

Inflammation and Endometrial Cancer: A Hypothesis

Francesmary Modugno,^{1,2} Roberta B. Ness,^{1,2} Chu Chen,^{3,4,5} and Noel S. Weiss^{3,5}¹Department of Epidemiology, Graduate School of Public Health, University of Pittsburgh; ²University of Pittsburgh Cancer Institute, Pittsburgh, Pennsylvania; ³Program in Epidemiology, Public Health Sciences Division, Fred Hutchinson Cancer Research Center; ⁴Department of Otolaryngology: Head and Neck Surgery; and ⁵Department of Epidemiology, School of Public Health and Community Medicine, University of Washington, Seattle, Washington

Abstract

Endometrial cancer is the most common gynecologic malignancy in the United States. Substantial epidemiologic data implicate an imbalance of estrogens and progestogens in the etiology of this disease. We propose that inflammation also plays a role in endometrial cancer development. Emerging laboratory data suggest that elevated levels of prostaglandin E₂ may underlie the transformation of normal endometrium to neoplastic tissue and that *in vitro* nonsteroidal anti-inflammatory drugs may inhibit endometrial cancer cell growth. In this review, we suggest that the risk factors for endometrial cancer—unopposed estrogens, anovulation, polycystic ovary syndrome, excessive menstruation,

early menarche, and late menopause—may be viewed as factors increasing the exposure of the endometrium to inflammation, whereas pregnancy and smoking, two likely protective factors, have the opposite effect. Chronic inflammation can induce rapid cell division, increasing the possibility for replication error, ineffective DNA repair, and subsequent mutations. A proinflammatory milieu can also directly increase estrogen production. Hence, inflammation may work in conjunction with or in addition to estrogen exposure in the development of endometrial cancer. (Cancer Epidemiol Biomarkers Prev 2005;14(12):2840–7)

Introduction

Substantial epidemiologic data implicate an imbalance of estrogens and progestogens in the etiology of endometrial cancer (Table 1): early menarche (1, 2), late menopause (3, 4), anovulation (5, 6), prolonged menstruation, obesity (7–9), polycystic ovary syndrome (10), and diabetes mellitus (11–13), characteristics or conditions involving a relative increase in exposure to endogenous estrogens, have been associated with an increased risk of the disease. All three prospective studies that have examined hormone levels observed an increase in endometrial cancer risk with increasing circulating levels of estrogen (14–16). Unopposed estrogen therapy (17–19) and tamoxifen (20, 21), both of which exert proliferative effects on the endometrium, have also been associated with an elevated incidence of the disease. Conversely, increased parity (4, 11, 22, 23) and combination oral contraceptive use (4, 24–26), both characterized by a relatively high degree of exposure to progestogens, are associated with reduced risk of endometrial cancer. Exposure to endogenous or exogenous estrogens not adequately opposed by progestogens leads to an increase in the mitotic activity of endometrial epithelial cells (27). This excessive activity results in increased DNA replication and repair errors, which, in turn, can lead to somatic mutations that may ultimately give rise to endometrial hyperplasia and subsequent malignancy. In this review, we propose that inflammation may work in conjunction with or in addition to estrogen exposure in the development of endometrial cancer (Fig. 1). Support for our hypothesis comes from several observations, including the fact that the effect of the menstrual

cycle resembles an inflammatory process, that unopposed estrogens have an inflammatory effect on the endometrium, and that many of the established risk factors for endometrial cancer can be viewed as producing inflammation in the endometrium. This proinflammatory milieu can initiate and promote neoplastic transformation directly. It can also increase estrogen production, which may facilitate carcinogenesis by disrupting the estrogen-progestogen balance.

The Inflammation-Cancer Link

Epidemiologic studies have documented a relationship between local tissue inflammation and cancer development at that site. Examples include hepatitis and liver cancer (28) and colitis and colon cancer (29). The inverse association between long-term use of nonsteroidal anti-inflammatory drugs (NSAID) and reduced risk of several cancers (30–34) further supports an inflammation-cancer link. Although the means by which local inflammation facilitates cancer development is unknown, inflammatory cells and, in particular, the production of proinflammatory cytokines, such as tumor necrosis factor- α (TNF- α), by both local tissue and infiltrating inflammatory cells, seem to play a key role (reviewed in refs. 35–40). These inflammatory cells induce rapid cell division and produce increased concentrations of free radicals (41) that may subsequently damage DNA. Increased rates of cell turnover are associated with a greater likelihood of replication errors and ineffective DNA repair at key regulatory sites, such as within tumor suppressor genes, which may increase the probability of converting DNA lesions to mutations (42). Moreover, inflammatory cytokines induce a range of inflammatory enzymes, including cyclooxygenase-2 (COX-2). COX-2 cyclizes and oxygenates arachidonic acid, eventually producing prostaglandin E₂ (PGE₂; ref. 43). PGE₂ can facilitate tumorigenesis by increasing production of cytokines and growth factors necessary for tumor growth, invasion, and metastases, including interleukin (IL)-6, IL-8, vascular endothelial growth factor, and matrix metalloproteases (44). PGE₂ also increases production of inducible nitric oxide

Received 7/6/05; revised 9/19/05; accepted 9/27/05

Grant support: National Cancer Institute grant K07-CA80668.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Requests for reprints: Francesmary Modugno, Department of Epidemiology, Graduate School of Public Health, University of Pittsburgh, 516A Parran Hall, 130 DeSoto Street, Pittsburgh, PA 15261. Phone: 412-383-2601; Fax: 412-383-2653. E-mail: modugno+@pitt.edu

Copyright © 2005 American Association for Cancer Research.

doi:10.1158/1055-9965.EPI-05-0493

Table 1. Risk factors for endometrial cancer in relation to the estrogen/progestogen imbalance hypothesis and the proposed inflammation hypothesis

Risk/protective factor	Proposed mechanism—estrogen/progestogen imbalance hypothesis*	Proposed mechanism—inflammation hypothesis
Risk factors		
Unopposed estrogen therapy use	Unopposed estrogen exposure in the endometrium	Increased exposure to estrogens increases inflammatory response in the endometrium
Obesity	Increased systemic exposure to unopposed estrogens via aromatization of androgens in adipose tissue and via decreased sex hormone-binding globulin production Decreased progesterone exposure leading to unopposed estrogen exposure due to anovulation	Increased proinflammatory systemic milieu
Diabetes mellitus	?	Increased proinflammatory systemic milieu
Polycystic ovary syndrome	Increased systemic exposure to unopposed estrogens via aromatization of androgens	Increased proinflammatory systemic milieu
Early menarche/late menopause	Increased lifetime exposure to estrogen in the endometrium	Increased lifetime exposure to inflammation via increased number of menstruations
Anovulation	Decreased progesterone production leading to unopposed estrogen exposure in the endometrium	Decreased exposure to progesterone increases inflammatory response in endometrium
Menorrhagia	?	Increased exposure to inflammation via menstruation
Protective factors		
Pregnancy	Increased exposure to progesterone	Decreases exposure to menstruation Increased exposure to anti-inflammatory effect of progesterone Decreased inflammatory milieu in the uterus
Smoking	Decreased exposure to estrogens due to increased metabolic clearance and production of less estrogenic metabolites	Production of anti-inflammatory estrogen metabolite
Oral contraceptive use	Increased exposure to progestins	Increased exposure to anti-inflammatory progestins Decreased exposure to menstruation

*Adapted from ref. 145.

synthetase, an enzyme involved in free radical generation (44). Further evidence for a role of COX-2 in cancer formation comes from the observation that carcinogenesis is inhibited in mice treated with COX-2-selective inhibitors and in COX-2 knockout mice (45, 46). Hence, neoplastic transformation may be initiated by DNA damage incurred by cells as a result of free radical generation. These initiated cells may be further promoted via COX-2-mediated up-regulation of PGE₂, which causes the production of factors supporting growth, invasion, and metastases.

More recently, laboratory data suggest that the molecular pathway underlying the cancer inflammation association involves the nuclear factor- κ B (NF- κ B) transcription factor and its inhibitor κ B kinase (IKK) complex. NF- κ B is a transcription factor that regulates apoptosis, cell proliferation, and cell growth arrest, as well as enhances angiogenesis via vascular endothelial growth factor expression (47). The NF- κ B pathway is also a crucial second messenger system for inflammatory cytokine signaling (48). In most normal cells, NF- κ B remains in the cytoplasm and is inactive until the cell is stimulated by an appropriate ligand (47). One group of activators of NF- κ B are the proinflammatory cytokines, including TNF- α and IL-1 β , which after complexing to their receptors recruit kinases that activate the IKK complex. The activated IKK complex phosphorylates I κ B, the inhibitory chaperone molecule bound to NF- κ B. NF- κ B is then liberated and translocated to the nucleus, where it activates a variety of genes, including those involved in inflammation and proliferation (49). Constitutive IKK activity resulting in altered NF- κ B

activation is a hallmark for inflammatory diseases (50). Liberated NF- κ B protein may also activate malignancy-promoting signaling pathways in both cancer cells and tumor-associated inflammatory cells (51), and aberrant NF- κ B activity has been reported for several cancers, including estrogen-associated breast cancer (52, 53). Recently, animal models have further elucidated the role of NF- κ B in tumorigenesis. In an inflammation-associated murine model of liver cancer, TNF- α produced by adjacent stromal cells activated NF- κ B in hepatocytes undergoing malignant transformation (54). Inhibition of TNF- α in the stromal cells subsequently induced hepatocyte apoptosis, thereby reducing tumor formation. Moreover, in an inflammation-associated model of colon cancer, deletion of IKK-B in intestinal epithelial cells increased apoptosis, thereby reducing the development of intestinal tumors, although IKK-B deletion did not decrease epithelial inflammation (54). In addition, selective deletion of IKK-B from inflammatory infiltrates in malignant and premalignant tumors resulted in decreased proinflammatory cytokine expression within the infiltrates as well as reduced tumor formation. Together, these data suggest that the NF- κ B pathway invoked by proinflammatory cytokines may be involved in tumor promotion by inhibiting apoptosis in initiated cells as well as by further stimulating production of proinflammatory cytokines by myeloid and lymphoid cells within the tumor mass. These proinflammatory cells can then feed back into the NF- κ B pathway, further promoting the proliferative, antiapoptotic, and proinflammatory process (51).

Thus, chronic inflammation with its subsequent generation of free radicals and up-regulation of COX-2 can lead to a cascade of events that may eventually initiate malignant transformation. Initiated cells can be further promoted by NF-κB proteins freed as a result of up-regulated proinflammatory cytokines. Activated NF-κB both inhibits apoptosis and stimulates production of proinflammatory cytokines, which can further promote proliferation of initiated cells. Hence, tissue exposed to chronic inflammation or to a proinflammatory milieu may be more susceptible to the carcinogenic process.

The Human Endometrium and the Menstrual Cycle: Shared Features with a Chronic Inflammatory Process. The human endometrium consists of glandular and surface epithelia, a surrounding stroma, and a vascular system found only in menstruating species (55). Throughout the childbearing years, the endometrium undergoes cycles of rapid growth, remodeling, differentiation, and angiogenesis. These changes are directly and indirectly caused by steroid hormones and are the result of cytokines synthesized and released by the epithelial, stromal, and vascular cells of the endometrium (56). During the menstrual cycle, the endometrium goes through the proliferative phase, marked by an increase in endometrial thickness; the secretory phase, characterized by the development of spiral arteries and stromal edema after ovulation; and the menstrual phase, marked by the shedding of the endometrial lining. During the three phases, steroid hormone levels vary. Estradiol increases and reaches its peak during the mid-proliferative phase. There is another, smaller increase and peak in estradiol during the midsecretory phase. Progesterone levels are extremely low during the menstrual and early proliferative phases, begin to increase toward the end of the proliferative phase, and peak during the midsecretory phase (57).

The menstrual phase of the endometrium includes inflammation as a physiologic component. Thus, in the absence of a pregnancy, the endometrium can be viewed as being cyclically exposed to a chronic inflammatory-like process. Inflammation is the response of the body to tissue damage resulting from a physical, chemical, or infectious agent, and involves the release of a host of factors, including cytokines, growth factors, and prostaglandins (40). After an insult, thrombi are formed, polynuclear and later mononuclear cells are recruited, fibroblasts proliferate, angiogenesis occurs, and collagen is deposited. The granulation tissue is gradually replaced by a scar and the epithelial tissue is repaired. A similar "injury" scene with its resulting cascade of inflammatory events occurs during the endometrial cycle. During the late endometrial secretory phase, there is a loss of integrity of the lysosomes and acid

hydrolases are released, leading to tissue damage including autodigestion of several cell components, including the membrane (58). The resulting injury causes platelets to amass, thrombi to form, and prostaglandins to be released (59). During this period, ischemic necrosis due to contraction of the endometrial vessels occurs and stromal granulocytes infiltrate the endometrium, reaching a peak during the menstrual phase (60). The vasoconstriction-vasodilation cycles and resulting hypoxia releases a plethora of reactive oxidative species resulting in a degradation of IκB proteins (61) and activation of NF-κB (62, 63). NF-κB induces COX-2, prostaglandin, and inflammatory cytokine release (56), a response that ultimately leads to menstruation (61). Endometrial damage is most apparent during the early menstrual phase when the epithelial and surrounding stroma are detached from the underlying basal layer (60) leaving a thin, denuded endometrium that retains the basal layer and a small portion of the surface epithelium. Migration and proliferation of these epithelial cells, together with angiogenesis of the arterial stumps left in the basal layer, are the main means of endometrial surface repair.

Given the inflammatory component of menstruation (64), it is not surprising that menstruation involves the synthesis and release of inflammatory factors, including cytokines, growth factors, COX-2, and prostaglandins (56, 60, 65), as well as activation of NF-κB (63). Conceivably, these factors could bear on the initiation and progression of endometrial malignancies (66, 67).

Endometrial Cancer Risk Factors Are Associated with a Proinflammatory Milieu. Host factors as well as lifestyle factors play a role in cytokine expression and, hence, can influence the inflammatory nature of the local and systemic milieu. For example, systemic IL-6 and TNF-α levels increase with age (68, 69) and body mass index (70, 71). Indeed, evidence suggests that obesity, a strong risk factor for endometrial cancer, may be a systemic inflammatory condition (72). In addition to elevated serum levels of IL-6 and TNF-α, healthy obese individuals have elevated circulating levels of C-reactive protein, leptin, and macrophage migration inhibitory factor, three markers of inflammation (73, 74). These elevated circulating levels are attributed to increased production of these and other proinflammatory proteins within adipose tissue, and, in the case of C-reactive protein, within the liver as a result of increased expression of IL-6 by adipocytes (75). Thus, proinflammatory cytokines secreted by adipose tissue modulate the immune system in favor of a state characterized by chronic low-level systemic inflammation (76). Given the high population attributable risk for endometrial cancer associated with excess adiposity [~56.8% in the

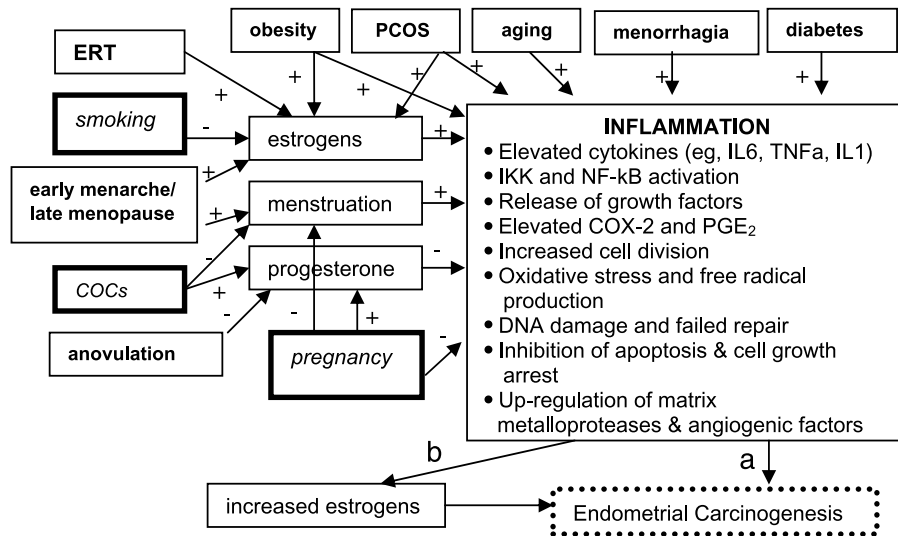


Figure 1. Proposed relationships among endometrial cancer risk/protective factors, inflammation, and endometrial carcinogenesis. Endometrial cancer risk factors (*in bold*) either influence inflammation directly or influence factors that increase inflammation (estrogen, menstruation) or decrease inflammation (progesterone). Protective factors (*in italics*) exert the opposite effects. The effects of inflammation can cause mutagenesis, ultimately leading to endometrial carcinogenesis either directly (**a**) or indirectly (**b**) by increasing estrogen levels. *ERT*, unopposed estrogen therapy; *COC*, combined oral contraceptives; *PCOS*, polycystic ovary syndrome.

United States and 45.2% in Europe (77) for body mass index $>25.0 \text{ kg/m}^2$], it is important to evaluate the means by which obesity exerts its detrimental influence. There is also mounting evidence that diseases involving insulin insensitivity that are associated with endometrial cancer, such as diabetes, also show increased proinflammatory cytokine production, especially IL-1, IL-6, and TNF- α (78, 79). TNF- α is also increased in women with polycystic ovary syndrome, with the effect being most notable in lean women (80). Notably, this proinflammatory milieu associated with endometrial cancer risk factors may increase estrogen production (81).

Estrogen metabolism may also affect cytokine expression. In the liver, estrogens are predominantly metabolized via either the 2-hydroxylation or 16 α -hydroxylation pathways (82). In the uterus, 4-hydroxylation of estradiol has also been observed (83). Because these enzymes compete for a limited substrate pool, an increase in one pathway will reduce the amount of product in the competing pathway. Host factors, such as weight (84), as well as lifestyle factors, such as cigarette smoking (85), influence which pathway predominates. Greater weight is associated with a shift to the 16 α -hydroxylation pathway, whereas cigarette smoking, a putative protective factor (86), is associated with a shift to the 2-hydroxylation pathway. Notably, 2-hydroxyestradiol and its methoxy derivative do not possess uterotrophic activity, whereas both the 4-hydroxylation and 16 α -hydroxylation metabolites are potent estrogens with uterine activity (87). Moreover, 2-methoxy-estradiol (an *O*-methylated product of 2-hydroxylation) seems to inhibit tumor growth, induce apoptosis, and alter microtubule stability (88). It also acts to inhibit the production and actions of both IL-6 and TNF- α (88, 89). Hence, the protective effect of smoking or the increased risk associated with obesity may result from the shift in estrogen metabolism, which exerts a direct effect on proinflammatory cytokines. Finally, whereas embryo implantation evokes an inflammatory response (90), pregnancy is associated with a temporary shift in the cytokine pattern to an anti-inflammatory endometrial milieu (91, 92). In particular, the first trimester decidua is characterized by an increase in I κ B α , the inhibitory chaperone molecule for NF- κ B (63).

Hence, factors associated with endometrial cancer risk, such as obesity and diabetes, also favor the production of proinflammatory cytokines. In contrast, factors associated with reduced risk, such as smoking and pregnancy, favor an anti-inflammatory milieu.

Interrelationship of Sex Hormones with Cytokines and Growth Factors in the Endometrium. Estrogen and progesterone are the well-established regulators of the human endometrium. However, less clear is whether this regulation is through direct interaction with endometrial epithelial cells or indirectly through interaction with stromal and vascular cells (93, 94). Emerging data suggest that estrogen and progesterone exert their effects by influencing the production of cytokines and growth factors found in the endometrium (56, 95), which act on the endometrium as well as the stromal and perivascular cells (93). For example, expression of many cytokines and growth factors in the endometrium is menstrual cycle dependent: Epidermal growth factor, insulin-like growth factor-I, and transforming growth factor- β and their receptors are most highly expressed in the proliferative phase, whereas IL-1, IL-6, and TNF- α are most highly expressed during the secretory and menstrual phases (96, 97). Estrogen directly regulates the endometrial production of many of these cytokines and growth factors as well as their receptors (95, 98, 99).

Progesterone also influences cytokine, growth factor, and prostaglandin production in the endometrium. The ingress of platelets during the proliferative phase of the human endometrium leads to an increase in prostaglandins, which further stimulates platelet ingress (93). This spiraling inflammatory response is blunted by the sudden increase in progesterone

produced by the corpus luteum after ovulation (93). Specifically, progesterone stimulates production of prostaglandin dehydrogenase (100), thereby facilitating the breakdown of prostaglandins. Progestins can also inhibit cytokine-induced transcription of COX-2 (101). In addition, during the proliferative phase of the endometrium, NF- κ B is activated (102) and its resulting inflammatory, proliferative, and antiapoptotic effects are inhibited by progesterone and its receptor (103). The observation that a strong uterine inflammation occurs in knockout mice lacking a progesterone receptor further supports the anti-inflammatory effects of progesterone in the endometrium (104). Thus, natural menstrual cycles lacking an ovulation or an endometrium that is exposed only to unopposed estrogens would be characterized by an inflammatory milieu. As discussed, this proinflammatory milieu may directly facilitate carcinogenesis. Moreover, it can also increase estrogen production (81). In particular, IL-6 can stimulate estrogen synthesis and can act synergistically with TNF- α to enhance the activities of aromatase (105), 17 β -hydroxysteroid dehydrogenase (106, 107), and estrone sulfatase (108), the three enzymes involved in the production of estradiol from androstenedione, estrone, and estone sulfate, respectively. Thus, a proinflammatory milieu can also contribute to an estrogen-progesterone imbalance, possibly predisposing the endometrium to the neoplastic process.

Hence, there is a complex interaction between sex steroid hormones and cytokine/growth factors within the endometrium. In particular, the absence of progesterone can lead to an inflammatory milieu, which can both increase estrogen production and further induce proinflammatory cytokines, thereby potentially creating an environment susceptible to tumor initiation and promotion.

Evidence Linking Inflammation and Endometrial Cancer.

In addition to the circumstantial evidence linking inflammation and endometrial cancer just described, emerging laboratory data suggest a link as well. Studies on excised human tissue show that members of the NF- κ B family are expressed in the proliferating endometrium, in endometrial hyperplasia, and in endometrial carcinoma (102). Notably, a decrease in NF- κ B expression coincides with an increase in apoptosis in low-grade carcinomas, suggesting that up-regulation of NF- κ B may prevent apoptosis in endometrial hyperplasia and in early carcinoma (102). Moreover, NF- κ B factors are aberrantly expressed in the nuclei of a majority of endometrial cancer tumors, with a strong association between the two components of the heterodimer that characterizes the "classic form" of NF- κ B (109). This same classic form has been frequently reported to localize in the nucleus of breast cancer cell lines and some breast tumors (110, 111).

Besides its role as a transcription factor for cell survival and proinflammatory genes, NF- κ B and its inhibitor I κ B seem to regulate expression of COX-2 in endometrial cancer cells (112). In malignant endometrial epithelial cells, NF- κ B induces COX-2, which ultimately leads to enhanced production of PGE₂ and COX-2 in both the malignant cells and in adjacent endometrial stromal cells (113). Up-regulation of COX-2 is found in many cancers, including endometrial cancer, where its expression seems related to tumor aggressiveness (114). Both NF- κ B and COX-2 are expressed in endometrial cancer and during the proliferative phase of the endometrium (102, 115-117). In particular, endometrial explant cultures show that 17 β -estradiol up-regulates COX-2 expression during the early proliferative phase of the endometrium (118) and it is the surge in progesterone as a result of ovulation that may up-regulate I κ B, the inhibitory chaperone molecule for NF- κ B (63), as well as down regulate COX-2 expression (93, 101, 119-121). Consistent with these observations, COX-2 is not found in the secretory phase of the endometrium in women who have ovulated (115) nor in the endometrium of postmenopausal

women using a combined estrogen + progestin hormone therapy regimen (122). Hence, a natural menstrual cycle lacking an ovulation and thus a progesterone surge (and possibly a hormone therapy regimen lacking progestin) would not inactivate NF- κ B nor down-regulate COX-2, leaving the endometrium exposed to greater levels of COX-2 enzyme and subsequently PGE₂.

COX-2 and PGE₂ are also elevated in uterine tissue of women with menorrhagia (excessive menstruation), a known risk factor for endometrial cancer (123). PGE₂ is also commonly elevated in malignant endometrial epithelial cells (113). Moreover, elevated PGE₂ levels caused by COX-2 up-regulation may underlie the transformation of normal endometrium to neoplastic tissue (66). In particular, within the endometrium, elevated COX-2 and PGE₂ can facilitate angiogenesis (124), increase cell proliferation (125), decrease apoptosis (125), inhibit B- and T-cell proliferation and macrophage function (thereby allowing defective cells to proliferate undetected by the immune system; ref. 126) and facilitate tissue invasion (127).

Aspirin has been shown *in vitro* to inhibit endometrial cancer cell growth through the induction of apoptosis in a dose-dependent manner (128). Other NSAIDs have also been shown to reduce endometrial cancer cell proliferation and induce apoptosis in a dose- and time-dependent manner (129, 130). Consistent with data from colon (131) and other cancer cell lines, these laboratory experiments suggest that NSAIDs exert their anticancer effects through both COX-2-dependent and COX-2-independent mechanisms (129). This latter observation suggests that if NSAIDs do protect against endometrial cancer, there may be several underlying mechanisms by which they exert their effects, such as by inhibiting aromatase (132), improving insulin signaling (133-138), trapping reactive oxygen species (139), ameliorating hypoxia/reoxygenation in injured tissues (139-141), and inhibiting macrophage expression of TNF- α (142), a proinflammatory cytokine whose expression varies with estrogen levels in the endometrium. Hence, aspirin and other NSAIDs might exert their effects in the endometrium through both inflammation-dependent and inflammation-independent mechanisms.

Summary and Conclusions

We propose that inflammation may play a role in the genesis of endometrial cancer. In general, chronic local inflammation may predispose to tumor development by generating free radicals and up-regulating COX-2 and PGE₂, which in turn can damage DNA and induce cell proliferation, thus initiating and promoting neoplastic transformation. Chronic inflammation can also dysregulate the NF- κ B pathway, thereby inhibiting apoptosis, blocking cell cycle arrest, and further stimulating production of proinflammatory cytokines. A proinflammatory milieu can feed back into this production cycle, further facilitating tumorigenesis. As discussed, the endometrial cycle resembles a state of cyclic chronic inflammation and endometrial cancer risk factors are associated with a proinflammatory milieu. In premenopausal women, the estrogen-dominated proliferative phase of the endometrium is characterized by an increase in NF- κ B activity and up-regulation of COX-2 and PGE₂, which could lead to a spiraling inflammatory response in the absence of progesterone. A similar effect could be expected in the postmenopausal endometrium of women who receive unopposed estrogen therapy. Menstruation is also associated with elevated COX-2 and PGE₂ expression, as well as with NF- κ B activation. Thus, the consistent epidemiologic risk factors for endometrial cancer—unopposed estrogen use, anovulation, polycystic ovary syndrome, excessive menstruation, early menarche, and late menopause—may be viewed as factors increasing the exposure of the endometrium to inflammation. Moreover, obesity and diabetes, two other

conditions also associated with an increased risk, are also characterized by a shift to a proinflammatory milieu. Pregnancy, which is associated with a reduced risk of endometrial cancer, has the effect of minimizing the number of menstrual cycles, and therefore reducing cumulative exposure to inflammation. Pregnancy also induces a temporary shift to an anti-inflammatory milieu and is associated with an inhibition of NF- κ B activity. Women who smoke cigarettes have relatively low risk of endometrial cancer, and smoking shifts estrogen metabolism to a pathway that seems to inhibit inflammatory cytokines. Together, these data suggest an inflammation-endometrial cancer link. Further support for our hypothesis derives from laboratory data showing that NSAIDs inhibit endometrial cancer cell growth *in vitro*; that NF- κ B activity is increased in the proliferating endometrium, in endometrial hyperplasia, and in endometrial carcinoma; and that elevated levels of COX-2 typically found in the endometrium exposed to unopposed estrogens lead to increases in PGE₂, which can initiate and promote the neoplastic process.

There are several ways to test the hypothesis put forth in this article. First, use of anti-inflammatory medications would be expected to reduce the risk of endometrial cancer. To date, no epidemiologic studies have assessed the association between NSAID use and endometrial cancer. Studies in populations of women with substantial exposure to these medications, such as those with connective tissue diseases, can also elucidate any potential relationship as long as the disease does not inflame the endometrium. Cytokine levels and activity may be altered by cytokine gene variations (143, 144). Thus, individual variations in these genes, either alone or in combination with host and lifestyle factors, may affect endometrial cancer risk. Similarly, susceptibility to the effects of inflammation may be modulated by variation in DNA repair genes, such that individuals with more active repair capabilities may be less susceptible to the effects of endometrial inflammation. The prevalence of these genetic variants in women with and without endometrial cancer can be assessed, thereby shedding light on the biological mechanisms underlying endometrial cancer. Finally, animal models can also be used to assess whether suppression of menstruation-associated inflammation by NSAIDs reduces the development of atypical hyperplasia and other markers of endometrial transformation.

Acknowledgments

We thank Chandra Marriott and Dr. Jeffrey L. Eppinger for their help with the manuscript.

References

1. Brinton LA, Berman ML, Mortel R, et al. Reproductive, menstrual, and medical risk factors for endometrial cancer: results from a case-control study. *Am J Obstet Gynecol* 1992;167:1317-25.
2. La Vecchia C, Franceschi S, Decarli A, Gallus G, Tognoni G. Risk factors for endometrial cancer at different ages. *J Natl Cancer Inst* 1984;73:667-71.
3. Kalandidi A, Tzonou A, Lipworth L, Gamatsi I, Filippa D, Trichopoulos D. A case-control study of endometrial cancer in relation to reproductive, somatometric, and life-style variables. *Oncology* 1996;53:354-9.
4. Pettersson B, Adami HO, Bergstrom R, Johansson ED. Menstruation span—a time-limited risk factor for endometrial carcinoma. *Acta Obstet Gynecol Scand* 1986;65:247-55.
5. McPherson CP, Sellers TA, Potter JD, Bostick RM, Folsom AR. Reproductive factors and risk of endometrial cancer. The Iowa Women's Health Study. *Am J Epidemiol* 1996;143:1195-202.
6. Parazzini F, La Vecchia C, Negri E, Fedele L, Balotta F. Reproductive factors and risk of endometrial cancer. *Am J Obstet Gynecol* 1991;164:522-7.
7. Javert CT, Renning EL. Endometrial Cancer. Survey of 610 cases at Woman's Hospital (1919-1960). *Cancer* 1963;16:1057-71.
8. Wynder EL, Escher GC, Mantel N. An epidemiological investigation of cancer of the endometrium. *Cancer* 1966;19:489-520.
9. Gangemi M, Meneghetti G, Predebon O, Scappatura R, Rocco A. Obesity as a risk factor for endometrial cancer. *Clin Exp Obstet Gynecol* 1987;14:119-22.
10. Hardiman P, Pillay OC, Atiomo W. Polycystic ovary syndrome and endometrial carcinoma. *Lancet* 2003;361:1810-2.
11. Elwood JM, Cole P, Rothman KJ, Kaplan SD. Epidemiology of endometrial cancer. *J Natl Cancer Inst* 1977;59:1055-60.

12. O'Mara BA, Byers T, Schoenfeld E. Diabetes mellitus and cancer risk: a multisite case-control study. *J Chronic Dis* 1985;38:435-41.
13. Parazzini F, La Vecchia C, Negri E, et al. Diabetes and endometrial cancer: an Italian case-control study. *Int J Cancer* 1999;81:539-42.
14. Austin H, Austin JM, Jr., Partridge EE, Hatch KD, Shingleton HM. Endometrial cancer, obesity, and body fat distribution. *Cancer Res* 1991; 51:568-72.
15. Potischman N, Hoover RN, Brinton LA, et al. Case-control study of endogenous steroid hormones and endometrial cancer. *J Natl Cancer Inst* 1996;88:1127-35.
16. Zeleniuch-Jacquotte A, Akhmedkhanov A, Kato I, et al. Postmenopausal endogenous oestrogens and risk of endometrial cancer: results of a prospective study. *Br J Cancer* 2001;84:975-81.
17. Ziel HK, Finkle WD. Increased risk of endometrial carcinoma among users of conjugated estrogens. *N Engl J Med* 1975;293:1167-70.
18. Smith DC, Prentice R, Thompson DJ, Herrmann WL. Association of exogenous estrogen and endometrial carcinoma. *N Engl J Med* 1975;293: 1164-7.
19. Grady D, Gebretsadik T, Kerlikowske K, Ernster V, Petitti D. Hormone replacement therapy and endometrial cancer risk: a meta-analysis. *Obstet Gynecol* 1995;85:304-13.
20. Fisher B, Costantino JP, Redmond CK, Fisher ER, Wickerham DL, Cronin WM. Endometrial cancer in tamoxifen-treated breast cancer patients: findings from the National Surgical Adjuvant Breast and Bowel Project (NSABP) B-14. *J Natl Cancer Inst* 1994;86:527-37.
21. Killackey MA, Hakes TB, Pierce VK. Endometrial adenocarcinoma in breast cancer patients receiving antiestrogens. *Cancer Treat Rep* 1985;69: 237-8.
22. Kvale G, Heuch I, Ursin G. Reproductive factors and risk of cancer of the uterine corpus: a prospective study. *Cancer Res* 1988;48:6217-21.
23. Lambe M, Wu J, Weiderpass E, Hsieh CC. Childbearing at older age and endometrial cancer risk (Sweden). *Cancer Causes Control* 1999;10:43-9.
24. Kelsey JL, LiVolsi VA, Holford TR, et al. A case-control study of cancer of the endometrium. *Am J Epidemiol* 1982;116:333-42.
25. Schlesselman JJ. Risk of endometrial cancer in relation to use of combined oral contraceptives. A practitioner's guide to meta-analysis. *Hum Reprod* 1997;12:1851-63.
26. Weiderpass E, Adami HO, Baron JA, et al. Risk of endometrial cancer following estrogen replacement with and without progestins. *J Natl Cancer Inst* 1999;91:1131-7.
27. Key TJ, Pike MC. The dose-effect relationship between "unopposed" oestrogens and endometrial mitotic rate: its central role in explaining and predicting endometrial cancer risk. *Br J Cancer* 1988;57:205-12.
28. Robinson WS. Hepatitis B virus and hepatocellular carcinoma. In: Parsonnet J, editor. *Microbes and malignancy: infection as a cause of human cancers*. New York: Oxford University Press; 1999.
29. Lewis JD, Deren JJ, Lichtenstein GR. Cancer risk in patients with inflammatory bowel disease. *Gastroenterol Clin North Am* 1999;28:459-77.
30. Thun MJ, Namboodiri MM, Calle EE, Flanders WD, Heath CW, Jr. Aspirin use and risk of fatal cancer. *Cancer Res* 1993;53:1322-7.
31. Schreinemachers DM, Everson RB. Aspirin use and lung, colon, and breast cancer incidence in a prospective study. *Epidemiology* 1994;5:138-46.
32. Paganini-Hill A, Chao A, Ross RK, Henderson BE. Aspirin use and chronic diseases: a cohort study of the elderly. *BMJ* 1989;299:1247-50.
33. Akhmedkhanov A, Toniolo P, Zeleniuch-Jacquotte A, Kato I, Koenig KL, Shore RE. Aspirin and epithelial ovarian cancer. *Prev Med* 2001;33: 682-7.
34. Harris RE, Chlebowski RT, Jackson RD, et al. Breast cancer and nonsteroidal anti-inflammatory drugs: prospective results from the Women's Health Initiative. *Cancer Res* 2003;63:6096-101.
35. O'Byrne KJ, Dalglish AG. Chronic immune activation and inflammation as the cause of malignancy. *Br J Cancer* 2001;85:473-83.
36. Balkwill F, Charles KA, Mantovani A. Smoldering and polarized inflammation in the initiation and promotion of malignant disease. *Cancer Cell* 2005;7:211-7.
37. Schwartzburd PM. Age-promoted creation of a pro-cancer microenvironment by inflammation: pathogenesis of discoordinated feedback control. *Mech Ageing Dev* 2004;125:581-90.
38. Philip M, Rowley DA, Schreiber H. Inflammation as a tumor promoter in cancer induction. *Semin Cancer Biol* 2004;14:433-9.
39. Ristimaki A. Cyclooxygenase 2: from inflammation to carcinogenesis. *Novartis Found Symp* 2004;256:215-21; discussion 21-6, 59-69.
40. Coussens LM, Werb Z. Inflammation and cancer. *Nature* 2002;420:860-7.
41. Hussain SP, Hofseth LJ, Harris CC. Radical causes of cancer. *Nat Rev Cancer* 2003;3:276-85.
42. Ames BN, Gold LS, Willett WC. The causes and prevention of cancer. *Proc Natl Acad Sci U S A* 1995;92:5258-65.
43. Morita I. Distinct functions of COX-1 and COX-2. *Prostaglandins Other Lipid Mediat* 2002;68-9:165-75.
44. Gasparini G, Longo R, Sarmiento R, Morabito A. Inhibitors of cyclooxygenase 2: a new class of anticancer agents? *Lancet Oncol* 2003;4:605-15.
45. Jacoby RF, Seibert K, Cole CE, Kelloff G, Lubet RA. The cyclooxygenase-2 inhibitor celecoxib is a potent preventive and therapeutic agent in the min mouse model of adenomatous polyposis. *Cancer Res* 2000; 60:5040-4.
46. Peluffo GD, Stillitani I, Rodriguez VA, Diament MJ, Klein SM. Reduction of tumor progression and paraneoplastic syndrome development in murine lung adenocarcinoma by nonsteroidal antiinflammatory drugs. *Int J Cancer* 2004;110:825-30.
47. Kumar A, Takada Y, Boriek AM, Aggarwal BB. Nuclear factor- κ B: its role in health and disease. *J Mol Med* 2004;82:434-48.
48. Karin M, Delhase M. The I κ B kinase (IKK) and NF- κ B: key elements of proinflammatory signalling. *Semin Immunol* 2000;12:85-98.
49. Karin M, Ben-Neriah Y. Phosphorylation meets ubiquitination: the control of NF- κ B activity. *Annu Rev Immunol* 2000;18:621-63.
50. Marok R, Winyard PG, Coumbe A, et al. Activation of the transcription factor nuclear factor- κ B in human inflamed synovial tissue. *Arthritis Rheum* 1996;39:583-91.
51. Balkwill F, Coussens LM. Cancer: an inflammatory link. *Nature* 2004;431: 405-6.
52. Romieu-Mourez R, Landesman-Bollag E, Seldin DC, Traish AM, Mercurio F, Sonenshein GE. Roles of IKK kinases and protein kinase CK2 in activation of nuclear factor- κ B in breast cancer. *Cancer Res* 2001; 61:3810-8.
53. Biswas DK, Shi Q, Baily S, et al. NF- κ B activation in human breast cancer specimens and its role in cell proliferation and apoptosis. *Proc Natl Acad Sci U S A* 2004;101:10137-42.
54. Pikarsky E, Porat RM, Stein I, et al. NF- κ B functions as a tumour promoter in inflammation-associated cancer. *Nature* 2004;431:461-6.
55. Blaustein A. *Pathology of the female genital tract*. 2nd ed. New York; 1982.
56. Kelly RW, King AE, Critchley HO. Cytokine control in human endometrium. *Reproduction* 2001;121:3-19.
57. Strauss J, Coutifaris C. The endometrium and myometrium: regulation and dysfunction. In: Yen S, Jaffe R, Barbieri R, editors. *Reproductive endocrinology*. Philadelphia: WB Saunders and Company; 1996.
58. Epifanova OI. Effects of hormones on the cell cycle. In: Baserga R, editor. *The cell cycle and cancer*. New York: M. Dekker; 1971. p. 145.
59. Srivastava KC. Prostaglandins and platelet function. *S Afr J Sci* 1978;74:290.
60. Tabibzadeh S. Human endometrium: an active site of cytokine production and action. *Endocr Rev* 1991;12:272-90.
61. Sugino N, Karube-Harada A, Taketani T, Sakata A, Nakamura Y. Withdrawal of ovarian steroids stimulates prostaglandin F $_{2\alpha}$ production through nuclear factor- κ B activation via oxygen radicals in human endometrial stromal cells: potential relevance to menstruation. *J Reprod Dev* 2004; 50:215-25.
62. Royds JA, Dower SK, Qwarnstrom EE, Lewis CE. Response of tumour cells to hypoxia: role of p53 and NF κ B. *Mol Pathol* 1998;51:55-61.
63. King AE, Critchley HO, Kelly RW. The NF- κ B pathway in human endometrium and first trimester decidua. *Mol Hum Reprod* 2001;7:175-83.
64. Baird DT, Cameron ST, Critchley HO, et al. Prostaglandins and menstruation. *Eur J Obstet Gynecol Reprod Biol* 1996;70:15-7.
65. Kelly RW. Pregnancy maintenance and parturition: the role of prostaglandin in manipulating the immune and inflammatory response. *Endocr Rev* 1994;15:684-706.
66. Sales KJ, Jabbour HN. Cyclooxygenase enzymes and prostaglandins in pathology of the endometrium. *Reproduction* 2003;126:559-67.
67. Balkwill F, Mantovani A. Inflammation and cancer: back to Virchow? *Lancet* 2001;357:539-45.
68. Daynes RA, Araneo BA, Ershler WB, Maloney C, Li GZ, Ryu SY. Altered regulation of IL-6 production with normal aging. Possible linkage to the age-associated decline in dehydroepiandrosterone and its sulfated derivative. *J Immunol* 1993;150:5219-30.
69. Wei J, Xu H, Davies JL, Hemmings GP. Increase of plasma IL-6 concentration with age in healthy subjects. *Life Sci* 1992;51:1953-6.
70. Mohamed-Ali V, Goodrick S, Rawesh A, et al. Subcutaneous adipose tissue releases interleukin-6, but not tumor necrosis factor- α , *in vivo*. *J Clin Endocrinol Metab* 1997;82:4196-200.
71. Hotamisligil GS, Arner P, Caro JF, Atkinson RL, Spiegelman BM. Increased adipose tissue expression of tumor necrosis factor- α in human obesity and insulin resistance. *J Clin Invest* 1995;95:2409-15.
72. Das UN. Is obesity an inflammatory condition? *Nutrition* 2001;17:953-66.
73. Dandona P, Aljada A, Ghanim H, et al. Increased plasma concentration of macrophage migration inhibitory factor (MIF) and MIF mRNA in mononuclear cells in the obese and the suppressive action of metformin. *J Clin Endocrinol Metab* 2004;89:5043-7.
74. Visser M, Bouter LM, McQuillan GM, Wener MH, Harris TB. Elevated C-reactive protein levels in overweight and obese adults. *JAMA* 1999;282:2131-5.
75. Banks RE, Forbes MA, Storr M, et al. The acute phase protein response in patients receiving subcutaneous IL-6. *Clin Exp Immunol* 1995; 102:217-23.
76. Wisse BE. The inflammatory syndrome: the role of adipose tissue cytokines in metabolic disorders linked to obesity. *J Am Soc Nephrol* 2004;15:2792-800.
77. Calle EE, Kaaks R. Overweight, obesity and cancer: epidemiological evidence and proposed mechanisms. *Nat Rev Cancer* 2004;4:579-91.
78. Pradhan AD, Manson JE, Rifai N, Buring JE, Ridker PM. C-reactive protein, interleukin 6, and risk of developing type 2 diabetes mellitus. *JAMA* 2001;286:327-34.
79. Grimbale RF. Inflammatory status and insulin resistance. *Curr Opin Clin Nutr Metab Care* 2002;5:551-9.
80. Gonzalez F, Thusu K, Abdel-Rahman E, Prabhala A, Tomani M, Dandona P. Elevated serum levels of tumor necrosis factor α in normal-weight women with polycystic ovary syndrome. *Metabolism* 1999;48:437-41.

81. Purohit A, Newman SP, Reed MJ. The role of cytokines in regulating estrogen synthesis: implications for the etiology of breast cancer. *Breast Cancer Res* 2002;4:65–9.
82. Zhu BT, Conney AH. Functional role of estrogen metabolism in target cells: review and perspectives. *Carcinogenesis* 1998;19:1–27.
83. Lihhr JG, Ricci MJ, Jefcoate CR, Hannigan EV, Hokanson JA, Zhu BT. 4-Hydroxylation of estradiol by human uterine myometrium and myoma microsomes: implications for the mechanism of uterine tumorigenesis. *Proc Natl Acad Sci U S A* 1995;92:9220–4.
84. Longcope C, Gorbach S, Goldin B, et al. The effect of a low fat diet on estrogen metabolism. *J Clin Endocrinol Metab* 1987;64:1246–50.
85. Michnovicz JJ, Hershcopf RJ, Naganuma H, Bradlow HL, Fishman J. Increased 2-hydroxylation of estradiol as a possible mechanism for the anti-estrogenic effect of cigarette smoking. *N Engl J Med* 1986;315:1305–9.
86. Terry PD, Rohan TE, Franceschi S, Weiderpass E. Cigarette smoking and the risk of endometrial cancer. *Lancet Oncol* 2002;3:470–80.
87. Martucci C, Fishman J. Direction of estradiol metabolism as a control of its hormonal action-uterotrophic activity of estradiol metabolites. *Endocrinology* 1977;101:1709–15.
88. Purohit A, Singh A, Ghilchik MW, Reed MJ. Inhibition of tumor necrosis factor α -stimulated aromatase activity by microtubule-stabilizing agents, paclitaxel and 2-methoxyestradiol. *Biochem Biophys Res Commun* 1999;261:214–7.
89. Purohit A, Reed MJ. Regulation of estrogen synthesis in postmenopausal women. *Steroids* 2002;67:979–83.
90. Finn CA. Implantation, menstruation and inflammation. *Biol Rev Camb Philos Soc* 1986;61:313–28.
91. Wegmann TG, Lin H, Guilbert L, Mosmann TR. Bidirectional cytokine interactions in the maternal-fetal relationship: is successful pregnancy a TH2 phenomenon? *Immunol Today* 1993;14:353–6.
92. Lin H, Mosmann TR, Guilbert L, Tuntipopipat S, Wegmann TG. Synthesis of T helper 2-type cytokines at the maternal-fetal interface. *J Immunol* 1993;151:4562–73.
93. Kelly RW, King AE, Critchley HO. Inflammatory mediators and endometrial function—focus on the perivascular cell. *J Reprod Immunol* 2002;57:81–93.
94. Tabibzadeh S, Sun XZ. Cytokine expression in human endometrium throughout the menstrual cycle. *Hum Reprod* 1992;7:1214–21.
95. Tabibzadeh S. Cytokines and the hypothalamic-pituitary-ovarian-endometrial axis. *Hum Reprod* 1994;9:947–67.
96. Tazuke SI, Giudice LC. Growth factors and cytokines in endometrium, embryonic development, and maternal: embryonic interactions. *Semin Reprod Endocrinol* 1996;14:231–45.
97. von Wolff M, Thaler CJ, Zepf C, Becker V, Beier HM, Strowitzki T. Endometrial expression and secretion of interleukin-6 throughout the menstrual cycle. *Gynecol Endocrinol* 2002;16:121–9.
98. Jacobs AL, Sehgal PB, Julian J, Carson DD. Secretion and hormonal regulation of interleukin-6 production by mouse uterine stromal and polarized epithelial cells cultured *in vitro*. *Endocrinology* 1992;131:1037–46.
99. Laird SM, Li TC, Bolton AE. The production of placental protein 14 and interleukin 6 by human endometrial cells in culture. *Hum Reprod* 1993;8:793–8.
100. van der Burg B, van der Saag PT. Nuclear factor- κ B/steroid hormone receptor interactions as a functional basis of anti-inflammatory action of steroids in reproductive organs. *Mol Hum Reprod* 1996;2:433–8.
101. Ishihara O, Matsuoka K, Kinoshita K, Sullivan MH, Elder MG. Interleukin-1 β -stimulated PGE₂ production from early first trimester human decidual cells is inhibited by dexamethasone and progesterone. *Prostaglandins* 1995;49:15–26.
102. Vaskivuo TE, Stenback F, Tapanainen JS. Apoptosis and apoptosis-related factors Bcl-2, Bax, tumor necrosis factor- α , and NF- κ B in human endometrial hyperplasia and carcinoma. *Cancer* 2002;95:1463–71.
103. Davies S, Dai D, Feldman I, Pickett G, Leslie KK. Identification of a novel mechanism of NF- κ B inactivation by progesterone through progesterone receptors in Hec50co poorly differentiated endometrial cancer cells: induction of A20 and ABIN-2. *Gynecol Oncol* 2004;94:463–70.
104. Lydon JP, DeMayo FJ, Funk CR, et al. Mice lacking progesterone receptor exhibit pleiotropic reproductive abnormalities. *Genes Dev* 1995;9:2266–78.
105. Macdiarmid F, Wang D, Duncan LJ, Purohit A, Ghilchick MW, Reed MJ. Stimulation of aromatase activity in breast fibroblasts by tumor necrosis factor α . *Mol Cell Endocrinol* 1994;106:17–21.
106. Duncan LJ, Coldham NG, Reed MJ. The interaction of cytokines in regulating oestradiol 17 β -hydroxysteroid dehydrogenase activity in MCF-7 cells. *J Steroid Biochem Mol Biol* 1994;49:63–8.
107. Adams EF, Rafferty B, White MC. Interleukin 6 is secreted by breast fibroblasts and stimulates 17 β -oestradiol oxidoreductase activity of MCF-7 cells: possible paracrine regulation of breast 17 β -oestradiol levels. *Int J Cancer* 1991;49:118–21.
108. Purohit A, Duncan LJ, Wang DY, Coldham NG, Ghilchick MW, Reed MJ. Paracrine control of oestrogen production in breast cancer. *Endocr Relat Cancer* 1997;4:323–30.
109. Pallares J, Martinez-Guitarte JL, Dolcet X, et al. Abnormalities in the NF- κ B family and related proteins in endometrial carcinoma. *J Pathol* 2004;204:569–77.
110. Yoshimatsu K, Golijanin D, Paty PB, et al. Inducible microsomal prostaglandin E synthase is overexpressed in colorectal adenomas and cancer. *Clin Cancer Res* 2001;7:3971–6.
111. Nakshatri H, Bhat-Nakshatri P, Martin DA, Goulet RJ, Jr., Sledge GW, Jr. Constitutive activation of NF- κ B during progression of breast cancer to hormone-independent growth. *Mol Cell Biol* 1997;17:3629–39.
112. St-Germain ME, Gagnon V, Parent S, Asselin E. Regulation of COX-2 protein expression by Akt in endometrial cancer cells is mediated through NF- κ B/I κ B pathway. *Mol Cancer* 2004;3:7.
113. Tamura M, Sebastian S, Yang S, et al. Up-regulation of cyclooxygenase-2 expression and prostaglandin synthesis in endometrial stromal cells by malignant endometrial epithelial cells. A paracrine effect mediated by prostaglandin E₂ and nuclear factor- κ B. *J Biol Chem* 2002;277:26208–16.
114. Ferrandina G, Legge F, Ranelletti FO, et al. Cyclooxygenase-2 expression in endometrial carcinoma: correlation with clinicopathologic parameters and clinical outcome. *Cancer* 2002;95:801–7.
115. Uotila PJ, Erkkola RU, Klemi PJ. The expression of cyclooxygenase-1 and -2 in proliferative endometrium and endometrial adenocarcinoma. *Ann Med* 2002;34:428–33.
116. Cao QJ, Einstein MH, Anderson PS, Runowicz CD, Balan R, Jones JG. Expression of COX-2, Ki-67, cyclin D1, and P21 in endometrial endometrioid carcinomas. *Int J Gynecol Pathol* 2002;21:147–54.
117. Jeon YT, Kang S, Kang DH, et al. Cyclooxygenase-2 and p53 expressions in endometrial cancer. *Cancer Epidemiol Biomarkers Prev* 2004;13:1538–42.
118. Punyadeera C, Dunselman G, Marbaix E, et al. Triphasic pattern in the *ex vivo* response of human proliferative phase endometrium to oestrogens. *J Steroid Biochem Mol Biol* 2004;92:175–85.
119. Kelly RW, Smith SK. Progesterone and antiprogesterins, a comparison of their effect on prostaglandin production by human secretory phase endometrium and decidua. *Prostaglandins Leukoc Med* 1987;29:181–6.
120. Abel MH, Baird DT. The effect of 17 β -estradiol and progesterone on prostaglandin production by human endometrium maintained in organ culture. *Endocrinology* 1980;106:1599–606.
121. Critchley HO, Jones RL, Lea RG, et al. Role of inflammatory mediators in human endometrium during progesterone withdrawal and early pregnancy. *J Clin Endocrinol Metab* 1999;84:240–8.
122. Hsu SC, Long CY, Yang CH, Wu CH, Chen CH, Liu FI. Cyclooxygenase-2 expression in the endometrium at the end of 2 years' continuous combined hormone replacement therapy. *Maturitas* 2003;46:295–9.
123. Makarainen L, Ylikorkala O. Primary and myoma-associated menorrhagia: role of prostaglandins and effects of ibuprofen. *Br J Obstet Gynaecol* 1986;93:974–8.
124. Tsujii M, Kawano S, Tsuji S, Sawaoka H, Hori M, DuBois RN. Cyclooxygenase regulates angiogenesis induced by colon cancer cells. *Cell* 1998;93:705–16.
125. Tsujii M, DuBois RN. Alterations in cellular adhesion and apoptosis in epithelial cells overexpressing prostaglandin endoperoxide synthase 2. *Cell* 1995;83:493–501.
126. DeWitt DL. Prostaglandin endoperoxide synthase: regulation of enzyme expression. *Biochim Biophys Acta* 1991;1083:121–34.
127. Tsujii M, Kawano S, DuBois RN. Cyclooxygenase-2 expression in human colon cancer cells increases metastatic potential. *Proc Natl Acad Sci U S A* 1997;94:3336–40.
128. Arango HA, Icelly S, Roberts WS, Cavanagh D, Becker JL. Aspirin effects on endometrial cancer cell growth. *Obstet Gynecol* 2001;97:423–7.
129. Gao J, Niwa K, Sun W, et al. Non-steroidal anti-inflammatory drugs inhibit cellular proliferation and upregulate cyclooxygenase-2 protein expression in endometrial cancer cells. *Cancer Sci* 2004;95:901–7.
130. Li HL, Zhang HW, Chen DD, Zhong L, Ren XD, St-Tu R. JTE-522, a selective COX-2 inhibitor, inhibits cell proliferation and induces apoptosis in RL95–2 cells. *Acta Pharmacol Sin* 2002;23:631–7.
131. Piazza GA, Alberts DS, Hixson LJ, et al. Sulindac sulfone inhibits azoxymethane-induced colon carcinogenesis in rats without reducing prostaglandin levels. *Cancer Res* 1997;57:2909–15.
132. Zhao Y, Agarwal VR, Mendelson CR, Simpson ER. Estrogen biosynthesis proximal to a breast tumor is stimulated by PGE₂ via cyclic AMP, leading to activation of promoter II of the CYP19 (aromatase) gene. *Endocrinology* 1996;137:5739–42.
133. Fang Y, Foye W, Robinson SM, Jenkins HJ. Hypoglycemic activity and chemical structure of the salicylates. *J Pharm Sci* 1968;57:2111–6.
134. Graef I, Gibbons DM. Salicylates and carbohydrate metabolism. *Diabetes* 1960;9:416–8.
135. Powell ED, Field RA. Studies on salicylates and complement in diabetes. *Diabetes* 1966;15:730–3.
136. McRae JR, Chen M, Robertson RP. Improvement of defective insulin responses to glucose, arginine, and β -adrenergic stimulation in diabetics by sodium salicylate. *Adv Prostaglandin Thromboxane Res* 1980;8:1287–9.
137. Baron SH. Salicylates as hypoglycemic agents. *Diabetes Care* 1982;5:64–71.
138. Hundal RS, Petersen KF, Mayerson AB, et al. Mechanism by which high-

- dose aspirin improves glucose metabolism in type 2 diabetes. *J Clin Invest* 2002;109:1321–6.
139. Amann R, Peskar BA. Anti-inflammatory effects of aspirin and sodium salicylate. *Eur J Pharmacol* 2002;447:1–9.
140. Colantoni A, de Maria N, Caraceni P, Bernardi M, Floyd RA, Van Thiel DH. Prevention of reoxygenation injury by sodium salicylate in isolated-perfused rat liver. *Free Radic Biol Med* 1998;25:87–94.
141. van Jaarsveld H, Kuyt JM, van Zyl GF, Barnard HC. Salicylate in the perfusate during ischemia/reperfusion prevented mitochondrial injury. *Res Commun Mol Pathol Pharmacol* 1994;86:287–95.
142. Shackelford RE, Alford PB, Xue Y, Thai SF, Adams DO, Pizzo S. Aspirin inhibits tumor necrosis factor α gene expression in murine tissue macrophages. *Mol Pharmacol* 1997;52:421–9.
143. Kroeger KM, Steer JH, Joyce DA, Abraham LJ. Effects of stimulus and cell type on the expression of the –308 tumour necrosis factor promoter polymorphism. *Cytokine* 2000;12:110–9.
144. Terry CF, Loukaci V, Green FR. Cooperative influence of genetic polymorphisms on interleukin 6 transcriptional regulation. *J Biol Chem* 2000;275:18138–44.
145. Akhmedkhanov A, Zeleniuch-Jacquotte A, Toniolo P. Role of exogenous and endogenous hormones in endometrial cancer: review of the evidence and research perspectives. *Ann N Y Acad Sci* 2001;943:296–315.

Inflammation and Endometrial Cancer: A Hypothesis

Francesmary Modugno, Roberta B. Ness, Chu Chen, et al.

Cancer Epidemiol Biomarkers Prev 2005;14:2840-2847.

Updated version Access the most recent version of this article at:
<http://cebp.aacrjournals.org/content/14/12/2840>

Cited articles This article cites 141 articles, 29 of which you can access for free at:
<http://cebp.aacrjournals.org/content/14/12/2840.full#ref-list-1>

Citing articles This article has been cited by 19 HighWire-hosted articles. Access the articles at:
<http://cebp.aacrjournals.org/content/14/12/2840.full#related-urls>

E-mail alerts [Sign up to receive free email-alerts](#) related to this article or journal.

Reprints and Subscriptions To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions To request permission to re-use all or part of this article, use this link
<http://cebp.aacrjournals.org/content/14/12/2840>.
Click on "Request Permissions" which will take you to the Copyright Clearance Center's (CCC) Rightslink site.