The Role of Human Papillomavirus Genotyping in Cervical Cancer Screening: A Large-Scale Evaluation of the cobas HPV Test

Mark Schiffman1, Sean Boyle2, Tina Raine-Bennett3, Hormuzd A. Katki1, Julia C. Gage1, Nicolas Wentzensen1, Janet R. Kornegay2, Raymond Apple2, Carrie Aldrich2, Henry A. Erlich2, Thanh Tam2, Brian Befano4, Robert D. Burk5, and Philip E. Castle6,7

Abstract

Background: The cobas HPV Test (‘cobas’; Roche Molecular Systems) detects HPV16 and HPV18 individually, and a pool of 12 other high-risk (HR) HPV types. The test is approved for (i) atypical squamous cells of undetermined significance (ASC-US) triage to determine need for colposcopy, (ii) combined screening with cytology (“cotesting”), and (iii) primary HPV screening.

Methods: To assess the possible value of HPV16/18 typing, >17,000 specimens from a longitudinal cohort study of initially HPV-negative women (HC2, Qiagen) were retested with cobas. To study accuracy, cobas genotyping results were compared with those of an established method, the Linear Array HPV Genotyping Test (LA, Roche Molecular Systems). Clinical value of the typing strategy was evaluated by linking the cobas results (supplemented by other available typing results) to 3-year cumulative risks of CIN3+.

Results: Grouped hierarchically (HPV16, else HPV18, else other HR types, else negative), the \( \kappa \) statistic for agreement between cobas and LA was 0.86 [95% confidence interval (CI), 0.86–0.87]. In all three scenarios, HPV16-positive women were at much higher 3-year risk of CIN3+ than HPV16-negative women: women ages 21 and older with ASC-US (14.5%; 95% CI, 13.5%–15.5% vs. 3.5%; 95% CI, 3.3–3.6); women ages 30 years and older that were HPV-positive cytology-negative (10.3%; 95% CI, 9.6–11.1 vs. 2.3%; 95% CI, 2.2–2.4); and all women 25 years and older that were HPV-positive (18.5%; 95% CI, 17.8–19.2 vs. 4.3%; 95% CI, 4.2–4.4).

Conclusion: The cobas and LA results show excellent agreement. The data support HPV16 typing.

Impact: HPV16 typing is useful in the management of HPV-positive/cytology-negative women in cotesting, of all HPV-positive women in primary HPV testing, and perhaps in the management of HPV-positive women with ASC-US. Cancer Epidemiol Biomarkers Prev; 24(9); 1304–10. ©2015 AACR.

See related commentary by Del Mistro, p. 1302

Introduction

Human papillomavirus (HPV) testing is an increasingly important part of cervical screening (1). Effective implementation of HPV testing requires the use of thoroughly validated assays, and restricts their use to clinical indications supported by data (2, 3).

The Roche cobas HPV Test (Roche Molecular Systems) is one of 4 HPV tests currently approved by the FDA (4). Its indications include: (i) to triage patients 21 years and older with ASC-US (atypical squamous cells of undetermined significance) cervical cytology results to determine the need for referral to colposcopy, (ii) to test women 30 years and older, adjunctively with cytology, in the context of general screening (“cotesting”) and (iii) for primary HPV screening alone (without cytology) among women 25 years and older.

The cobas assay targets DNA of 14 HPV types. It simultaneously provides HPV type-specific results for HPV16 and HPV18, the two most important high-risk (HR) genotypes as well as a pooled result for 12 other HR types. The 12 other HR group includes the 10 other established carcinogenic types (HPV31, HPV33, HPV35, HPV39, HPV45, HPV52, HPV56, HPV58, and HPV59), a probably carcinogenic HPV type (HPV68) and a possibly carcinogenic type (HPV66; ref. 5).

The clinical value of HPV16 and HPV18 genotyping in each of three approved test settings is not clearly established. Current U.S. consensus guidelines do not recommend use of HPV16 and HPV18 typing among women with HPV-positive ASC-US, who are uniformly referred to colposcopy instead (6, 7). Guidelines do recommend use of HPV16/18 genotyping for women with cotesting results of HPV-positive, cytology-negative; those with HPV16/18 are immediately referred to colposcopy while those with other types are retested at 1 year (6, 7). Formal guidelines for the use of genotyping as part of primary HPV testing are not yet established; interim guidance (8) recommends colposcopy for HPV16/18-positive women, with cytology reserved for women positive for one or more infections with the 12 other HR HPV types.
By performing a large-scale study in a cohort of clinical specimens, we assessed the accuracy of HPV16/18 genotyping by cobas and the clinical utility of its typing strategy in the three possible clinical scenarios. To assess typing accuracy, we compared the genotype-specific clinical performance ofcobas to that of an established research use-only HPV typing method, Linear Array HPV Genotyping Test (LA, Roche Molecular Systems). To assess utility, we examined the risk stratification provided by type-specific HPV16 and HPV18 results for women ages 21 and older with ASC-US; HPV-positive, cytology-negative women ages 30 years and above, and all HPV-positive women ages 25 and above. Specifically, we assessed 3-year risk of CIN3+ as the measure of risk; using 3-year risk rather than cross-sectional risk helped to maximize ascertainment of outcomes regardless of immediate management.

Materials and Methods

Study population

To achieve the study objectives, *a posteriori* retesting of a large population of known HPV-positive women was performed. To assess typing accuracy, we analyzed paired cobas and LA test data on more than 17,000 specimens from a cohort study of initially Hybrid Capture 2 (HC2)-positive (Qiagen) women being conducted collaboratively by the NCI (Bethesda, MD) and Kaiser Permanente Northern California (KPNC, Oakland, CA; refs. 9, 10).

The NCI-KPNC cohort study from which the specimens were drawn is called the Persistence and Progression (PaP) Cohort (9). The PaP cohort is designed to explore viral and host determinants of cervical precancer among HPV-positive women; as the major outcome, we assessed 3-year risk of CIN3+ by typing result.

For the comparison of cobas and LA, a very large convenience sample of the initially HC-positive cohort was retested, namely all available paired cobas/LA test results from nested (completed and ongoing) case–control studies of prevalent and incident precursor or cancer. Women in the cohort can contribute many specimens in longitudinal fashion, as they return for follow-up. Thus, the tested specimens included both HC2-positive enrollment specimens and subsequently collected follow-up specimens from the same women (both HC2-positive and HC2-negative). In contrast, for the assessment of clinical utility of HPV typing among HPV-positive women, data from case–control analyses were used among HC2-positive specimens only from enrollment without consideration of longitudinal results from the same women (see details on Definitions of Cases and Controls below).

The PaP cohort was nested within routine practice at KPNC, where HPV testing using HC2 has long been used to triage ASC-US cytology. KPNC introduced cytology cotesting in 2003 for cervical screening at 3-year intervals for women 30 years and older (11). Pap tests are interpreted at KPNC regional and local laboratories; HPV testing of a co-collected second cervical specimen, collected into specimen transport medium (STM; Qiagen), from a randomly selected majority of women who tested HC2-positive. From 2007 to 2011, 45,302 HC2-positive women were enrolled. An additional small percentage (~7%) of women opted-out in response to a letter informing them of the PaP study. NCI and KPNC institutional review boards have approved the study yearly.

The current study within the PaP cohort is based on Roche Molecular Systems research testing of 17,262 selected archived specimens from the PaP cohort. Clinical follow-up cytology and histology were obtained on the cohort as described elsewhere (13). NCI selected the samples for Roche testing, which was performed masked to all other data. LA and cobas results were generated in tandem on the same aliquot of specimen.

As detailed below, to evaluate the performance of the cobas typing strategy for clinical risk stratification, we estimated cumulative 3-year risk of CIN3+ by typing result. However, a group of PaP cohort specimens from enrollment (those from the prevalent cases and controls who at that time had not developed CIN2+) had been tested only in another laboratory (RD Burk, Albert Einstein College of Medicine, The Bronx, NY), using another (MY09/11) L1 PCR-based method (14). To generate cumulative 3-year risk estimates, we included those results as well, as described below. Of note, the two assays varied in several details, including specimen input (cobas used more than MY09/11), extraction efficiency, and PCR efficiency. Nonetheless, this pooling of typing data was justified by good comparability of the additional data to cobas typing results (HPV16, else HPV18, else HR, else negative) as indicated by a κ of 0.65 [95% confidence interval (CI), 0.57–0.73] for 233 specimens tested by both assays.

HPV testing

Laboratory methods have been described fully elsewhere (13). In brief, denatured STM specimens were neutralized within 14 hours to minimize DNA damage.

For the cobas HPV testing, the x480 sample preparation module was used to prepare and aliquot the master mix and to perform sample addition. Twenty-five microliters of sample was added to 25 μl of master mix in a 96-well PCR plate. This plate was then manually sealed and transferred to the x480 real-time amplification and detection module of the cobas 4800 system as per the manufacturer’s protocol, using spectrally unique fluorescent dyes to label TaqMan probes for HPV16, HPV18, and the 12 other HR-HPV genotypes.

The LA assay targets the 14 HR types included in the cobas assay and also HPV6, 11, 26, 40, 42, 53, 54, 55, 61, 62, 64, 67, 69, 70, 71, 72, 73 (MM9), 81, 82 (MM4), 83 (MM7), 84 (MM8), IS939 (MM subtype), and 89 (CP6108). In brief, automated sample extraction was performed on the neutralized STM sample using the x480 sample extraction module of the cobas 4800 system. The HPV LA test was carried out according to the manufacturer’s protocol available within the package insert of the kit with minor modifications (13).

For the subset of specimens from prevalent cases and controls, whose results were needed to complete 3-year risk estimates, genotyping results generated with another PCR-based method
in the Burk laboratory were included. The MY09/M11 L1 degenerate primer PCR (MY09/11 PCR) system used by the Burk laboratory to test prevalent cases and controls has been described previously (14).

Definitions of cases and controls for risk estimation

In the comparison of cobas with LA, all specimens with paired results were considered, disregarding case–control status. For analyses assessing clinical utility, 3-year cumulative risk of CIN3– or CIN2– were calculated on the basis of results from enrollment specimens. Cases diagnosed before a repeat screening-type visit (i.e., a second cotest) were classified as prevalent, while those cases diagnosed after a second screening visit were considered incident; admittedly, this division is arbitrary. The analyses focused on CIN3– as the main case group and surrogate of cancer risk; but also considered CIN2–, despite lack of diagnostic reproducibility (15), because CIN2 lesions are commonly treated. The CIN3– or CIN2– cases were compared with a random sample of HC2–positive control women that had not been diagnosed with CIN2– at the time of selection, and had returned at least for one subsequent screening at approximately 1 year after enrollment (10). As shown below, this analysis was conducted separately for all selected women 21 years and older with ASC-US to address triage, all selected women ages 30 years and above that were HC2– and cytology negative to address cotesting, and all selected women ages 25 years and older that were HC2– to address primary HPV screening. The testing fractions are shown in Table 1. As shown in the table, the supplementary testing by MY09/11 PCR applied to prevalent cases and controls diagnosed during the enrollment period. However, some prevalent cases developed more severe diagnoses during follow-up (e.g., CIN2 was followed by CIN3), and some prevalent controls developed CIN2+. For analysis, these individuals were categorized as incident cases instead according to their worst diagnosis.

Statistical analyses

To assess typing accuracy, the LA and cobas assays were compared at the level of HPV16, HPV18, 12 other HR types, or negative for the 14 targeted types, using the κ statistic and asymptotic χ². This analysis was hierarchical, using “else if” logic. Thus, any specimen with HPV16 detected was grouped as HPV16, only specimens without HPV16 could be grouped as HPV18, and only specimens negative for both HPV16 and HPV18 could be grouped as 12 HR. (This analysis was confirmed with nonhierarchical analyses, in which a specimen concurrently could have HPV16, and/or HPV18, and/or other HR types, but we do not present those very similar results here). The comparison was repeated, stratified by correlates of HPV viral load, that is, concurrent HC2 result (positive/negative), cytologic result (negative; ASC-US; LSIL; and high-grade, including AGC, ASC-H, HSIL, AIS, and cancer), and also by case status (worst histopathologic result).

Results were also examined at the HPV type–specific level, with type determined by LA. For clarity, this analysis was restricted to specimens that were positive for only a single one of the 14 cobastargeted types (whether or not other, e.g., low-risk types, were also present). To address a limitation of LA (16), HPV52 was called positive only when no other type in the LA pooled probe used to define HPV52 was individually positive. As there was no attempt to adjudicate absolute truth, discrepancies between the 2 assays could be described only in relative terms (e.g., in the case of additional positives by LA, it was not possible to distinguish between true and false positives). Moreover, the interassay comparison was conducted at the level of specimens; some women contributed more than 1 specimen (enrollment plus 1 or more follow-up specimens) to the analysis. We did not adjust for the remote possibility of statistical autocorrelation.

The second major part of the statistical analysis was to determine whether detecting the presence of HPV16 or HPV18 at enrollment, as provided by cobas, would meaningfully alter the risk of CIN3– (or secondarily, CIN2+) in a clinically actionable manner; that is, changing management from colposcopy to 1-year follow-up in the case of HPV-positive ASC-US or, conversely, changing management to immediate colposcopy for cotesting or primary HPV screening. Because, in the course of the nested case–control studies, only a random sample of 10% to 15% of the noncases (and most of the cases) were tested by cobas, with the supplementation of Burk lab PCR, it was necessary to use the sampling fractions to weight up estimates from the tested group to properly represent the full PaF cohort (Table 1). The tested group was multiplied by the inverse of the sampling fractions to estimate

### Table 1. Sampling and testing methods for three subcohorts for which 3-year cumulative risks of CIN3– and CIN2– were estimated; HC2– enrollment specimens were tested for HPV16, else HPV18, else 12 other high-risk types

<table>
<thead>
<tr>
<th>Case–control status</th>
<th>Total N in subcohort</th>
<th>Tested by cobas</th>
<th>Tested by MY09/MY11 PCR or cobas</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>N</td>
<td>% of Total</td>
<td>N</td>
</tr>
</tbody>
</table>

A. ASC-US Triage: Sample size of women with HC2+: ASC-US, ages 21 years and older (median 33), tested by cobas or MY09/MY11 PCR

- Control 11,966
- Prevalent CIN2+ 1,300
- Incident CIN2+ 624

<table>
<thead>
<tr>
<th></th>
<th>N</th>
<th>% of Total</th>
<th>N</th>
<th>% of Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>1,244</td>
<td>10.4%</td>
<td>1,670</td>
<td>14.0%</td>
</tr>
<tr>
<td>Prevalent CIN2+</td>
<td>528</td>
<td>40.6%</td>
<td>1,171</td>
<td>90.1%</td>
</tr>
<tr>
<td>Incident CIN2+</td>
<td>426</td>
<td>68.3%</td>
<td>450</td>
<td>72.1%</td>
</tr>
</tbody>
</table>

B. Cotesting: sample size of women with HC2+/negative cytology, ages 30 years or older (median 40), tested by cobas or MY09/MY11 PCR

- Control 17,425
- Prevalent CIN2+ 204
- Incident CIN2+ 1,176

<table>
<thead>
<tr>
<th></th>
<th>N</th>
<th>% of Total</th>
<th>N</th>
<th>% of Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>2,773</td>
<td>15.9%</td>
<td>3,490</td>
<td>20.0%</td>
</tr>
<tr>
<td>Prevalent CIN2+</td>
<td>25</td>
<td>12.3%</td>
<td>194</td>
<td>95.1%</td>
</tr>
<tr>
<td>Incident CIN2+</td>
<td>897</td>
<td>76.3%</td>
<td>970</td>
<td>82.5%</td>
</tr>
</tbody>
</table>

C. Primary HPV testing: sample size of women with HC2+, ages 25 years or older (median 38), tested by cobas or MY09/MY11 PCR

- Control 35,134
- Prevalent CIN2+ 3,290
- Incident CIN2+ 2,315

<table>
<thead>
<tr>
<th></th>
<th>N</th>
<th>% of Total</th>
<th>N</th>
<th>% of Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>4,708</td>
<td>13.4%</td>
<td>6,047</td>
<td>17.2%</td>
</tr>
<tr>
<td>Prevalent CIN2+</td>
<td>761</td>
<td>23.1%</td>
<td>2,989</td>
<td>90.9%</td>
</tr>
<tr>
<td>Incident CIN2+</td>
<td>1,683</td>
<td>72.7%</td>
<td>1,827</td>
<td>79.9%</td>
</tr>
</tbody>
</table>

NOTE: A control was Hybrid Capture 2 (HC2) positive at enrollment but, during observation, never developed CIN2+. A prevalent case of CIN2+ (or CIN3+) was diagnosed after enrollment (HC2+) screening, prior to a second screen. An incident case of CIN2+ (or CIN3+) was HC2+ at enrollment, but diagnosed after at least the second screen. The table demonstrates that cobas data were supplemented by MY09/MY11 PCR data mainly for prevalent cases of CIN2+. Combining both testing methods, a high percentage of cases of CIN2+ were typed, permitting reasonably precise estimation of cumulative risk. A smaller fraction of HC2+ controls was tested.

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the distributions for the whole population, and the cumulative 3-year risks and 95% CIs were calculated using weighted Kaplan–Meier methods (using SAS-callable SUDAAN version 11.0.1) to adjust for censoring and incomplete follow-up. The consideration of HPV types was hierarchical. To determine this order, an iterative approach was used in which each of the preceding higher risk channels was excluded from consideration for the analyses of the remaining HPV channels. We looked at each HPV channel individually and chose the one with the highest positive predictive value for the 3-year risk of CIN3+, that is, HPV16 (see Results).

We then sought to determine, given an HPV16-negative status, whether testing positive for any other type would indicate the need for colposcopic referral. Consequently, we excluded all women testing positive for HPV16 and repeated the risk calculations among the remaining women to determine the next highest risk channel.

Results
Agreement between cobas and LA
Overall, 17,262 specimens from the PaP study were tested by both cobas and LA. The specimens were collected from 8,451 women, whose median age was 37 at enrollment (range, 21–87; interquartile range, 30–47). There was a close concordance between cobas and grouped LA results. As shown in Table 2, when the results were grouped hierarchically (HPV16, else HPV18, else other HR types, else negative), the x² statistic for agreement between the assays was 0.86 (95% CI, 0.86–0.87), indicating excellent agreement. As interassay was not perfect, the discrepancies were further examined. There was a steady trend for agreement to be higher in subgroups of the cohort with presumed higher HPV viral load (i.e., when HC2 was positive and/or cytology was abnormal). Thus, the lowest but still good agreement (x² = 0.65; 95% CI, 0.62–0.69) was observed among specimens that were concurrently HC2-negative and cytology-negative; at the other extreme, agreement among HC2-positive cases with CIN3+ diagnoses was very high (x² = 0.90; 95% CI, 0.88–0.92). The cobas assay tended to be slightly less specific than HPV16-positive than cobas (x² for agreement on HPV16 > 0.92, 20.8% vs. 19.3%, respectively; P < 0.001 by McNemar test). Most HPV16 positives, LA-negative specimens were classified by LA as having other HR HPV types; no particular type predominated (data not shown). The HPV16 cobas-positive, LA-negative specimens (n = 344) were as likely as HPV16 LA-positive, cobas-negative specimens (n = 91) to come from women with CIN3+ (~15% in both groups), but either set of discrepant results was less likely to derive from women with CIN3+ than concordantly positive results (~30% of 3,229, data not shown).

Further type-specific analyses of the positivity of cobas relative to LA are shown in Table 3; in this analysis, we were particularly interested in exploring LA-positive specimens that were negative by cobas. LA results were used to classify single-type infections for each of the 14 types targeted by cobas. Multiple concurrent infections with one of the 14 cobas-targeted types were excluded for the comparison shown in Table 3. Each type by LA was compared with the corresponding cobas result (HPV16, HPV18, or each of the other 12 HR types). The results indicate that, with LA as the reference-standard typing assay, cobas detected a very high (>95%) percentage of all single-type infections for almost all targeted types. As the exceptions, the cobas HR was negative for 19.9% of 1,223 specimens that LA classified as single HPV52, 16.2% of 648 specimens that LA classified as single HPV58, and 9.8% of 594 specimens that LA classified as single HPV51. When multiple infections were included rather than excluded from the analysis, cobas detection of LA-positive HPV16 or HPV18 slightly declined (data not shown).

To explore the fraction of specimens that tested positive by cobas but negative by LA for the 14 types targeted by both assays, we examined whether cobas cross-reacted with additional (e.g., low-risk) types detected by LA. There was no evidence of cross-reactivity for this group. In fact, the cobas assay was less likely to be positive (for at least one HPV channel) when LA detected only 1 or more types not targeted by cobas (9.8% cobas-positive).

Table 2. Comparison of hierarchical HPV results from cobas and LA, grouped as HPV16 else HPV18 else 12 high-risk (HR) HPV, else negative for 14 targeted types

<table>
<thead>
<tr>
<th>LA result</th>
<th>Cobas HPV16, n</th>
<th>Cobas HPV18, n</th>
<th>Cobas high-risk HPV, n</th>
<th>Cobas HPV-negative, n</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>LA HPV16</td>
<td>3,229</td>
<td>5</td>
<td>46</td>
<td>40</td>
<td>3,320</td>
</tr>
<tr>
<td>LA HPV18</td>
<td>40</td>
<td>904</td>
<td>21</td>
<td>7</td>
<td>972</td>
</tr>
<tr>
<td>LA high-risk HPV</td>
<td>7,767</td>
<td>537</td>
<td>446</td>
<td>4,312</td>
<td>8,591</td>
</tr>
<tr>
<td>LA HPV−</td>
<td>263</td>
<td>1,011</td>
<td>8,280</td>
<td>4,307</td>
<td>17,176</td>
</tr>
<tr>
<td>Total</td>
<td>3,573</td>
<td>1,071</td>
<td>8,767</td>
<td>4,393</td>
<td>17,717</td>
</tr>
</tbody>
</table>

NOTE: x² statistic = 0.86 (95% CI, 0.86–0.87, indicating excellent overall agreement), test of symmetry P < 0.0001 (indicating relatively increased HPV16 and HPV18 positivity of cobas relative to LA). Of note, all specimens with both LA and cobas paired results were tested, including enrollment specimens (all HC2-positive) and subsequent longitudinal specimens from the same women (many of which were HC2-negative). Frequency missing = 86.

Bolded values represent inter-assay agreement, while off-diagonal, unbolded values represent lack of agreement.

Table 3. Individual type positivity by cobas, restricted to samples positive by LA for only one of 14 cobas-targeted high-risk types

<table>
<thead>
<tr>
<th>LA single HPV type</th>
<th>Total number LA single-type positive</th>
<th>% cobas positive on corresponding channel</th>
</tr>
</thead>
<tbody>
<tr>
<td>HPV16</td>
<td>2,143</td>
<td>98.4%*</td>
</tr>
<tr>
<td>HPV18</td>
<td>605</td>
<td>98.5%*</td>
</tr>
<tr>
<td>HPV31</td>
<td>1,108</td>
<td>98.9%</td>
</tr>
<tr>
<td>HPV33</td>
<td>258</td>
<td>100.0%</td>
</tr>
<tr>
<td>HPV35</td>
<td>407</td>
<td>96.8%</td>
</tr>
<tr>
<td>HPV39</td>
<td>626</td>
<td>96.7%</td>
</tr>
<tr>
<td>HPV45</td>
<td>450</td>
<td>97.8%</td>
</tr>
<tr>
<td>HPV51</td>
<td>594</td>
<td>90.2%</td>
</tr>
<tr>
<td>HPV52</td>
<td>1,223</td>
<td>80.1%</td>
</tr>
<tr>
<td>HPV56</td>
<td>516</td>
<td>99.0%</td>
</tr>
<tr>
<td>HPV58</td>
<td>648</td>
<td>83.8%</td>
</tr>
<tr>
<td>HPV59</td>
<td>506</td>
<td>95.3%</td>
</tr>
<tr>
<td>HPV66</td>
<td>391</td>
<td>98.0%</td>
</tr>
<tr>
<td>HPV68</td>
<td>292</td>
<td>96.6%</td>
</tr>
</tbody>
</table>

NOTE: Multiple infection with other, i.e., low-risk types, was not reason for exclusion. HPV16 positivity by LA was compared with the cobas HPV16 channel, HPV18 positivity by LA was compared with the cobas HPV18 channel, and 12 other high-risk types by LA were compared with the cobas HR2 channel. *15% of 7622 LA negative for HPV16 (among this group with single-type infections) were cobas-positive for HPV16, and 0.6% of 9162 LA negative for HPV18 were cobas positive for HPV18.
ASC-US triage: HPV typing and risk of CIN3+, among women with positive HC2 results and ASC-US cytology

A possible use of cobas typing (HPV16 and HPV18) not recommended by current guidelines is among women with HPV-positive ASC-US, who are routinely referred to colposcopy. The question is whether women with ASC-US who are negative for HPV16 and HPV18 might be at sufficiently low risk not to need immediate colposcopy; if so, perhaps they could be asked to return in 1 year instead. Thus, 3-year cumulative risk of CIN3+ was estimated by HPV16 and HPV18 typing results among women with HPV-positive ASC-US.

As shown in Table 4, overall, women 21 years and older with ASC-US who tested HC2+ had a 5.2% 3-year risk of CIN3+ overall, but those negative for HPV16 were at much lower risk of CIN3+ (3.5%, 95% CI, 3.3%–3.6%) than HPV16-positive women (14.5%, 95% CI, 13.5–15.5%). Thus, testing negative for HPV16 yielded a risk that was lower than the current colposcopy threshold, which is benchmarked in guidelines to the risk (here, 5.2%) of all HPV-positive ASC-US (6, 7). Among HPV16-negative women, HPV18 or other HR detection only slightly stratified risk in this group of HC2-positive women with ASC-US. Repeating the analysis using CIN2+ as the disease definition yielded similar conclusions (Supplementary Table S1). An ancillary analysis restricting to cobas data (excluding the MY09/11 PCR data) resulted in falsely low cumulative risk estimates (because many prevalent cases were excluded) but the pattern of risk stratification was unchanged (data not shown). There was no meaningful change in the pattern of results when age was stratified into tertiles (data not shown).

Cotesting: HPV typing and risk of CIN3+ in cotesting, among women ages 30 years and above with positive HC2 results and negative cytology

To evaluate clinical use of the HPV16 and HPV18 typing in cotesting, specifically in the triage of HPV-positive/Pap-negative results, the analysis was restricted to HPV-positive (by HC2), cytology-negative enrollment specimens. Again, 3-year cumulative risk of CIN3+ was calculated by typing result. In the absence of HPV HPV16 and HPV18 typing results, guidelines recommend that such women return in 1 year (6, 7). The question was whether HPV16 and HPV18 typing by cobas would stratify risk in such a way that it might justify immediate colposcopy.

Table 4. Use of cobas-like partial HPV typing in triage of ASC-US

<table>
<thead>
<tr>
<th>HPV-type positive</th>
<th>3-Year risk of CIN3+</th>
<th>95% CI</th>
<th>3-Year risk of CIN3+</th>
<th>95% CI</th>
</tr>
</thead>
<tbody>
<tr>
<td>HPV16</td>
<td>14.5%</td>
<td>13.5%–15.5%</td>
<td>3.5%</td>
<td>3.3%–3.6%</td>
</tr>
<tr>
<td>HPV18</td>
<td>4.8%</td>
<td>4.1%–5.7%</td>
<td>3.3%</td>
<td>3.2%–3.5%</td>
</tr>
<tr>
<td>HPV HR</td>
<td>3.5%</td>
<td>3.3%–3.6%</td>
<td>2.8%</td>
<td>2.5%–3.2%</td>
</tr>
</tbody>
</table>

NOTE: The estimated CIN3+ risk among all women with ASC-US testing HC2+, prior to typing, was 5.2%; by current U.S. guidelines all would be referred to colposcopy. Three-year cumulative risk of CIN3 or worse (CIN3+) among women with Hybrid Capture 2-positive (HC2+) ASC-US, ages 21 years and older, stratified by HPV type status (HPV16, else HPV18, else other 12 high-risk (HR) types targeted by cobas).

As shown in Table 5, the 3-year cumulative risk of CIN3+ among women ages 30 years and older who tested HC2-positive but cytology-negative was 3.5% overall; as expected, this risk is lower than the colposcopy threshold according to current guidelines as applied to this group, which is set by the risk for LSIL (5.9%) or all HPV+ ASC-US (5.5%). HPV-positive, cytology-negative women that were positive for HPV16 were at much higher cumulative risk of CIN3+ (10.3%; 95% CI, 9.6–11.1), compared with HPV16-negative women (2.3%; 95% CI, 2.2–2.4). Thus, the magnitude of risk among HPV16-positive women exceeded the threshold of risk at which current guidelines recommend referral to colposcopy. Among HPV16-negative women, HPV18 detection only slightly stratified risk (Table 5). In the absence of either HPV16 or HPV18, positivity for the other HR types did not meaningfully change risk of CIN3+ in this group of HC2-positive women. Repeating the analysis using CIN2+ as the disease definition yielded similar conclusions (Supplementary Table S2), as did restricting to cobas data (data not shown).

Primary HPV testing: HPV typing and risk of CIN3+ in primary HPV testing, among women ages 25 years and older with positive HC2 results

To evaluate clinical use of the HPV16 and HPV18 typing in primary HPV testing, the analysis was restricted to HPV-positive (by HC2) specimens among women ages 25 and older, that is, the age group for which the FDA recently approved primary HPV testing by cobas. Again, 3-year cumulative risk of CIN3+ by typing result was calculated. The question was whether HPV16 and HPV18 typing would stratify risk in such a way that it would justify immediate colposcopy rather than repeat testing in 1 year. Of note, the 25- to 29-year-old group tested by HC2 in KPNC was dominated by ASC-US triage (approximately half had ASC-US); thus, this population had a higher intrinsic risk than a true population sample of all women ages 25 years and older.

As shown in Table 6, the 3-year cumulative risk of CIN3+ among women 25 years and older who tested HC2-positive was 6.7% overall. Formal guidelines do not yet exist for colposcopy referral following primary HPV testing (8); of note, the risks in this group were very similar when comparing all HC2-positive women, to those with LSIL (6.0%) and those with HPV+ ASC-US (5.5%). However, those positive for HPV16 were at much higher cumulative risk of CIN3+ (18.5%, 95% CI, 17.8–19.2), compared with HPV16-negative women (4.3%, 95% CI, 4.2–4.4). Among HPV16-negative women, HPV18 detection only slightly stratified risk (Table 6) in these HC2-positive women ages 25 years and older.
above. In the absence of either HPV16 or HPV18, positivity for the other HR types did not meaningfully change risk of CIN3+ in this group of HC2-positive women. Repeating the analysis using CIN2+ as the disease definition yielded similar conclusions (Supplementary Table S3), as did restricting to cobas results (data not shown).

Discussion

Agreement between different HPV assays is rarely if ever perfect (17). As expected from previous work (18), we observed that agreement between cobas and LA (at the level of HPV16, else HPV18, else 12 other HR types, else negative) was at least good in all subgroups, but tended to be stronger when viral load was higher (as indicated by HC2 positivity, abnormal cytology, or diagnosis of CIN3+ compared with controls). Also, cobas positivity for HPV16 or HPV18 was slightly higher when LA indicated the presence of a single HPV type rather than multiple HR infections. The cobas assay tended to yield HPV16-positive results slightly more often than LA, but the association with CIN3+ was equal for the two assays. Among the targeted, 12 other HR types, cobas sensitivity relative to LA was somewhat lowered for HPV52, HPV58, and HPV51. We used a stringent definition for HPV52 (calling it positive only when none of the other types in the mixed probe were present) to avoid false LA HPV52 positivity. No cancers were missed by cobas that were detected by HC2 and/ or LA but the sample of cancers was small. There was no indication of cobas cross-reactivity with non-HR types detected by LA.

The clinical performance demonstrated among women with HPV-positive enrollment results supports the recent interim clinical guideline (8) regarding primary HPV testing among women 25 and older, which recommends colposcopy for those with HPV16. The estimates of 3-year cumulative risk were quite similar to those generated in the ATHENA trial, which was presented to the FDA for approval of primary HPV testing. Specifically, for the 3 cobas channels, the KPNC versus ATHENA estimates were quite close, as follows: HPV16 (18.5% vs. 25.2%, respectively), else HPV18 (7.8% vs. 11.0%) else HR (4.3% vs. 5.4%; ref. 19).

Clinical performance among women with HPV-positive Pap-negative results among women ages 30 years and older supports recent consensus guidelines regarding cotesting, which recommend colposcopy for those with HPV16 (6, 7). In both the primary HPV testing and cotesting scenarios, the 3-year cumulative risk of CIN3+ (or CI for women with HPV16 was substantially greater than the benchmark risk thresholds for colposcopy, that is, higher than the risks associated with LSIL or HPV-positive ASC-US in the same populations. In contrast, in both cases (primary HPV testing or cotesting), the 3-year risk of CIN3+ (or CIN2+) was not (particularly elevated (for all HPV-positive women ages 25 years and older or those 30 years and above with negative cytology, respectively) among HPV18-positive women in the absence of HPV16. It is not surprising that our 3-year analyses based on CIN3+ or CIN2+ did not demonstrate increased risk linked to HPV18. The clinical concern of HPV18 infection is not based on short-term risk of precancer, rather it is based mainly on the link between HPV18 and hard-to-detect precancerous and invasive lesions (particularly glandular lesions) that are observed in prospective studies with longer follow-up periods (20, 21).

The 2012 guidelines do not recommend HPV typing of women with HPV-positive ASC-US; such women all are currently referred to colposcopy (7). However, our findings, in contrast to the scant data considered for the management guidelines, suggest that the absence of HPV16 might lower risk below the threshold for colposcopy. If confirmed and found to be cost-effective, this finding could lead to consideration of HPV typing for HPV16 (and HPV18, for the aforementioned separate reason) as part of ASC-US triage.

There are limitations to our large performance study of the cobas HPV test. The study was limited to a large convenience sample of longitudinal specimens from women that initially tested positive by HC2, using the leftover routine cotest specimen collected after cytology. An analysis in which cobas was used to determine HPV positivity might have yielded slightly different results, despite the fact that the two HPV tests have similar performance (17). Our cobas-LA comparison included only Roche testing; however, to permit 3-year cumulative risks necessitated including some MY09/11 PCR results from another laboratory, therefore, our risk stratification estimates are not based solely on cobas data. Restricting to cobas data led to the same conclusions, despite artificial lowering of the cumulative risk estimates (data not shown). Our HC2-negative specimens were derived from follow-up of women who were initially positive for HPV by the HC2 test, and cannot be generalized to the larger HPV-negative population. Also, both cobas and LA testing were performed out of a remaining aliquot of the HC2 test specimen, collected into the Qiagen STM buffer, denatured, and neutralized. This is not an FDA-approved process for cobas testing. These factors limit, albeit to an unknown extent, the generalizability of our findings to cobas performed in the FDA-approved manner. Finally, it is worth noting that this study was conducted at a large integrated health system in the United States, not in a population-based screening program like those in some other countries. The optimal screening and management strategies could differ when organized screening with wide coverage and high compliance with follow-up intervals are present. Specifically, reliance on longer periods of follow-up to differentiate benign HPV infections from those leading to precancer is more practical in an organized national program.

The strength of the study is its large size, which permitted a statistically powerful, detailed examination of cobas performance relative to LA typing and of typing relative to CIN3+ outcomes. In conclusion, the results demonstrate that cobas and LA results show excellent but not perfect agreement. The data support the current clinical guideline recommending HPV16 typing in the management of HPV-positive women, among HPV-positive/Pap-negative women, and suggest that knowing
HPV16 is absent might be useful in the management of HPV-positive women with ASC-US.

Disclosure of Potential Conflicts of Interest
M. Schiffman and J.C. Gage report receiving commercial research support from Roche and BD. P.E. Castle has received speakers bureau honoraria from Roche and Cepheid, is a consultant/advisory board member for ClearPath, Inovio, GE Healthcare, Cepheid, Roche, Gentecel, BD, Hologic, and Guided Therapeutics, and has provided expert testimony for Merck. No potential conflicts of interest were disclosed by the other authors.

Authors’ Contributions
Conception and design: M. Schiffman, H.A. Katki, N. Wentzensen, P.E. Castle
Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): M. Schiffman, S. Boyle, T. Raine-Bennett, J.R. Kornegay, C. Aldrich, P.E. Castle
Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): M. Schiffman, S. Boyle, H.A. Katki, J.C. Gage, N. Wentzensen, H.A. Ehrlich, T. Tam, B. Befano, P.E. Castle

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