoma. The genotype for each SNP was treated as a nonordered categorical variable to test for heterogeneity and as an ordered categorical variable with three levels (0, 1, and 2) assigned to each genotype to test for a gene-dose effect. Pair-wise linkage disequilibrium ( $D'$ ) and correlation coefficients ( $r^2$ ) were estimated using the HAPLOVIEW program (37). We also created a variable reflecting all possible combinations of *BsmI-ApaI-TaqI* genotypes for each SNP. All genotypes were included into the model, and homozygous carriers of *bbaaTT* genotype (including common alleles of all three SNPs among our study participants) were used as a reference group. The distribution of the following variables was examined by genotype for each SNP among Caucasians and Japanese control subjects separately: age (continuous), education ( $\leq 12$  years for "high school or less," 13-14 years for "post-high school training other than academic college or partial academic college," and  $\geq 15$  years for "college or professional school"); family history of ovarian cancer among first-degree relatives (yes, no); gravidity (0, 1, 2-3, 4, or more); history of a tubal ligation procedure (yes, no); hysterectomy (yes, no); use of contraceptive steroids (yes, no); menopausal status (premenopausal versus postmenopausal); use of menopausal hormones (estrogen alone, progesterone alone, combination of estrogen and progesterone). There was no difference in the distribution of any of these variables by *FokI* and *Cdx-2* genotypes. The distributions of age, education, gravidity, use of contraceptive, and menopausal hormones were different by *ApaI*, *BsmI*, and *TaqI* genotypes in Caucasian women. Education and use of menopausal hormones were distributed differently by genotypes of several of the 3'untranslated region SNPs in Japanese women. Variables that were associated with ovarian cancer risk and which were distributed nonrandomly by *VDR* genotypes in control subjects were included into the statistical models (38). Including all of the covariates did not change associations of *VDR* SNPs with risk. All  $P$  values were based on two-tailed tests.

## Results

The mean age of the study participants was 55.0 years (SD 13.8, range 18-88). Participant characteristics are presented in Table 1. Participant with a family history of ovarian cancer in first-degree relatives had a higher ovarian cancer risk. Education, gravidity, tubal ligation, use of contraceptive steroids, premenopausal status, and the use of menopausal estrogen in combination with progesterone were inversely associated with ovarian cancer risk.

The distributions of the alleles among control subjects within each ethnic group were consistent with Hardy-Weinberg equilibrium, except for the *BsmI* SNP in Japanese and other Asian women ( $P = 0.001$ ) in whom *BB* genotype frequency was very low. The minor allele frequency distributions differed significantly by ethnicity

**Table 1. Participant characteristics**

Characteristics	No. participants (%)	
	Cases (n = 313)	Controls (n = 574)
Age (y)		
<45	70 (22)	117 (21)
45-54	90 (29)	173 (30)
55-64	75 (24)	146 (25)
>64	78 (25)	138 (24)
Ethnicity		
Caucasian	72 (23)	148 (26)
Japanese	94 (30)	173 (30)
Filipino	36 (12)	79 (14)
Hawaiian	69 (22)	106 (18)
Other Asian	17 (5)	27 (5)
Mixed	25 (8)	41 (7)
Education (y)		
≤12	128 (41)	169 (29)
13-14	98 (31)	190 (33)
≥15	87 (28)	215 (38)
Family history of ovarian cancer		
Yes	16 (5)	9 (2)
No	297 (95)	565 (98)
Gravidity		
Nulligravid	74 (24)	56 (10)
1	42 (13)	67 (12)
2-3	121 (39)	246 (43)
≥4	76 (24)	205 (36)
Used oral contraceptives		
Yes	144 (46)	394 (69)
No	169 (54)	180 (31)
Had tubal ligation		
Yes	48 (15)	181 (32)
No	265 (85)	393 (68)
Menopausal status		
Premenopausal	111 (35)	215 (37)
Postmenopausal	202 (65)	359 (63)
Type of menopause		
Hysterectomy	32 (16)	47 (13)
Natural menopause	170 (84)	312 (87)
Use of menopausal hormones		
Never used	106 (52)	149 (42)
Estrogen only	32 (16)	50 (14)
Progesterone only	12 (6)	19 (5)
Combined estrogen and progesterone	52 (26)	141 (39)

for all polymorphisms under study (Table 2). Strong linkage disequilibrium ( $D' > 0.90$ ;  $r^2 > 0.90$ ) was observed between *BsmI*, *ApaI*, and *TaqI* among Japanese and Caucasian women and was weaker among other ethnic groups. *FokI* and *Cdx-2* SNPs were not in linkage disequilibrium with other SNPs.

Caucasian carriers of the *ApaI* A allele were at increased risk of ovarian cancer ( $P_{\text{trend}} = 0.02$ ; Table 3).

A nonsignificant increased risk associated with this allele was also observed among Japanese women. Although *BsmI* B and *TaqI* t allele carriers among Caucasian and Japanese women were at higher ovarian cancer risk, these associations were not statistically significant. Among Caucasians, carriers of the *FokI* f allele were at significantly increased risk of ovarian carcinoma compared with women who were homozygous for the common allele ( $P_{\text{trend}} = 0.04$ ). The *Cdx-2* A allele was associated with a decreased ovarian cancer risk in Japanese women: carriers of the GA genotype had a significantly reduced risk of ovarian cancer (OR 0.5, 95% CI 0.3-0.9;  $P_{\text{trend}} = 0.03$ ). When all three SNPs in the 3' region were studied simultaneously, five common *BsmI*-*ApaI*-*TaqI* genotypes were observed. Homozygous carriers of the *bbatT* genotype, including all common alleles of all three polymorphisms, represented a majority of our population (74%). Among Caucasian women, significantly higher risk was observed for carriers of the *bbatT* and *BBAAt* genotypes when compared with homozygous *bbatT* carriers.

## Discussion

In this multiethnic study, we explored the association of several common *VDR* gene polymorphisms with ovarian cancer risk. Frequency distributions of the minor alleles for all five SNPs varied significantly by ethnicity and were similar to the data in the National Center for Biotechnology Information data base and in the published literature (18, 32, 39, 40). The association of *VDR* polymorphisms with ovarian cancer risk was generally inconsistent among ethnic groups. In Caucasian women, the *ApaI* A allele was associated with an increased risk of ovarian carcinoma, as were two of the 3' area genotypes, *bbatT* and *BBAAt*. Although Japanese women who were carriers of the *ApaI* A allele and *bbatT* and *BBAAt* genotypes were at increased risk, the results were not statistically significant. The *FokI* f allele was associated with increased ovarian cancer risk in Caucasian women, but not in other ethnic groups. In contrast, the *Cdx-2* GA genotype was associated with a decreased risk of ovarian cancer only in Japanese women.

An association of the *FokI* f allele with increased risk of ovarian cancer was consistent with our hypothesis. The f variant was found to have lower transcriptional activation in laboratory studies (29, 30, 41, 42) and has been associated with an increased breast cancer risk in Caucasian women when studied in combination with the poly(A) polymorphism in the 3' region of *VDR* (43).

**Table 2. Frequency distributions of minor alleles among control subjects by ethnic group**

SNPs*	Minor allele*	Restriction site present	Caucasians (n = 148)	Japanese (n = 173)	Other Asian (n = 27)	Filipino (n = 79)	Hawaiians (n = 106)	Mixed (n = 41)	P, $\chi^2$ test
rs10735810 ( <i>FokI</i> )	T (f)	Yes	0.38	0.25	0.48	0.30	0.36	0.40	0.04
rs11568820 ( <i>Cdx-2</i> )	A		0.18	0.44	0.44	0.46	0.45	0.35	<0.0001
rs731236 ( <i>TaqI</i> )	C (t)	Yes	0.41	0.11	0.06	0.03	0.22	0.28	<0.0001
rs7975232 ( <i>ApaI</i> )	C (a)	Yes	0.50	0.68	0.80	0.78	0.49	0.65	<0.0001
rs1544410 ( <i>BsmI</i> )	A (B)	No	0.42	0.12	0.02	0.08	0.28	0.30	<0.0001

\*Minor allele in Caucasians. Nomenclature is based on the National Center of Biotechnology Information. In brackets, variant allele designations often used in publications.

**Table 3. VDR SNP and 3' area BsmI-ApaI-TaqI combined genotype associations with epithelial ovarian cancer risk**

Genotype	Caucasian			Japanese		
	Cases n (%)	Controls n (%)	OR (95% CI)*	Cases n (%)	Controls n (%)	OR (95% CI)†
rs7975232 ( <i>ApaI</i> )						
<i>aa</i>	11 (16)	33 (24)	1.0 reference	38 (41)	75 (45)	1.0 (reference)
<i>aA</i>	37 (53)	72 (52)	2.8 (1.2-7.0)	47 (50)	79 (47)	1.2 (0.7-2.1)
<i>AA</i>	22 (31)	34 (24)	3.4 (1.3-9.1)	8 (9)	14 (8)	1.2 (0.5-3.1)
<i>P</i> <sub>trend</sub>			0.02			0.56
rs1544410 ( <i>BsmI</i> )						
<i>bb</i>	26 (27)	48 (33)	1.0 (reference)	72 (77)	137 (80)	1.0 (reference)
<i>bB</i>	30 (42)	70 (49)	1.2 (0.6-2.4)	19 (20)	28 (16)	1.4 (0.7-2.8)
<i>BB</i>	15 (21)	26 (18)	1.3 (0.5-3.3)	3 (3)	6 (4)	0.9 (0.2-4.0)
<i>P</i> <sub>trend</sub>			0.53			0.49
rs731236 ( <i>TaqI</i> )						
<i>TT</i>	26 (36)	48 (33)	1.0 reference	73 (79)	138 (80)	1.0 (reference)
<i>Tt</i>	30 (42)	77 (53)	1.1 (0.5-2.1)	17 (18)	31 (18)	1.1 (0.6-2.2)
<i>tt</i>	16 (22)	21 (14)	1.8 (0.7-4.4)	3 (3)	4 (2)	1.7 (0.4-7.9)
<i>P</i> <sub>trend</sub>			0.28			0.53
rs10735810 ( <i>FokI</i> )						
<i>FF</i>	16 (23)	58 (40)	1.0 reference	37 (40)	74 (43)	1.0 (reference)
<i>Ff</i>	44 (62)	64 (44)	2.5 (1.3-4.8)	48 (52)	80 (47)	1.2 (0.7-2.0)
<i>ff</i>	11 (15)	22 (15)	2.1 (0.8-5.2)	8 (9)	18 (10)	0.9 (0.4-2.2)
<i>P</i> <sub>trend</sub>			0.04			0.87
rs11568820 ( <i>Cdx-2</i> )						
<i>GG</i>	44 (63)	95 (66)	1.0 reference	36 (39)	45 (26)	1.0 (reference)
<i>GA</i>	21 (30)	44 (30)	1.0 (0.6-1.8)	41 (45)	99 (58)	0.5 (0.3-0.9)
<i>AA</i>	5 (7)	6 (4)	1.5 (0.4-5.2)	15 (16)	27 (16)	0.9 (0.4-1.8)
<i>P</i> <sub>trend</sub>			0.96			0.03
<i>BsmI-ApaI-TaqI</i>						
<i>bb/aa/TT</i>	11 (16)	33 (24)	1.0 reference	75 (45)	38 (41)	1.0 reference
<i>bb/aA/TT</i>	13 (19)	12 (9)	4.2 (1.3-13.1)	31 (33)	55 (33)	1.1 (0.6-1.9)
<i>bB/aA/Tt</i>	24 (35)	54 (40)	2.3 (0.9-5.9)	15 (16)	19 (11)	1.8 (0.8-4.0)
<i>bB/AA/Tt</i>	5 (7)	12 (9)	2.4 (0.6-9.7)	2 (2)	8 (5)	0.5 (0.1-2.4)
<i>BB/AA/tt</i>	15 (22)	16 (12)	5.2 (1.6-17.5)	3 (3)	1 (2)	2.3 (0.4-12.3)

\*For 3' area SNPs (*ApaI*, *BsmI*, and *TaqI*) and combined genotypes, adjusted for education, gravidity, use of oral contraceptives, and menopausal hormones.  
†For 3' area SNPs and combined genotypes, adjusted for education and use of menopausal hormones.

*ApaI* and two other SNPs in the 3' region of the gene do not affect the structure of the VDR protein but have been shown to affect VDR mRNA stability or may be in linkage disequilibrium with some other functional SNP (33). The association of the *Cdx-2 A* allele with decreased ovarian cancer risk may result from more efficient *Cdx-2* binding and higher VDR activity associated with this variant (31).

Ethnic differences in the association of VDR polymorphisms with ovarian cancer risk might be partially explained by differences in variant allele frequencies. Frequencies of the *ApaI A* allele, *TaqI t*, and *BsmI B* alleles were very low in Japanese and other Asian women. In addition, the distribution of *BsmI* in Japanese women was not consistent with Hardy-Weinberg equilibrium, which might influence its association with risk. Substantial racial/ethnic variation is observed in the incidence of ovarian cancer with the highest rates in Caucasian women and the lowest rates among Asian women (44). Lower ovarian cancer incidence in Japanese women, who have significantly lower prevalence of the *ApaI A* allele and *BBAAtt ApaI-BsmI-TaqI*, is consistent with our findings. On the other hand, the *Cdx-2* variant allele, which was associated with decreased ovarian cancer risk in Japanese women only, was much more common in Asian women than in Caucasians. Increased bone mineral density in Japanese postmenopausal women (31, 32), but not in Caucasian

women (39), with the *Cdx-2 A* allele has been reported. At present, the reason for this difference is not known and might be related to environmental factors, such as calcium intake (45).

VDR and its ligand vitamin D regulate fundamental processes of cellular proliferation, differentiation, and apoptosis that have the potential to influence ovarian cancer development. Specific mechanisms for the anti-proliferative effect of VDR have yet to be determined. This effect in neoplastic cells may be related to ability of liganded VDR to interact with cell cycle regulatory proteins p21 and p27 leading to G<sub>1</sub> arrest, as well as to control cell growth factors, such as *c-myc* and *c-fos*, or to elicit apoptosis by down-regulating *Bcl-2* (46). VDR may also inhibit androgen receptor expression found in majority of ovarian tumors (12). A role of androgen in ovarian carcinogenesis has been suggested (47, 48), and Ahonen et al. (12) reported that vitamin D can antagonize the growth-promoting effects of dehydroandrostenedione in ovarian cancer cells.

VDR polymorphisms have been reported in association with breast, colon, and prostate cancer risk, suggesting a common link with ovarian cancer risk. VDR was found to be down-regulated in colon and breast tumors (49), but was found to be up-regulated in ovarian tumors (13, 49) when compared with non-matched normal ovarian tissue. The basis for the up-regulation of VDR in ovarian tumors is not clear,

although higher levels of androgen in association with ovarian cancer may affect the expression of VDR (49). Dihydrotestosterone has been shown to increase VDR expression in the ovary (12). Studies comparing normal and cancerous tissue from the same individual might help to confirm if VDR is selectively up-regulated in ovarian tumors.

Strengths of this study include its population basis, histologic confirmation of all case diagnoses, and stringent laboratory quality control procedures. An advantage of our study design was its multiethnic nature, which might help to better understand genetic contributions to ovarian cancer risk. Although the study had sufficient power to detect significant associations in Caucasian and Japanese women, the number of women in other ethnic groups was small. Furthermore, power was limited to examine combinations of genetic variants. Larger studies are needed to explore the effects of these genetic variations.

Ovarian cancer is the fatal gynecologic malignancy, mostly because of its late clinical manifestations and absence of screening methods for early detection. As the majority of women who develop this cancer do not carry highly penetrant *BRCA* gene mutations, low-penetrant genes could be useful in detecting women at higher risk of ovarian cancer, allowing for an individualized approach to screening. Our study provides some evidence that the *VDR* gene might influence ovarian cancer susceptibility.

## References

- Garland CF, Mohr SB, Gorham ED, Grant WB, Garland FC. Role of ultraviolet B irradiance and vitamin D in prevention of ovarian cancer. *Am J Prev Med* 2006;31:512-4.
- Lefkowitz ES, Garland CF. Sunlight, vitamin D, ovarian cancer mortality rates in US women. *Int J Epidemiol* 1994;23:1133-6.
- Salazar-Martinez E, Lazcano-Ponce EC, Gonzalez Lira-Lira G, Escudero-De Los Rios P, Hernandez-Avila M. Nutritional determinants of epithelial ovarian cancer risk: a case-control study in Mexico. *Oncology* 2002;63:151-7.
- Kushi LH, Mink PJ, Folsom AR, et al. Prospective study of diet and ovarian cancer. *Am J Epidemiol* 1999;149:21-31.
- Genkinger JM, Hunter DJ, Spiegelman D, et al. Dairy products and ovarian cancer: a pooled analysis of 12 cohort studies. *Cancer Epidemiol Biomarkers Prev* 2006;15:364-72.
- Saunders DE, Christensen C, Wappler NL, et al. Inhibition of *c-myc* in breast and ovarian carcinoma cells by 1,25-dihydroxyvitamin D<sub>3</sub>, retinoic acid and dexamethasone. *Anticancer Drugs* 1993;4:201-8.
- Jiang F, Bao J, Li P, Nicosia SV, Bai W. Induction of ovarian cancer cell apoptosis by 1,25-dihydroxyvitamin D<sub>3</sub> through the down-regulation of telomerase. *J Biol Chem* 2004;279:53213-21.
- Saunders DE, Christensen C, Lawrence WD, et al. Receptors for 1,25-dihydroxyvitamin D<sub>3</sub> in gynecologic neoplasms. *Gynecol Oncol* 1992;44:131-6.
- Zhang X, Jiang F, Li P, et al. Growth suppression of ovarian cancer xenografts in nude mice by vitamin D analogue EB1089. *Clin Cancer Res* 2005;11:323-8.
- Whitfield GK, Jurutka PW, Haussler CA, Haussler MR. Steroid hormone receptors: evolution, ligands, and molecular basis of biologic function. *J Cell Biochem* 1999;Suppl 32-33:110-22.
- Baker AR, McDonnell DP, Hughes M, et al. Cloning and expression of full-length cDNA encoding human vitamin D receptor. *Proc Natl Acad Sci U S A* 1988;85:3294-8.
- Ahonen MH, Zhuang YH, Aine R, Ylikomi T, Tuohimaa P. Androgen receptor and vitamin D receptor in human ovarian cancer: growth stimulation and inhibition by ligands. *Int J Cancer* 2000;86:40-6.
- Villena-Heinsen C, Meyberg R, Axt-Flidner R, Reitnauer K, Reichrath J, Friedrich M. Immunohistochemical analysis of 1,25-dihydroxyvitamin-D<sub>3</sub>-receptors, estrogen and progesterone receptors and Ki-67 in ovarian carcinoma. *Anticancer Res* 2002;22:2261-7.
- Holick MF. Vitamin D: its role in cancer prevention and treatment. *Prog Biophys Mol Biol* 2006;92:49-59.
- Kinuta K, Tanaka H, Moriwake T, Aya K, Kato S, Seino Y. Vitamin D is an important factor in estrogen biosynthesis of both female and male gonads. *Endocrinology* 2000;141:1317-24.
- Cramer DW, Welch WR. Determinants of ovarian cancer risk. II. Inferences regarding pathogenesis. *J Natl Cancer Inst* 1983;71:717-21.
- Cramer DW. Epidemiologic aspects of early menopause and ovarian cancer. *Ann N Y Acad Sci* 1990;592:363-75.
- Uitterlinden AG, Fang Y, van Meurs JB, Pols HA, van Leeuwen JP. Genetics and biology of vitamin D receptor polymorphisms. *Gene* 2004;338:143-56.
- Chen WY, Bertone-Johnson ER, Hunter DJ, Willett WC, Hankinson SE. Associations between polymorphisms in the vitamin D receptor and breast cancer risk. *Cancer Epidemiol Biomarkers Prev* 2005;14:2335-9.
- Knight JA, Lesosky M, Barnett H, Raboud JM, Vieth R. Vitamin D and reduced risk of breast cancer: a population-based case-control study. *Cancer Epidemiol Biomarkers Prev* 2007;16:422-9.
- Curran JE, Vaughan T, Lea RA, Weinstein SR, Morrison NA, Griffiths LR. Association of A vitamin D receptor polymorphism with sporadic breast cancer development. *Int J Cancer* 1999;83:723-6.
- Kim HS, Newcomb PA, Ulrich CM, et al. Vitamin D receptor polymorphism and the risk of colorectal adenomas: evidence of interaction with dietary vitamin D and calcium. *Cancer Epidemiol Biomarkers Prev* 2001;10:869-74.
- Park K, Woo M, Nam J, Kim JC. Start codon polymorphisms in the vitamin D receptor and colorectal cancer risk. *Cancer Lett* 2006;237:199-206.
- Bodiwala D, Luscombe CJ, French ME, et al. Polymorphisms in the vitamin D receptor gene, ultraviolet radiation, and susceptibility to prostate cancer. *Environ Mol Mutagen* 2004;43:121-7.
- Taylor JA, Hirvonen A, Watson M, Pittman G, Mohler JL, Bell DA. Association of prostate cancer with vitamin D receptor gene polymorphism. *Cancer Res* 1996;56:4108-10.
- Valdivielso JM, Fernandez E. Vitamin D receptor polymorphisms and diseases. *Clin Chim Acta* 2006;371:1-12.
- Tung KH, Goodman MT, Wu AH, et al. Aggregation of ovarian cancer with breast, ovarian, colorectal, and prostate cancer in first-degree relatives. *Am J Epidemiol* 2004;159:750-8.
- Gross C, Krishnan AV, Malloy PJ, Eccleshall TR, Zhao XY, Feldman D. The vitamin D receptor gene start codon polymorphism: a functional analysis of FokI variants. *J Bone Miner Res* 1998;13:1691-9.
- Jurutka PW, Remus LS, Whitfield GK, et al. The polymorphic N terminus in human vitamin D receptor isoforms influences transcriptional activity by modulating interaction with transcription factor IIB. *Mol Endocrinol* 2000;14:401-20.
- Whitfield GK, Remus LS, Jurutka PW, et al. Functionally relevant polymorphisms in the human nuclear vitamin D receptor gene. *Mol Cell Endocrinol* 2001;177:145-59.
- Yamamoto H, Miyamoto K, Li B, et al. The caudal-related homeodomain protein Cdx-2 regulates vitamin D receptor gene expression in the small intestine. *J Bone Miner Res* 1999;14:240-7.
- Arai H, Miyamoto KI, Yoshida M, et al. The polymorphism in the caudal-related homeodomain protein Cdx-2 binding element in the human vitamin D receptor gene. *J Bone Miner Res* 2001;16:1256-64.
- Jurutka PW, Whitfield GK, Hsieh JC, Thompson PD, Haussler CA, Haussler MR. Molecular nature of the vitamin D receptor and its role in regulation of gene expression. *Rev Endocr Metab Disord* 2001;2:203-16.
- Surveillance, Epidemiology, and End Results Program. Summary Staging Guide for the Cancer Surveillance, Epidemiology, and End Results Reporting (SEER) Program. Bethesda, MD: U S. Department of Health and Human Services, Public Health Services, National Institute of Health., 1986.
- Oyama N, Johnson D B. Hawaii Health Surveillance Program Survey Methods and Procedures. R & S Report No 54. Honolulu, Hawaii: State Department of Health Research and Statistics Office., 1986.
- Tung KH, Goodman MT, Wu AH, et al. Reproductive factors and epithelial ovarian cancer risk by histologic type: a multiethnic case-control study. *Am J Epidemiol* 2003;158:629-38.
- Barrett JC, Fry B, Maller J, Daly MJ. Haploview: analysis and visualization of LD and haplotype maps. *Bioinformatics* 2005;21:263-5.
- Brennan P, Hsu CC, Moullan N, et al. Effect of cruciferous vegetables on lung cancer in patients stratified by genetic status: a mendelian randomisation approach. *Lancet* 2005;366:1558-60.

39. Fang Y, van Meurs JB, Bergink AP, et al. Cdx-2 polymorphism in the promoter region of the human vitamin D receptor gene determines susceptibility to fracture in the elderly. *J Bone Miner Res* 2003;18:1632–41.
40. Ferrari S, Rizzoli R, Bonjour JP. Genetic aspects of osteoporosis. *Curr Opin Rheumatol* 1999;11:294–300.
41. Arai H, Miyamoto K, Taketani Y, et al. A vitamin D receptor gene polymorphism in the translation initiation codon: effect on protein activity and relation to bone mineral density in Japanese women. *J Bone Miner Res* 1997;12:915–21.
42. Colin EM, Weel AE, Uitterlinden AG, et al. Consequences of vitamin D receptor gene polymorphisms for growth inhibition of cultured human peripheral blood mononuclear cells by 1, 25-dihydroxyvitamin D<sub>3</sub>. *Clin Endocrinol Oxf* 2000;52:211–6.
43. Guy M, Lowe LC, Bretherton-Watt D, et al. Vitamin D receptor gene polymorphisms and breast cancer risk. *Clin Cancer Res* 2004;10:5472–81.
44. Parkin DM, Whelan SL, Ferlay J, Teppo L. *Cancer Incidence in Five Continents*. Lyon, France: IARC, 2002.
45. Morita A, Iki M, Dohi Y, et al. Effects of the Cdx-2 polymorphism of the vitamin D receptor gene and lifestyle factors on bone mineral density in a representative sample of Japanese women: the Japanese Population-based Osteoporosis (JPOS) Study. *Calcif Tissue Int* 2005;77:339–47.
46. Haussler MR, Whitfield GK, Haussler CA, et al. The nuclear vitamin D receptor: biological and molecular regulatory properties revealed. *J Bone Miner Res* 1998;13:325–49.
47. Adashi EY. The climacteric ovary as a functional gonadotropin-driven androgen-producing gland. *Fertil Steril* 1994;62:20–7.
48. Risch HA. Hormonal etiology of epithelial ovarian cancer, with a hypothesis concerning the role of androgens and progesterone. *J Natl Cancer Inst* 1998;90:1774–86.
49. Anderson MG, Nakane M, Ruan X, Kroeger PE, Wu-Wong JR. Expression of VDR and CYP24A1 mRNA in human tumors. *Cancer Chemother Pharmacol* 2006;57:234–40.

## *Vitamin D Receptor Gene Polymorphisms and Epithelial Ovarian Cancer Risk*

Galina Lurie, Lynne R. Wilkens, Pamela J. Thompson, et al.

*Cancer Epidemiol Biomarkers Prev* 2007;16:2566-2571.

**Updated version** Access the most recent version of this article at:  
<http://cebp.aacrjournals.org/content/16/12/2566>

**Cited articles** This article cites 46 articles, 9 of which you can access for free at:  
<http://cebp.aacrjournals.org/content/16/12/2566.full#ref-list-1>

**Citing articles** This article has been cited by 5 HighWire-hosted articles. Access the articles at:  
<http://cebp.aacrjournals.org/content/16/12/2566.full#related-urls>

**E-mail alerts** [Sign up to receive free email-alerts](#) related to this article or journal.

**Reprints and Subscriptions** To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at [pubs@aacr.org](mailto:pubs@aacr.org).

**Permissions** To request permission to re-use all or part of this article, use this link  
<http://cebp.aacrjournals.org/content/16/12/2566>.  
Click on "Request Permissions" which will take you to the Copyright Clearance Center's (CCC) Rightslink site.