

# Deletion Polymorphism of UDP-Glucuronosyltransferase 2B17 and Risk of Prostate Cancer in African American and Caucasian Men

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## Abstract

**Purpose:** UDP-glucuronosyltransferases (UGT) are a family of enzymes that glucuronidate many endogenous chemicals, including androgens. This makes them more hydrophilic, alters biological activity, and facilitates their excretion. A deletion polymorphism in the *UGT2B17* gene was recently described that was associated with a reduced rate of glucuronidation *in vivo*. The purpose of this study was to determine if the deletion polymorphism is associated with susceptibility to prostate cancer. **Materials and Methods:** *UGT2B17* expression was determined by reverse transcription-PCR of pathologically normal prostate tissues ( $n = 5$ ). In a case-control study with 420 patients with incident primary prostate cancer (127 African Americans and 293 Caucasians) and 487 controls (120 African Americans and 367 Caucasians), the frequency of *UGT2B17* deletion polymorphism in genomic DNA was compared between cases and controls with PCR analysis.

**Results:** *UGT2B17* mRNA was detected only in individuals with at least one *UGT2B17* allele. The frequency of the null genotype was present in 0.11 and 0.12 of Caucasian and African American controls, respectively. When all subjects were considered, a significant association was found between the *UGT2B17* deletion polymorphism and prostate cancer risk [odds ratio (OR), 1.7; 95% confidence interval (95% CI), 1.2-2.6]. There was an increase in prostate cancer risk among individuals with *UGT2B17* deletion polymorphism in Caucasians (OR, 1.9; 95% CI, 1.2-3.0) but not in African Americans (OR, 1.3; 95% CI, 0.6-2.7).

**Conclusions:** These results suggest that the *UGT2B17* enzyme may play a role in the metabolism of androgens in prostate tissue and that the *UGT2B17* deletion polymorphism is associated with prostate cancer risk. (Cancer Epidemiol Biomarkers Prev 2006;15(8):1473-8)

## Introduction

Prostate cancer is the fourth most common cancer in men, comprising approximately one eighth of all male-specific cancers in the world (1). Prostate cancer is much more common in developed than in developing countries. Developed countries account for nearly 75% of all new prostate cancer cases. The incidence and mortality rates of prostate cancer among African American men are significantly higher than that of any other ethnic group in the United States and the world (1-3). The etiology of prostate cancer remains unknown and few risk factors for prostate cancer, such as race, age, family history, and steroid hormone levels, have been suggested (4). Variations in androgen levels have been suggested as a risk factor for prostate cancer but results are not consistent (5-9). The differences in androgen levels may be due to variation in activity or expression of the enzymes responsible for metabolism of androgens. Most epidemiologic studies have focused on enzymes involved in production of androgens, such as CYP3A4 (10-13), CYP17 (14-22), CYP3A43

(23), and SRD5A2 (22, 24). Although differences in androgen levels may reflect variation in catabolism, fewer studies (25-28) have evaluated this hypothesis.

UDP-glucuronosyltransferases (UGT) are a family of enzymes that glucuronidate many endogenous chemicals (e.g., bilirubin and steroid hormones) and xenobiotics (29, 30). Two groups of UGTs (UGT1 and UGT2) have been identified based on sequence homology and substrate specificity (31). Generally, UGT2B enzymes metabolize steroid hormones and xenobiotics (32, 33). Both *UGT2B15* and *UGT2B17* have been implicated in the metabolism of androgens, including androsterone, testosterone, androstane-3 $\alpha$ ,17 $\beta$ -diol, and dihydrotestosterone, which have been postulated to modify risk of prostate cancer (refs. 34-37; Fig. 1). *UGT2B17* shares 95% and 94% identity with *UGT2B15* cDNA and its amino acid sequence (36). The high level of cDNA and amino acid sequence homology between *UGT2B15* and *UGT2B17* suggests that these genes may have arisen from a gene duplication event (38).

We and others previously reported that *UGT2B15* Asp85Tyr polymorphism is significantly associated with prostate cancer risk (25, 27, 28). In addition, it has been suggested that a deletion polymorphism of *UGT2B17* is a risk factor for high transplant-related mortality because the *UGT2B17* enzyme can be immunogenic in the recipient with homozygous *UGT2B17* deletion (39, 40). A recent study suggested that *UGT2B17* deletion polymorphism is significantly associated with reduced glucuronidation activities in human liver microsomes (41) and the bimodal distribution of the testosterone excretion (42).

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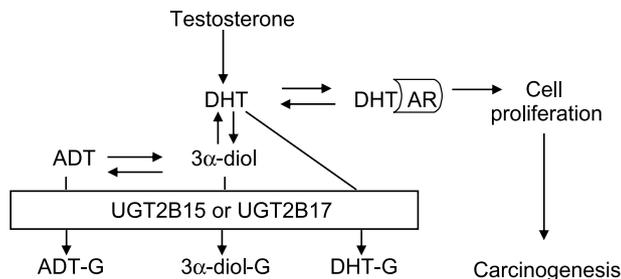
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**Figure 1.** Elimination of dihydrotestosterone in prostate cells (modified from ref. 67). Dihydrotestosterone is conjugated and inactivated either by the UGT2B15 or UGT2B17 enzyme and then excreted in urine. DHT, dihydrotestosterone; ADT, androsterone; 3 $\alpha$ -diol, androstane-3 $\alpha$ ,17 $\beta$ -diol; AR, androgen receptor.

Smoking is suggested as a contributing factor in prostate cancer mortality (43) and a putative risk factor for prostate cancer (44, 45). However, this association has been weak and inconsistent (46). One of many hypothesized biological mechanisms between smoking and prostate cancer is that smoking may be associated with higher levels of serum testosterone, which acts as a promoter of prostate cancer in men (47). Therefore, deletion of the UGT2B17 enzyme activity may be associated with prostate cancer risk because individuals with defective UGT2B17 activity may have a higher serum androgen level than subjects with normal activity. In the present study, we examined the potential association of the UGT2B17 deletion polymorphism with prostate cancer risk by assessing the expression pattern of the UGT2B17 enzyme in prostate tissues and by comparing UGT2B17 genotypes in prostate cancer patients and noncancer controls in African Americans and Caucasians.

## Materials and Methods

**Tissue Samples and UGT2B17 Expression Analysis.** Normal human prostate tissues were obtained from the Tissue Procurement Facility at the H. Lee Moffitt Cancer Center and Research Institute (Tampa, FL). All specimens came from subjects undergoing cancer surgery and were quick-frozen at  $-70^{\circ}\text{C}$  within 30 minutes of surgery. Total RNA was isolated from 0.5g samples of normal prostate tissues by the guanidinium isothiocyanate/cesium chloride method (48), followed by treatment with DNase I (Invitrogen, Carlsbad, CA).

Equimolar amounts of total RNA was reverse transcribed with the SuperScript reverse transcriptase amplification kit (Invitrogen) and 0.5 mg of oligo(dT)<sub>12-18</sub> primer as outlined in the protocol of the manufacturer. Sense (5'-gtgttgggaatattctgactataatata-3') and antisense (5'-caggtacataggaaggaggaa-3') primers homologous to UGT2B17 sequences were used in reverse transcription-PCR (RT-PCR); expected size, 242 bp).  $\beta$ -Actin sense (5'-tgatggtggcatgggtcag-3') and antisense (5'-gtgttggcgtacaggtctt-3') primers were added as an internal positive control in each RT-PCR assay (expected size, 756 bp). The primers were designed to amplify mRNA only from proposed genes by selecting primers that cross exon boundaries. Reactions without RNA were included as negative controls in all RT-PCR experiments. Ten- $\mu\text{l}$  aliquots were used from each PCR and resolved by electrophoresis in 8% polyacrylamide gels. PCR products were detected after staining the gels with 1  $\mu\text{g}/\text{mL}$  ethidium bromide and photography over UV light. The sequences of PCR products were confirmed by dideoxy sequencing (49) done at the Molecular Biology Core Facility.

**Study Populations and Sample Processing.** All cancer cases were histologically confirmed by the department of pathology

in each institute. A total of 307 incident cases (293 Caucasian and 14 African American) with primary adenocarcinoma of the prostate were recruited between 2002 and 2004 at the H. Lee Moffitt Cancer Center. Ninety-five percent of case subjects who were asked to participate in the study consented. Controls consisted of 377 subjects (367 Caucasians and 10 African Americans) who were visiting the Lifetime Cancer Screening Center, which is affiliated with the H. Lee Moffitt Cancer Center. At this center, routine screenings are offered to men for cancers of the prostate, colorectum, and skin. All control subjects were male and had no previous diagnosis of cancer. The control subjects were matched to cases on age at diagnosis ( $\pm 5$  years) and race. Eighty-three percent of the control subjects who were asked to participate in the study consented.

African Americans ( $n = 113$ ) with prostate cancer were recruited within 6 months of diagnosis between 1998 and 2003 at the University of Arkansas for Medical Sciences, University Hospital, the Central Arkansas Veterans Healthcare System (Little Rock, AR) and the Jefferson Regional Medical Center (Pine Bluff, AR). Controls ( $n = 110$ ), frequency matched to cases on age at diagnosis ( $\pm 5$  years) and race, were identified from Arkansas State Driver's License records, Centers for Medicare and Medicaid Services records, and a mass-mailing database that covers  $\sim 80\%$  of Arkansas residents. Exclusion criteria for the case-control study included a history of cancer. The recruitment rate for both cases and controls among African Americans was 55%. The appropriate Institutional Review Board approvals were obtained for the study protocol at each institute. Signed informed consent was obtained from all study subjects.

Nongenetic risk factor data for the current study were obtained through in-person interviews with cases and controls at the time of enrollment. The questionnaire covered (i) demographic information; (ii) family history of cancer; (iii) medical history; and (iv) detailed tobacco consumption. Smoking intensity (pack-years) was defined as the number of packs of cigarettes smoked per day multiplied by the number of years smoked. Study subjects who smoked 100 or fewer cigarettes in their lifetime were categorized as never smokers. Racial information was based on self-report and was broadly defined as Caucasian, African American, or other. Due to low numbers, individuals who reported other races were excluded ( $n = 8$ ). For cases, data on cancer stage, Gleason score, and prostate-specific antigen were abstracted from medical records.

Subjects were asked to provide a blood or buccal sample after the interview as a source of genomic DNA. DNA was extracted according to standardized protocols (50).

**Genotyping Assays.** The presence of the UGT2B17 deletion polymorphism was screened in all subjects using a multiplex-PCR assay. Because  $\beta$ -actin should be present in all cells, the amplification of this gene provides a positive control for each reaction. Therefore, samples in which only the  $\beta$ -actin gene is amplified were considered a deletion for UGT2B17. The multiplex PCR was done to amplify the part of UGT2B17 gene using UGT2B17-specific primers (sense, ggagttgtgaaagggtgct; antisense, cacagacgttatattatagtcag) with human  $\beta$ -actin primers (sense, ggccggcaccacatgtaccct; antisense, aggggccggactcgtact) as an internal positive control with  $60^{\circ}\text{C}$  annealing temperature. Negative controls were included in each PCR experiment. Products of the PCR reaction (expected size: UGT2B17, 350 bp;  $\beta$ -actin, 207 bp) were detected by electrophoresis on 8% PAGE gels and visualized using ethidium bromide stain and UV transillumination.

To ensure quality control of genotyping results, randomly selected PCR-amplified DNA samples ( $n = 3$ ) from subjects possessing UGT2B17 [+] genotype were examined by dideoxy DNA sequencing (49).

**Statistical Analysis.** Number (%) of subjects and mean values (SDs) were generated for all variables by case-control status.  $\chi^2$  test for categorical variables and two sample *t* test (or Wilcoxon rank-sum test) for continuous variables were used to test the statistical significance of any observed differences in demographic and clinical characteristics between cases and controls.

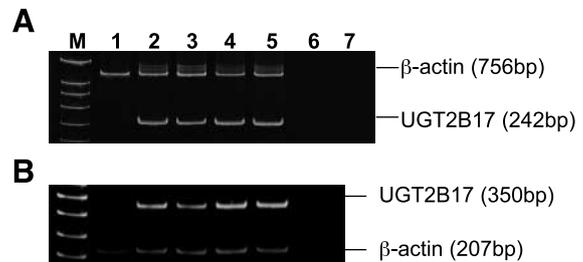
The risk for prostate cancer in relation to *UGT2B17* genotypes (positive versus null) was estimated with logistic regression modeling approaches. Potential confounding and gene-environment interaction effects were tested in the logistic regression model. Logistic regression model was also fitted separately among light smokers ( $\leq 26.5$  pack-years) and heavy smokers ( $> 26.5$  pack-years) for each race, with the cut point being the median among smoking controls. Final multivariable models for the *UGT2B17* were constructed using an empirical stepwise approach and they included the covariates of age (for both races). Odds ratios (OR) and their 95% confidence intervals (95% CI) were reported.

$P \leq 0.05$  (two sided) was considered to be statistically significant. The SAS software (v. 9.2, SAS, Cary, NC) was used for all analyses.

## Results

Table 1 provides descriptive characteristics of the cases and controls. Despite the frequency matching on age, cases tended to be older than controls ( $P < 0.0001$ ). Cases tended to smoke less than controls, but no differences were apparent among the African Americans. More men with prostate cancer (22%) than without (9%) reported having a first-degree family member with prostate cancer ( $P < 0.0001$ ). As expected, prostate-specific antigen levels of most patients are  $> 4$  ng/ml and Gleason scores range between 5 and 9.

To evaluate the potential physiologic importance of *UGT2B17* in prostate tissue and the association between expression and genotype, we did RT-PCR and genotyping analysis to determine whether the *UGT2B17* enzyme is expressed in normal human prostate tissues from individuals with different *UGT2B17* genotypes. As shown in Fig. 2A, RT-PCR amplification of a fragment of the expected size (242 bp) corresponding to that expected for *UGT2B17* mRNA was detected only in individuals with *UGT2B17* [+] genotype (Fig. 2B). These PCR products were confirmed by direct sequencing analysis (results not shown).



**Figure 2.** Expression and genotyping analysis of *UGT2B17* in normal prostate tissues. To investigate an association between *UGT2B17* genotypes and their expression, RNA and DNA samples extracted from same five individuals were used. **A.** *UGT2B17* mRNA was detected by RT-PCR of individual RNA samples. *M*, DNA marker; lanes 1 to 5, total prostate RNA from five different individuals. Only individuals with *UGT2B17* [+] genotype (subjects 2, 3, 4, and 5) show expression of *UGT2B17* mRNA. Upper bands indicate products from  $\beta$ -actin (756 bp) and lower bands are PCR products (242 bp) from *UGT2B17* mRNA. Lanes 6 and 7, negative controls for RT reaction and PCR, respectively. The primers were designed to amplify mRNA only from proposed genes by selecting primers that cross exon boundaries. **B.** The amplification of  $\beta$ -actin provides a positive internal control (expected size, 207 bp) for each PCR. A PCR amplification positive for  $\beta$ -actin but negative for *UGT2B17* (expected size: 350 bp) would indicate a null *UGT2B17* genotype, as shown in lane 1 (subject 1).

We obtained informative genotyping results of the *UGT2B17* null polymorphism from 97.9% of study population. Before beginning the analysis of the association of the polymorphisms with risk of prostate cancer, we first examined whether there were racial differences in genotype frequency. Racial differences in genotype frequency were not observed among the control groups ( $P = 0.46$ ). The frequencies of the *UGT2B17* deletion genotype among the control subjects were 11% and 12% in Caucasians and African Americans, respectively. The frequency of this polymorphism among African Americans in this study is significantly different from the one reported in a previous study (51).

To determine whether this genetic variant was associated with increased risk for prostate cancer, we compared genotypes in prostate cancer cases and controls (Table 2). When all subjects were considered, a significantly increased risk for

**Table 1. Selected characteristics of study subjects and comparison between cases and controls**

Variables	Cases ( <i>n</i> = 420)	Controls ( <i>n</i> = 487)	<i>P</i>
Mean age (range), y	64 (42-85)	60 (37-82)	<0.0001
Ethnicity (%)			
Caucasians	293 (70)	367 (75)	>0.05
African Americans	127 (30)	120 (25)	
Smoking			
Mean pack-years (SD) in Caucasians	20.0 (24.1)	29.2 (20.1)	<0.0001
Mean pack-years (SD) in African Americans	18.1 (25.9)	16.2 (29.0)	0.58
Family history of prostate cancer (%)*	51 of 233 (22)	33 of 353 (9)	<0.0001
Prostate-specific antigen (%)			
<4 ng/ml	45 (16)		
4-10 ng/ml	199 (69)		
>10 ng/ml	43 (15)		
Gleason score (%)			
5-6	165 (71)		
7-9	69 (29)		
Stage at diagnosis (%)			
I	171 (70)		
II	64 (26)		
III	8 (3)		
IV	1 (0.4)		

\*Men having a first-degree family member with prostate cancer.

**Table 2. Prostate cancer risk for the UGT2B17 genotypes alone and by race and smoking level**

UGT2B17 genotypes	Cases (%)	Controls (%)	Crude OR (95% CI)	Adjusted OR (95% CI)	P
All					
UGT2B17 [+]*	331 (82)	426 (88)	1.0 (reference)	1.0 (reference)	0.004
UGT2B17 null	75 (18)	56 (12)	1.7 (1.2-2.5)	1.7 (1.2-2.6) <sup>†</sup>	
Caucasians					
UGT2B17 [+]	237 (81)	325 (89)	1.0 (reference)	1.0 (reference)	0.006
UGT2B17 null	56 (19)	42 (11)	1.8 (1.2-2.8)	1.9 (1.2-3.0) <sup>‡</sup>	
African Americans					
UGT2B17 [+]	94 (83)	101 (88)	1.0 (reference)	1.0 (reference)	0.32
UGT2B17 null	19 (17)	14 (12)	1.5 (0.7-3.1)	1.3 (0.6-2.7) <sup>‡</sup>	
Light smokers ( $\leq 26.5$ pack-years)					
UGT2B17 [+]	99 (78)	171 (89)	1.0 (reference)	1.0 (reference)	0.011
UGT2B17 null	28 (22)	22 (11)	2.2 (1.2-4.0)	2.4 (1.2-4.8) <sup>†</sup>	
Heavy smokers (>26.5 pack-years)					
UGT2B17 [+]	99 (82)	168 (86)	1.0 (reference)	1.0 (reference)	0.382
UGT2B17 null	21 (18)	27 (14)	1.3 (0.7-2.5)	1.2 (0.6-2.3) <sup>†</sup>	

\*UGT2B17 [+]=UGT2B17 (+/+) homozygous wild and (+/0) heterozygous genotypes.

<sup>†</sup>Adjusted for race, age, and pack-years.

<sup>‡</sup>Adjusted for age and pack-years.

prostate cancer was observed for subjects with at least one UGT2B17 allele. The crude OR was 1.7 (95% CI, 1.2-2.5). The association remained (OR, 1.7; 95% CI, 1.2-2.6) after additional adjustment for age, race, and pack-years of smoking. Stratified analysis by race revealed an elevated prostate cancer risk among Caucasian individuals with UGT2B17 deletion polymorphism (OR, 1.9; 95% CI, 1.2-3.0). However, the main effect of the UGT2B17 deletion polymorphism on prostate cancer risk among African Americans was not statistically significant (OR, 1.3; 95% CI, 0.6-2.7). When tests for interaction were done, interaction between UGT2B17 polymorphism and race was not significant ( $P = 0.61$ ).

To examine whether the association between genotypes and prostate cancer risk varied by smoking history, we did a stratified analysis on genotype and smoking level (pack-years). There was a statistically significant association between UGT2B17 null genotype and prostate cancer risk among light smokers ( $\leq 26.5$  pack-years; OR, 2.4; 95% CI, 1.2-4.8) but not among heavy smokers (>26.5 pack-years; OR, 1.2; 95% CI, 0.6-2.3). Interaction between UGT2B17 polymorphism and smoking was not significant ( $P = 0.53$ ).

## Discussion

In the present study, we showed that the UGT2B17 enzyme is expressed in normal prostate tissues of individuals with UGT2B17 [+] genotype, consistent with a previous study (34). These data are consistent with the hypothesis that the UGT2B17 enzyme may play a role in degradation of dihydrotestosterone (52) and that an excessive amount of dihydrotestosterone may be associated with carcinogenesis in the prostate tissue (53). A significant association was found between the UGT2B17 deletion polymorphism and prostate cancer risk in Caucasians. However, we did not observe a similar association in African Americans. Potential explanations can be either biological differences between races in the androgen metabolism pathway or a small sample size. One hypothesis to explain the variation in prostate cancer incidence between racial groups is reference range differences among men of different races in androgens (54). For example, adult African American men have higher mean circulating concentrations of testosterone or other androgens than similarly aged white men (6). Therefore, against this background of high androgen levels in African Americans, the UGT2B17 deletion polymorphism, which is responsible for degradation of androgens, may have a diminished effect.

We previously reported a significant association between UGT2B15 polymorphism and prostate cancer risk (28). To the

best of our knowledge, no previous epidemiologic studies for UGT2B17 deletion polymorphism have examined an association with human cancer.

The clinical syndromes resulting from mutations or deletion of UGT2B members have not been described, although mutations of UGT1A family members have been well investigated and are responsible for the Crigler-Najjar and Gilbert syndromes (55-57). The deletion of UGT2B17 may be not clinically significant because results from various tests, such as normal physical exam, complete blood count, and serum chemistry, are normal. Notably, a higher prevalence (67-85%) of UGT2B17 deletion polymorphism has been reported in Asian individuals (40, 42) but the incidence rate of prostate cancer among Asian men is much lower than rates among Caucasian and African American men (58). These data suggest that other steroid-metabolizing enzymes may also have a role in prostate cancer risk.

The other UGT2Bs, especially UGT2B15, can glucuronidate similar androgens and are expressed in the prostate gland (34-36). However, *in vitro* studies suggest that UGT2B17 is more labile than UGT2B15, suggesting that regulation of UGT2B17 expression would lead to a more rapid change in the level of glucuronidated androgens (59). Furthermore, a recent study reported a different expression pattern between UGT2B17 and UGT2B15 in prostate tissues. UGT2B17 is expressed in basal cells where dehydroepiandrosterone is converted into androstane-3 $\alpha$ ,17 $\beta$ -diol and androsterone. However, expression of UGT2B15 was observed only in luminal cells, where dihydrotestosterone is formed from testosterone. These results suggest that UGT2B17 and UGT2B15 may have complementary roles and are expressed in cells where their specific substrates are synthesized (60). Therefore, comprehensive analyses of the UGT2B family and risk for prostate cancer are warranted. The proposed mechanism of carcinogenesis in prostate tissue involves the androgen receptor in the presence of dihydrotestosterone interacting with DNA in the proliferating prostate epithelial cells to produce permanent genomic mutations (Fig. 1).

Smoking has a positive correlation with level of serum testosterone, which acts as a promoter of prostate cancer and is degraded by the UGT2B17 enzyme (47). Dai et al. (61) found significant correlations between smoking level and serum androstenedione as well as testosterone in men. Therefore, excessive amount of dihydrotestosterone due to smoking and/or defective catabolism of androgens may increase prostate cancer risk. Another potential biological mechanism between smoking and prostate cancer is that UGT2B17 deletion polymorphism may enhance the effect of 4-(methyl-nitrosamino)-1-(3-pyridyl)-1-butanol exposure from smoking

on the risk of prostate cancer because the UGT2B17 enzyme not only metabolizes androgens but also detoxifies 4-(methyl-nitrosamino)-1-(3-pyridyl)-1-butanol, which is a lung carcinogen in rodents (41, 62, 63).

We observed a stronger association between UGT2B17 null polymorphism and prostate cancer among light smokers. This pattern is not unusual in previous studies examining polymorphism genotypes and cancer risk (64). It has been suggested that genetic variations in the ability to respond to tobacco carcinogens are most important in determining cancer risk at low levels of exposure and may be less relevant at higher smoking doses where high levels of carcinogen exposure overwhelm polymorphism-induced differences in enzyme activity and/or expression (65).

The prevalence of the UGT2B17 null polymorphism in Caucasians was similar to those reported from previous studies (39, 51) but significantly lower than that among Asian men (40). We also observed a higher prevalence in African Americans than previously reported (51). Although the reason for the different prevalence was not clear, these data suggest that variability in UGT2B17 genotype prevalence exists in African Americans from different regions of the United States, as we previously observed in GSTM1 null polymorphism (66).

Results from this study must be interpreted in light of limitations. We are aware of potential limitations of this study. Due to the nature of the genotyping analysis, we cannot distinguish between heterozygous (0/+) and homozygous (+/+) UGT2B17 genotypes. However, a recent study reported that a significant difference was observed when phenotypes from subjects who have at least one UGT2B17 allele were compared with those from UGT2B17 homozygous deletion subjects (41, 42). Second, although we had adequate power to detect main effects, the sample size is not large enough to adequately detect small interactions, especially among African Americans. Third, we observed significant age differences between cases and controls despite matching on age ( $\pm 5$  years). To address this, we adjusted for age when ORs were estimated. Finally, our control groups were not screened for prostate cancer by either prostate-specific antigen levels or digital rectal examination. Therefore, because of their ages, we estimate that some of them may have benign prostatic hyperplasia or even early stage of prostate cancer. This would result in misclassification and would bias the ORs to the null.

In conclusion, this study suggests that the UGT2B17 null polymorphism may play a role in prostate cancer risk. It will be important to replicate these observations in additional studies involving larger numbers of subjects.

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## References

- Potter JD. Food, nutrition and the prevention of cancer: a global perspective. American Institute for Cancer Research; 1997.
- Crawford ED. Epidemiology of prostate cancer. Urology 2003;62:3–12.
- McDavid K, Lee J, Fulton JP, Tonita J, Thompson TD. Prostate cancer incidence and mortality rates and trends in the United States and Canada. Public Health Rep 2004;119:174–86.
- Gronberg H. Prostate cancer epidemiology. Lancet 2003;361:859–64.
- Hsing AW, Reichardt JK, Stanczyk FZ. Hormones and prostate cancer: current perspectives and future directions. Prostate 2002;52:213–35.
- Ross R, Bernstein L, Judd H, Hanisch R, Pike M, Henderson B. Serum testosterone levels in healthy young black and white men. J Natl Cancer Inst 1986;76:45–8.
- Eaton NE, Reeves GK, Appleby PN, Key TJ. Endogenous sex hormones and prostate cancer: a quantitative review of prospective studies. Br J Cancer 1999;80:930–4.
- Mohr BA, Feldman HA, Kalish LA, Longcope C, McKinlay JB. Are serum hormones associated with the risk of prostate cancer? Prospective results from the Massachusetts Male Aging Study. Urology 2001;57:930–5.
- Heikkila R, Aho K, Heliövaara M, et al. Serum testosterone and sex hormone-binding globulin concentrations and the risk of prostate carcinoma: a longitudinal study. Cancer 1999;86:312–5.
- Zeigler-Johnson C, Friebe T, Walker AH, et al. CYP3A4, CYP3A5, and CYP3A43 genotypes and haplotypes in the etiology and severity of prostate cancer. Cancer Res 2004;64:8461–7.
- Keshava C, McCanlies EC, Weston A. CYP3A4 polymorphisms-potential risk factors for breast and prostate cancer: a HuGE review. Am J Epidemiol 2004;160:825–41.
- Spurdle AB, Goodwin B, Hodgson E, et al. The CYP3A4\*1B polymorphism has no functional significance and is not associated with risk of breast or ovarian cancer. Pharmacogenetics 2002;12:355–66.
- Westlind A, Lofberg L, Tindberg N, Andersson TB, Ingelman-Sundberg M. Interindividual differences in hepatic expression of CYP3A4: relationship to genetic polymorphism in the 5'-upstream regulatory region. Biochem Biophys Res Commun 1999;259:201–5.
- Kakinuma H, Tsuchiya N, Habuchi T, et al. Serum sex steroid hormone levels and polymorphisms of CYP17 and SRD5A2: implication for prostate cancer risk. Prostate Cancer Prostatic Dis 2004;7:333–7.
- Madigan MP, Gao YT, Deng J, et al. CYP17 polymorphisms in relation to risks of prostate cancer and benign prostatic hyperplasia: a population-based study in China. Int J Cancer 2003;107:271–5.
- Stanford JL, Noonan EA, Iwasaki L, et al. A polymorphism in the CYP17 gene and risk of prostate cancer. Cancer Epidemiol Biomarkers Prev 2002;11:243–7.
- Haiman CA, Stampfer MJ, Giovannucci E, et al. The relationship between a polymorphism in CYP17 with plasma hormone levels and prostate cancer. Cancer Epidemiol Biomarkers Prev 2001;10:743–8.
- Gsur A, Bernhofer G, Hinteregger S, et al. A polymorphism in the CYP17 gene is associated with prostate cancer risk. Int J Cancer 2000;87:434–7.
- Wadelius M, Andersson AO, Johansson JE, Wadelius C, Rane E. Prostate cancer associated with CYP17 genotype. Pharmacogenetics 1999;9:635–9.
- Lunn RM, Bell DA, Mohler JL, Taylor JA. Prostate cancer risk and polymorphism in 17 hydroxylase (CYP17) and steroid reductase (SRD5A2). Carcinogenesis 1999;20:1727–31.
- Ntais C, Polycarpou A, Ioannidis JP. Association of the CYP17 gene polymorphism with the risk of prostate cancer: a meta-analysis. Cancer Epidemiol Biomarkers Prev 2003;12:120–6.
- Forrest MS, Edwards SM, Houlston R, et al. Association between hormonal genetic polymorphisms and early-onset prostate cancer. Prostate Cancer Prostatic Dis 2005;8:95–102.
- Stone A, Ratnasingh LD, Emerson GL, et al. CYP3A43 Pro(340)Ala polymorphism and prostate cancer risk in African Americans and Caucasians. Cancer Epidemiol Biomarkers Prev 2005;14:1257–61.
- Zeigler-Johnson CM, Walker AH, Mancke B, et al. Ethnic differences in the frequency of prostate cancer susceptibility alleles at SRD5A2 and CYP3A4. Hum Hered 2002;54:13–21.
- MacLeod SL, Nowell S, Plaxco J, Lang NP. An allele-specific polymerase chain reaction method for the determination of the D85Y polymorphism in the human UDP-glucuronosyltransferase 2B15 gene in a case-control study of prostate cancer. Ann Surg Oncol 2000;7:777–82.
- Hajdinjak T, Zagradisnik B. Prostate cancer and polymorphism D85Y in gene for dihydrotestosterone degrading enzyme UGT2B15: Frequency of DD homozygotes increases with Gleason Score. Prostate 2004;59:436–9.
- Gsur A, Preyer M, Haidinger G, et al. polymorphism in the UDP-Glucuronosyltransferase 2B15 gene (D85Y) is not associated with prostate cancer risk. Cancer Epidemiol Biomarkers Prev 2002;11:497–8.
- Park J, Chen L, Shade K, et al. Asp85Tyr polymorphism in the udp-glucuronosyltransferase (UGT) 2B15 gene and the risk of prostate cancer. J Urol 2004;171:2484–8.
- Tephly TR, Burchell B. UDP-glucuronosyltransferases: a family of detoxifying enzymes. Trends Pharmacol Sci 1990;11:276–9.
- Gueraud F, Paris A. Glucuronidation: a dual control. Gen Pharmacol 1998;31:683–8.
- Mackenzie PI, Owens IS, Burchell B, et al. The UDP glycosyltransferase gene superfamily: recommended nomenclature update based on evolutionary divergence. Pharmacogenetics 1997;7:255–69.
- Tukey RH, Strassburg CP. Human UDP-glucuronosyltransferases: metabolism, expression, and disease. Annu Rev Pharmacol Toxicol 2000;40:581–616.
- Turgeon D, Carrier JS, Levesque E, Hum DW, Belanger A. Relative enzymatic activity, protein stability, and tissue distribution of human steroid-metabolizing UGT2B subfamily members. Endocrinology 2001;142:778–87.
- Belanger G, Beaulieu M, Marcotte B, et al. Expression of transcripts encoding steroid UDP-glucuronosyltransferases in human prostate hyperplastic tissue and the LNCaP cell line. Mol Cell Endocrinol 1995;113:165–73.
- Chen F, Ritter JK, Wang MG, McBride OW, Lubet RA, Owens IS. Characterization of a cloned human dihydrotestosterone/androstenediol UDP-glucuronosyltransferase and its comparison to other steroid isoforms. Biochemistry 1993;32:10648–57.
- Beaulieu M, Levesque E, Hum DW, Belanger A. Isolation and characterization of a novel cDNA encoding a human UDP-glucuronosyltransferase active on C19 steroids. J Biol Chem 1996;271:22855–62.
- Green MD, Oturu EM, Tephly TR. Stable expression of a human liver UDP-glucuronosyltransferase (UGT2B15) with activity toward steroid and xenobiotic substrates. Drug Metab Dispos 1994;22:799–805.

38. Turgeon D, Carrier JS, Levesque E, Beatty BG, Belanger A, Hum DW. Isolation and characterization of the human UGT2B15 gene, localized within a cluster of UGT2B genes and pseudogenes on chromosome 4. *J Mol Biol* 2000;295:489–504.
39. Murata M, Warren EH, Riddell SR. A human minor histocompatibility antigen resulting from differential expression due to a gene deletion. *J Exp Med* 2003;197:1279–89.
40. Terakura S, Murata M, Nishida T, et al. A UGT2B17-positive donor is a risk factor for higher transplant-related mortality and lower survival after bone marrow transplantation. *Br J Haematol* 2005;129:221–8.
41. Lazarus P, Zheng Y, Aaron Runkle E, Muscat JE, Wiener D. Genotype-phenotype correlation between the polymorphic UGT2B17 gene deletion and NNAL glucuronidation activities in human liver microsomes. *Pharmacogenet Genomics* 2005;15:769–78.
42. Jakobsson J, Ekstrom L, Inotsume N, et al. Large differences in testosterone excretion in Korean and Swedish men is strongly associated with an UDP-glucuronosyl transferase 2B17 polymorphism. *J Clin Endocrinol Metab* 2005;91:687–93.
43. Hsing AW, McLaughlin JK, Schuman LM, et al. Diet, tobacco use, and fatal prostate cancer: results from the Lutheran Brotherhood Cohort Study. *Cancer Res* 1990;50:6836–40.
44. Plaskon LA, Penson DF, Vaughan TL, Stanford JL. Cigarette smoking and risk of prostate cancer in middle-aged men. *Cancer Epidemiol Biomarkers Prev* 2003;12:604–9.
45. Giovannucci E, Rimm EB, Ascherio A, et al. Smoking and risk of total and fatal prostate cancer in United States health professionals. *Cancer Epidemiol Biomarkers Prev* 1999;8:277–82.
46. Hickey K, Do KA, Green A. Smoking and prostate cancer. *Epidemiol Rev* 2001;23:115–25.
47. Ferrini RL, Barrett-Connor E. Sex hormones and age: a cross-sectional study of testosterone and estradiol and their bioavailable fractions in community-dwelling men. *Am J Epidemiol* 1998;147:750–4.
48. Chomczynski P, Sacchi N. Single-step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction. *Anal Biochem* 1987;162:156–9.
49. Sanger F, Nicklen S, Coulson AR. DNA sequencing with chain-terminating inhibitors. *Proc Natl Acad Sci U S A* 1977;74:5463–7.
50. Park JY, Muscat JE, Ren Q, et al. CYP1A1 and GSTM1 polymorphisms and oral cancer risk. *Cancer Epidemiol Biomarkers Prev* 1997;6:791–7.
51. Wilson W III, Pardo-Manuel de Villena F, Lyn-Cook BD, et al. Characterization of a common deletion polymorphism of the UGT2B17 gene linked to UGT2B15. *Genomics* 2004;84:707–14.
52. Green MD, Tephly TR. Glucuronidation of amines and hydroxylated xenobiotics and endobiotics catalyzed by expressed human UGT1.4 protein. *Drug Metab Dispos* 1996;24:356–63.
53. Gann PH, Hennekens CH, Ma J, Longcope C, Stampfer MJ. Prospective study of sex hormone levels and risk of prostate cancer. *J Natl Cancer Inst* 1996;88:1118–26.
54. Platz EA, Giovannucci E. The epidemiology of sex steroid hormones and their signaling and metabolic pathways in the etiology of prostate cancer. *J Steroid Biochem Mol Biol* 2004;92:237–53.
55. Ritter JK, Yeatman MT, Ferreira P, Owens IS. Identification of a genetic alteration in the code for bilirubin UDP-glucuronosyltransferase in the UGT1 gene complex of a Crigler-Najjar type I patient. *J Clin Invest* 1992;90:150–5.
56. Moghrabi N, Clarke DJ, Burchell B, Boxer M. Cosegregation of intragenic markers with a novel mutation that causes Crigler-Najjar syndrome type I: implication in carrier detection and prenatal diagnosis. *Am J Hum Genet* 1993;53:722–9.
57. Bosma PJ, Chowdhury JR, Bakker C, et al. The genetic basis of the reduced expression of bilirubin UDP-glucuronosyltransferase 1 in Gilbert's syndrome. *N Engl J Med* 1995;333:1171–5.
58. Haqq C, Li R, Khodabakhsh D, et al. Ethnic and racial differences in prostate stromal estrogen receptor  $\alpha$ . *Prostate* 2005;65:101–9.
59. Guillemette C, Levesque E, Beaulieu M, Turgeon D, Hum DW, Belanger A. Differential regulation of two uridine diphospho-glucuronosyltransferases, UGT2B15 and UGT2B17, in human prostate LNCaP cells. *Endocrinology* 1997;138:2998–3005.
60. Chouinard S, Pelletier G, Belanger A, Barbier O. Cellular specific expression of the androgen-conjugating enzymes UGT2B15 and UGT2B17 in the human prostate epithelium. *Endocr Res* 2004;30:717–25.
61. Dai WS, Gutai JP, Kuller LH, Cauley JA. Cigarette smoking and serum sex hormones in men. *Am J Epidemiol* 1988;128:796–805.
62. Rivenson A, Hoffmann D, Prokopczyk B, Amin S, Hecht SS. Induction of lung and exocrine pancreas tumors in F344 rats by tobacco-specific and Areca-derived N-nitrosamines. *Cancer Res* 1988;48:6912–7.
63. Hecht SS. Biochemistry, biology, and carcinogenicity of tobacco-specific N-nitrosamines. *Chem Res Toxicol* 1998;11:559–603.
64. Park JY, Schantz SP, Stern JC, Kaur T, Lazarus P. Association between glutathione S-transferase  $\pi$  genetic polymorphisms and oral cancer risk. *Pharmacogenetics* 1999;9:497–504.
65. London SJ, Daly AK, Cooper J, Navidi WC, Carpenter CL, Idle JR. Polymorphism of glutathione S-transferase M1 and lung cancer risk among African-Americans and Caucasians in Los Angeles County, California. *J Natl Cancer Inst* 1995;87:1246–53.
66. Park LY, Muscat JE, Kaur T, et al. Comparison of GSTM polymorphisms and risk for oral cancer between African-Americans and Caucasians. *Pharmacogenetics* 2000;10:123–31.
67. Belanger A, Pelletier G, Labrie F, Barbier O, Chouinard S. Inactivation of androgens by UDP-glucuronosyltransferase enzymes in humans. *Trends Endocrinol Metab* 2003;14:473–9.

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