Urinary Biomarkers of Oxidative Status in a Clinical Model of Oxidative Assault

Dora Il’yasova1, Ivan Spasojevic1, Frances Wang1, Adviye A. Tolun2, Karel Base1, Sarah P. Young2, P. Kelly Marcom1, Jeffrey Marks1, Gabriel Mixon3, Richard DiGiulio4, and David S. Millington2

Abstract

Background: We used doxorubicin-based chemotherapy as a clinical model of oxidative assault in humans.

Methods: The study recruited newly diagnosed breast cancer patients (n = 23). Urine samples were collected immediately before (T0) and at 1 hour (T1) and 24 hours (T24) after i.v. administration of treatment. Measurements included allantoin and the isoprostanes iP(2α)-III, iP(2α)-VI, and 8,12-iso-iP(2α)-VI along with the prostaglandin 2,3-dinor-iP(2α)-III, a metabolite of iP(2α)-III. All biomarkers were quantified using liquid chromatography–tandem mass spectrometry.

Results: In all subjects, the levels of the biomarkers increased at T1: allantoin by 22% (P = 0.06), iP(2α)-III by 62% (P < 0.05), iP(2α)-VI by 41% (P < 0.05), 8,12-iso-iP(2α)-VI by 58% (P < 0.05), and 2,3-dinor-iP(2α)-III by 52% (P < 0.05). At T24, the F2-isoprostanes returned to their baseline levels; the levels of allantoin continued to increase, although the T24–T0 difference was not statistically significant.

Conclusions: These results indicate that urinary F2-isoprostanes are valid biomarkers and allantoin is a promising biomarker of oxidative status in humans.

Impact: The levels of biomarkers change quickly in response to oxidative assault and can be used to monitor oxidative status in humans in response to treatments related either to generation of free radicals (chemotherapy and radiation therapy) or to antioxidants (inborn metabolic diseases and Down syndrome). Cancer Epidemiol Biomarkers Prev; 19(6); 1506–10. ©2010 AACR.

Introduction

There is consensus that existing and newly developed indices of oxidative status should be validated against known oxidative insults in animal models and in human studies (1, 2). In response to this well-recognized need, the National Institute of Environmental Health Sciences has established an initiative to conduct a comparative study of biomarkers of oxidative stress (BOSS). The BOSS project tests the responsiveness of commonly used oxidative indices in an established model of oxidative stress—carbon tetrachloride (CCl4) poisoning in rodents. We developed an analogous approach to validate commonly used oxidative indices in humans; specifically, we used doxorubicin-based chemotherapy in breast cancer patients as a clinical model of oxidative assault.

Doxorubicin-based chemotherapy satisfies two major requirements for a model of oxidative assault. First, it is based on an established oxidative stressor; generation of hydroxyl radicals at pharmacologic levels of doxorubicin (1 μmol/L) has been shown in animal studies by direct measurement with electron spin resonance spectroscopy (3, 4). Second, doxorubicin-based chemotherapy presents a well-controlled oxidative exposure with an exact dose given to each patient at a certain time, which allows for timed collection of biological samples.

We examined the responsiveness of several indices of oxidative damage and antioxidant defense measured in blood and urine. The findings related to the blood measurements are reported elsewhere (5). Here, we report our findings on urinary biomarkers of oxidative lipid modification: four F2-isoprostanes and one oxidative product of uric acid (allantoin). Urinary F2-isoprostanes are well-studied indices of lipid peroxidation that have been validated by the BOSS project (6, 7). Allantoin has recently emerged as a promising biomarker of oxidative status that is specific to humans (therefore, it cannot be evaluated in animal models). Humans (as well as other hominoid primates and birds) lack urate oxidation—therefore, it cannot be evaluated in animal models). Humans (as well as other hominoid primates and birds) lack urate oxidation; recent studies indicate that uric acid, the terminal product of purine metabolism in humans, is a potent antioxidant and scavenger of reactive oxygen species (9). Allantoin in human bodily fluids is generated by nonenzymatic oxidation by free...
radicals. Gruber and colleagues (10) recently showed in a human study that allantoin increases in nasal lavage fluid in response to ozone exposure. In this study, we evaluated responsiveness of allantoin to a systemic oxidative stressor.

Materials and Methods

Study subjects
We recruited 23 women with newly diagnosed breast cancer scheduled to undergo standard chemotherapy (doxorubicin, 60 mg/m² and cyclophosphamide, 600 mg/m² × 4). The eligibility criteria were the following: (a) histologically confirmed invasive breast cancer; (b) no evidence of metastasis; (c) age ≥18 years; (d) ≥2 weeks since surgery; (e) adequate bone marrow, hepatic, and renal function; and (f) ability to give informed consent. Exclusion criteria included concomitant anticancer medications with myelosuppression effects, low functional status, serious comorbidities, pregnancy, and prior treatment with weekly paclitaxel. The study protocol was approved by the Duke University Medical Center Institutional Review Board.

Urine samples
Urinary samples were collected from participants at three time points: immediately before treatment (T0) and after treatment at 1 hour (T1) and 24 hours (T24). Urine samples were stored at −80°C.

Urinary creatinine
Creatinine was assayed by a fast electrospray ionization–tandem mass spectrometry (MS/MS) method as described previously (11).

Four urinary F2-isoprostanes
Four isomers of F2-isoprostanes—iPF(2α)-III, 2,3-dinor-iPF(2α)-III, iPF(2α)-VI, and 8,12-iso-iPF(2α)-VI—were quantified by liquid chromatography–MS/MS (LC-MS/MS; refs. 12, 13) on Shimadzu 20A series LC and Applied Biosystems API 4000 QTrap MS/MS instruments. Based on creatinine measurements, the urine samples were diluted to 0.65 mg/mL creatinine, and samples with creatinine levels equal to or below this value were analyzed without dilution. Sample preparation included addition of internal standards [iPF(2α)-III-d4, 8,12-iso-iPF(2α)-VI-d11, and iPF(2α)-VI-d14] and 10 μL of 1 mol/L HCl, washing of samples (500 μL) with 1 mL hexane, extraction of the analytes by ethyl acetate/hexane mixture (3:1, v/v), evaporation of the liquid, and resuspension of the residue in 150 μL of a mixture containing 70% mobile phase A (0.1% formic acid in water) and 30% methanol. The samples (100 μL) were then injected into the LC-MS/MS system. Two solid core C18 columns (Phenomenex Kinetex C18, 150 × 4.6 mm) in series were used to achieve chromatographic separation of the F2-isoprostane isomers. The mass spectrometer was operated in negative mode with the following multiple reaction monitoring transitions (m/z): 353/193 [iPF(2α)-III], 357/197 [iPF(2α)-III-d4], 325/237 [2,3-dinor-iPF(2α)-III], 353/115 [iPF(2α)-VI and 8,12-iso-iPF(2α)-VI], 364/115 [iPF(2α)-VI-d11], and 357/115 [8,12-iso-iPF(2α)-VI-d4]. Calibration samples covering the expected range of concentrations were prepared by adding pure material into pooled human urine, injected before and after the patient samples. Lower limits of quantification (>80% accuracy) were 0.007, 0.34, 0.25, and 0.12 mg/mL for iPF(2α)-III, 2,3-dinor-iPF(2α)-III, iPF(2α)-VI, and 8,12-iso-iPF(2α)-VI, respectively.

Urinary allantoin
Allantoin was quantified using ultra performance LC-MS/MS (UPLC-MS/MS) with an Acquity UPLC system and TQD triple quadrupole mass spectrometer equipped with an electrospray ionization source (Waters Corp.). Allantoin (Sigma-Aldrich) and DL-allantoin-5-13C;1-15N (C/D/N Isotopes) were used as the standard and internal standard, respectively. Synthetic urine matrix (14) was used for preparation of samples and calibrants. Sample preparation included initial vortex mixing and centrifugation (15,000 × g for 10 min), addition of internal standard (25 μL urine + 25 μL internal standard, 100 μmol/L) and matrix (450 μL), and a second round of vortex mixing and centrifugation. In addition, 5 μL of sample were injected onto an Acquity UPLC BEH HILIC, 1.7 μm, 2.1× 100 mm column (Waters) heated to 40°C. Chromatographic separation was achieved by isocratic elution using 0.5% formic acid in acetonitrile/DI-H2O (95:5, v/v) as the mobile phase, with a flow rate of 200 μL/min. Allantoin and the internal standard were detected in positive ion mode using the following multiple reaction monitoring transitions (m/z): 159/116 and 159/61 as primary and secondary, respectively, for allantoin and 161/118 and 161/61 for the internal standard. The primary transitions were used to quantify the allantoin, and the secondary transitions were used as qualifier ions. Allantoin calibrators covered concentrations from 1 to 500 μmol/L; lower limit of quantification (accuracy, >80%) was 0.06 pmol.

Other variables
Data on age, tumor stage, estrogen receptor (ER) and progesterone receptor (PR) status, height, and weight were obtained from medical records. Body mass index (BMI) was calculated using the following formula: weight (kg)/[height (m)]². Data on supplement intake were collected using a questionnaire completed by participants at the time of recruitment.

Statistical analysis
We compared change in the mean levels of oxidative indices at the three time points using general linear models that allowed us to control for the covariance structure. Because adjustment for age and BMI did not change our estimates, our final models included only the time effect. Correlations between the biomarkers...
Results

The study population included women from 18 to 63 years of age, with 60% in the perimenopausal age range (40-54 y); participants were almost equally distributed across the conventional obesity categories (Table 1). Most women were Caucasian and had been diagnosed with a stage I or II breast cancer tumor; six had aggressive tumors (ER/PR negative/negative; Table 1). Approximately one third of the participants reported taking antioxidants and/or vitamin and mineral complexes. None of the urinary biomarkers examined at baseline correlated with any of these patient characteristics.

The levels of all examined urinary biomarkers increased at 1 hour after the doxorubicin injection: allantoin by 22% (P = 0.06), iPF(2α)-III by 62% (P < 0.05), iPF(2α)-VI by 41% (P < 0.05), 8,12-iso-iPF(2α)-VI by 58% (P < 0.05), and 2,3-dinor-iPF(2α)-III by 52% (P < 0.05; Table 2). The 1-hour changes in F2-isoprostanes were statistically significant, whereas the 1-hour change in allantoin was sizable (22%) and borderline significant. None of the urinary biomarkers increased at 1 hour (T1). At 24 hours (T24) after doxorubicin injection, the F2-isoprostanes returned to their baseline levels, whereas allantoin levels continued to increase (Table 2). The statistically significant 1-hour increases in F2-isoprostanes were greater than the increase in allantoin; nonetheless, the change in allantoin was sizable (22%) and borderline significant (P = 0.06). Although allantoin may be a less sensitive marker, this result points to its potential importance in diversifying the panel of oxidative indices.

Our study showed that all five urinary measurements are sensitive markers of oxidative stress in humans. Our data on urinary F2-isoprostanes in humans are consistent with the results of the BOSS study using CCl4 as an oxidative stressor in animals (6, 7). Moreover, our results on allantoin are consistent with a study that tested the effect of 2-hour exposure to 0.2 ppm ozone in healthy volunteers (n = 15; ref. 10). The ozone study found increased allantoin levels in nasal lavage at 1 hour into the actual exposure and immediately after the 2 hours of exposure compared with the pre-exposure levels.

The timeline of biomarker changes in our study and the ozone study is noteworthy (10). Both studies, carried out with humans rather than in animal models, showed an increase in oxidative indices at 1 hour after the exposure for both systemic (doxorubicin injection) and local (ozone exposure) oxidative stressors. In the ozone study, allantoin levels in nasal lavage had returned to pre-exposure levels at 6 hours after exposure (10). In our study, the urinary F2-isoprostanes returned to baseline levels by 24 hours after doxorubicin injection. Although the timeline of our post-exposure measurements did not allow us to ascertain whether the decrease in these biomarkers occurred earlier in the 24-hour interval, a consideration of doxorubicin pharmacokinetics suggests that this is unlikely. The peak of doxorubicin in plasma was expected and was seen in our study at 20 to 30 minutes after injection; however, the half-life of doxorubicin and its metabolites in the human body has been estimated to be 17 to 28 hours (15). The long half-life of doxorubicin suggests that oxidative stress should persist for 24 hours. In fact, dead cells in urine were seen in 24-hour samples practically invariably but not in 1-hour urine samples.

Our study raises questions as to why the half-life of doxorubicin and its metabolites and the cell-killing doxorubicin effect do not correspond to the dynamics of the biomarkers with its sharp increase at 1 hour after injection. One plausible explanation is that the antioxidant defense response of the body is quick, and biomarker levels in

<table>
<thead>
<tr>
<th>Table 1. Study subjects</th>
</tr>
</thead>
<tbody>
<tr>
<td>Characteristic</td>
</tr>
<tr>
<td>Age (y)</td>
</tr>
<tr>
<td>18-39 (premenopausal)</td>
</tr>
<tr>
<td>40-54 (premenopausal)</td>
</tr>
<tr>
<td>55-63 (menopausal)</td>
</tr>
<tr>
<td>Race</td>
</tr>
<tr>
<td>African-American</td>
</tr>
<tr>
<td>Caucasian</td>
</tr>
<tr>
<td>BMI</td>
</tr>
<tr>
<td>&lt;25</td>
</tr>
<tr>
<td>25-29.9</td>
</tr>
<tr>
<td>≥30</td>
</tr>
<tr>
<td>Tumor stage</td>
</tr>
<tr>
<td>I</td>
</tr>
<tr>
<td>II</td>
</tr>
<tr>
<td>III</td>
</tr>
<tr>
<td>ER/PR status</td>
</tr>
<tr>
<td>Negative/negative</td>
</tr>
<tr>
<td>Negative/positive</td>
</tr>
<tr>
<td>Positive/negative</td>
</tr>
<tr>
<td>Positive/positive</td>
</tr>
<tr>
<td>Use antioxidants and/or vitamin and mineral complexes</td>
</tr>
</tbody>
</table>

*The oldest patient in this study was 63 y old.
bodily fluids increase and decrease rapidly. Compared with this whole-body antioxidant defense reaction, damaged cells are slaughtered slowly due to the more drawn-out process of cytotoxic effects. Obtaining direct evidence to support or disprove this assumption would require timed experiments either with cell cultures or using animal models. It should also be noted that many studies have used plasma levels and urinary levels of F2-isoprostanes interchangeably as biomarkers of oxidative status despite evidence that they may differ. In short, it is possible that the increase and subsequent decrease in F2-isoprostane plasma levels may occur earlier compared with what we observed measuring urinary excretion of F2-isoprostanes (16).

In conclusion, we validated four biomarkers of oxidative status in a clinical model of oxidative stress. Urinary F2-isoprostanes, specifically iPF(2α)-III and its metabolite 2,3-dinor-iPF(2α)-III as well as iPF(2α)-VI and 8,12-iso-iPF(2α)-VI, are sensitive indices of systemic oxidative stress in humans. In addition, our results indicate that allantoin is a promising biomarker. Although its response to systemic oxidative stress is not as pronounced as those observed in F2-isoprostanes, this biomarker can serve an important role in diversifying the panel of oxidative indices available for the assessment of oxidative status in humans.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

Acknowledgments

We thank Dr. Kirk Kitchin (U.S. Environmental Protection Agency) for his critical review of this work.

Grant Support

Duke Comprehensive Cancer Center pilot study award, NIH Specialized Program of Research Excellence grant 5P50 CA68438, and Anna Merills’ Fund for Down Syndrome Research Foundation.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received 02/25/2010; revised 03/23/2010; accepted 03/25/2010; published OnlineFirst 05/25/2010.

References


Table 2. Oxidative indices before (T0) and 1 h (T1) and 24 h (T24) after administration of treatment

<table>
<thead>
<tr>
<th>Measurement</th>
<th>Mean (SD), n</th>
<th>T0</th>
<th>T1</th>
<th>T24</th>
</tr>
</thead>
<tbody>
<tr>
<td>Allantoin (mmol/mol creatinine)</td>
<td>7.2 (3.7)</td>
<td>20</td>
<td>19</td>
<td>19</td>
</tr>
<tr>
<td>T1 minus T0</td>
<td></td>
<td>1.6</td>
<td></td>
<td></td>
</tr>
<tr>
<td>T1 minus T24</td>
<td></td>
<td></td>
<td>2.1</td>
<td>0.3</td>
</tr>
<tr>
<td>iPF(2α)-III (ng/mg creatinine)</td>
<td>0.26 (0.15)</td>
<td>19</td>
<td>18</td>
<td>18</td>
</tr>
<tr>
<td>T1 minus T0</td>
<td></td>
<td>0.16</td>
<td></td>
<td>0.0004</td>
</tr>
<tr>
<td>T1 minus T24</td>
<td></td>
<td></td>
<td>0.07</td>
<td>0.2</td>
</tr>
<tr>
<td>2,3-dinor-iPF(2α)-III (ng/mg creatinine)</td>
<td>12.0 (7.8)</td>
<td>19</td>
<td>18</td>
<td>18</td>
</tr>
<tr>
<td>T1 minus T0</td>
<td></td>
<td>6.0</td>
<td></td>
<td>0.003</td>
</tr>
<tr>
<td>T1 minus T24</td>
<td></td>
<td></td>
<td>1.2</td>
<td>0.6</td>
</tr>
<tr>
<td>iPF(2α)-VI (ng/mg creatinine)</td>
<td>3.4 (2.0)</td>
<td>19</td>
<td>18</td>
<td>18</td>
</tr>
<tr>
<td>T1 minus T0</td>
<td></td>
<td>1.3</td>
<td></td>
<td>0.001</td>
</tr>
<tr>
<td>T1 minus T24</td>
<td></td>
<td></td>
<td>−0.3</td>
<td>0.6</td>
</tr>
<tr>
<td>8,12-iso-iPF(2α)-VI (ng/mg creatinine)</td>
<td>8.5 (4.6)</td>
<td>19</td>
<td>18</td>
<td>18</td>
</tr>
<tr>
<td>T1 minus T0</td>
<td></td>
<td>4.7</td>
<td></td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>T1 minus T24</td>
<td></td>
<td></td>
<td>−1.1</td>
<td>0.4</td>
</tr>
</tbody>
</table>


Urinary Biomarkers of Oxidative Status in a Clinical Model of Oxidative Assault

Dora Il'yasova, Ivan Spasojevic, Frances Wang, et al.

Cancer Epidemiol Biomarkers Prev  Published OnlineFirst May 25, 2010.

Updated version  Access the most recent version of this article at: doi:10.1158/1055-9965.EPI-10-0211

E-mail alerts  Sign up to receive free email-alerts related to this article or journal.
Reprints and Subscriptions  To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.
Permissions  To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.