Associations between Environmental Exposures and Incident Colorectal Cancer by ESR2 Protein Expression Level in a Population-Based Cohort of Older Women


Abstract

Background: Cigarette smoking (smoking), hormone therapy (MHT), and folate intake (folate) are each thought to influence colorectal cancer risk, but the underlying molecular mechanisms remain incompletely defined. Expression of estrogen receptor β (ESR2) has been associated with colorectal cancer stage and survival.

Methods: In this prospective cohort study, we examined smoking, MHT, and folate-associated colorectal cancer risks by ESR2 protein expression level among participants in the Iowa Women’s Health Study (IWHS). Self-reported exposure variables were assessed at baseline. Archived, paraffin-embedded colorectal cancer tissue specimens were collected and evaluated for ESR2 protein expression by IHC. Multivariate Cox regression models were fit to estimate relative risks (RR) and 95% confidence intervals (CI) for associations between smoking, MHT, or folate and ESR2-defined colorectal cancer subtypes.

Results: Informative environmental exposure and protein expression data were available for 491 incident colorectal cancer cases. Positive associations between ESR2-low and -high tumors and several smoking-related variables were noted, most prominently with average number of cigarettes per day (RR, 4.24; 95% CI, 1.81–9.91 for ESR2-low and RR, 2.15; 95% CI, 1.05–4.41 for ESR2-high for ≥40 cigarettes compared with nonsmokers). For MHT, a statistically significant association with ESR2-low tumors was observed with longer duration of exposure (RR, 0.54; 95% CI, 0.26–1.13 for >5 years compared with never use). No associations were found for folate.

Conclusions: In this study, smoking and MHT were associated with ESR2 expression patterns.

Impact: These data support possible heterogeneous effects from smoking and MHT on ERβ-related pathways of colorectal carcinogenesis in older women. Cancer Epidemiol Biomarkers Prev; 24(4): 713–9. ©2015 AACR.

Introduction

Colorectal cancer represents the third most common incident and fatal cancer in the United States (with estimates of 136,830 new cases and 50,310 attributable deaths in 2014; ref. 1). Cigarette smoking has been shown by us and others to increase the risk for colorectal cancer (2–4), whereas hormone therapy (MHT) has protective effects (5–8). Less clear is the role that folate intake has on colorectal cancer risk (9). Kim and colleagues found an increase in folate modestly decreased risk, although other studies have yielded mixed results (10, 11).

Molecular heterogeneity in colorectal carcinogenesis is well established (12–14). Concordantly, emerging data from our group and others demonstrate differential associations between common environmental exposures, including smoking, MHT and folate, and incident colorectal cancers defined by microsatellite instability (MSI), CpG island methylator phenotype (CIMP), KRAS and BRAF mutation status (2, 3, 15–18), and TP53 protein expression (19), among other phenotypic markers. Most significantly, postmenopausal MHT was associated with a lower risk for MSI-L/MSS tumors (15) and smoking was shown to be associated with MSI-high, CIMP-positive, and BRAF-mutated tumors (2).

To date, relatively few studies have examined subtype-specific colorectal cancer risks by ESR2 (ERβ) expression levels (20, 21). ESR2 (ERβ) is the main estrogen receptor (ER) expressed in colon tissue (22). Although the exact mechanism is yet to be determined, it appears that ESR2 signaling has a role in the protective effect of MHT against colon tumor development (23). ESR2 is
highly expressed in normal colonic mucosa but declines in colon adenocarcinoma. ESR2 loss in colon tissue is associated with progressing cancer and cell dedifferentiation (24, 25) as well as advanced cancer stage and poor survival (26). Both tobacco carcinogens and estrogen utilize some of the same enzymes for metabolites. Smoking induces the expression of genes that are involved in estrogen metabolism and, in lung tissue, has been shown to increase the carcinogenic estrogen metabolite 4-OHE. So it seems biologically plausible that their pathways may overlap and smoking may influence the estrogen pathway (27). Further clarification of the risk factors for molecularly defined colorectal cancer subtypes could inform more targeted prevention, early detection, and treatment strategies.

In this current study, we used baseline data and archived tumor tissue specimens from the prospective Iowa Women’s Health Study (IWHS) to examine exposures associated with ESR2-defined colorectal cancer subtypes in older women. Smoking, MHT, and folate were investigated as potentially modifiable lifestyle, medication, and dietary factors, respectively. On the basis of previous reports from our group and others (2, 3, 15, 16, 18, 19), these exposures may be plausibly linked to heterogeneous pathways of colorectal carcinogenesis.

Materials and Methods

This study was reviewed and approved by the Institutional Review Boards for Human Research of the University of Iowa (Iowa City, IA), University of Minnesota (Minneapolis, MN), and Mayo Clinic (Rochester, MN).

Subjects

Recruitment and enrollment methods for the IWHS have been reported elsewhere (28). Briefly, a 16-page baseline questionnaire was used to collect comprehensive self-reported demographic, dietary, lifestyle, and medication data from 41,836 Iowa women, ages 55 to 69 years, who held a valid driver’s license at baseline in 1986. Subjects were excluded for the present study based on the following factors (not mutually exclusive): history of any malignancy other than skin cancer (n = 3830); follow-up less than one day (n = 10); incomplete baseline exposure information (n = 660 for smoking and n = 200 for MHT); incomplete premenopausal or menopause status (for MHT analyses only, n = 569); or invalid dietary data (for folate analyses only, >30 missing dietary variables, self-reported intakes of <600 calories per day, or ≥5,000 calories per day, n = 3,096). Vital status and state of residence were determined by mailed follow-up surveys and through linkage to Iowa death-certificate records.

Risk factor assessment

Smoking patterns, including smoking status (never, ever, former, current), smoking duration (years), average number of cigarettes smoked per day, and cumulative pack-years were collected. Dietary habits were assessed using a semiquantitative food frequency questionnaire adapted from the 126-item instrument developed by Willett and colleagues (29). Folate was computed by multiplying the frequency response by the nutrient content of the specified portion sizes, with additional intake from supplement use included when indicated. Previous or current MHT and duration of MHT exposure were also collected, as described previously (15). Potential confounding variables acquired from the baseline questionnaire included body mass index (BMI), waist-to-hip ratio (WHR), physical activity level, alcohol consumption, age at menarche, age at menopause, oral contraceptive use, history of diabetes mellitus and daily intake of total calories, fat, sucrose, red meat, calcium, vitamin E, and methionine.

Case ascertainment

Incident colorectal cancer cases were identified through annual linkage with the Iowa Cancer Registry, which is a member of the National Cancer Institute’s Surveillance, Epidemiology and End Results program (30). Colorectal cancer cases were identified using International Classification for Diseases in Oncology (ICD-O) codes of 18.0, 18.2 to 18.9, 19.9, and 20.9, with tumors located in the cecum, ascending colon, hepatic flexure, transverse colon, and splenic flexure defined as proximal colon cancers and tumors located in the descending colon, sigmoid colon, rectosigmoid junction, and rectum defined as distal colorectal cancers (31, 32).

Tissue selection and processing

Beginning in 2006, archived, paraffin-embedded tissue specimens were requested from incident colorectal cancer cases diagnosed through December 31, 2002. In total, tissue specimens were retrieved from 732 of 1,255 (58%) cases, which is similar to colorectal cancer tissue retrieval rates recently reported from the Health Professionals Follow-up Study (51%; ref. 33) and the Nurses’ Health Study (58%; ref. 34). Women with tissue available for ER analysis were slightly older than those for whom tissue was not available (mean 73.9 vs. 72.1 years of age). Otherwise, subject demographics, exposure patterns, and tumor characteristics did not differ significantly between colorectal cancer cases with retrieved versus nonretrieved tissue specimens. All incident colorectal cancer diagnoses were confirmed by a single gastrointestinal pathologist. A total of 563 of 732 (77%) cases met criteria for the present study (i.e., confirmed first primary colorectal cancer with sufficient tissue for the planned laboratory analyses). Paraffin blocks were serially sectioned to 5 or 10 μm slices and placed on slides. The last slide was stained with hematoxylin and eosin (H&E) so that areas of neoplastic (defined as >50% dysplastic cells) and normal tissue could be defined and marked. From these marked slides, three tumor cores and two normal cores were taken from each block and placed into a tissue microarray (TMA) block along with liver controls. The TMA was produced by the Mayo Clinic Pathology Research Core (PRC) laboratory using the Beecher ATA-27 automated array. From the TMA, 5 μm sections were cut and placed on slides for H&E or IHC staining.

Characterization of ESR2 protein expression by IHC

IHC for ESR2 expression was performed by the PRC at the Mayo Clinic. Briefly, slides were deparaffinized and hydrated with distilled water, antigen retrieval was done by soaking slides in EDTA in a 98 to 100°C steamer for 30 minutes. A protein block was applied (DAKO X0909) and the primary antibody [ERβ antibody, clone PPG5/10 (Thermo Scientific #MA1-27412) at 1:25 dilution] was applied. The secondary horseradish peroxidase-labeled antibody was applied (DAKO K4061), chromagen DAB (DAKO K3468) was used, and the sections were counterstained with hematoxylin. Breast cancer tissue was used as a positive control and liver tissue for negative controls. Each section or core was scored by a pathologist (T.C. Smyrk) using a combination of the staining intensity (0–3) and percentage of cells stained (0%–0%, 1%–10%, 11%–30%, 31%–67%, 68%–100%).
The two scores were added for a combined score (0–8) as reported by Harvey and colleagues. Each case was classified as ESR2 negative if the combined score was 0, ESR2-low if the score was 1 to 5, and ESR2-high for a score of 6 to 8 (ref. 35; representative examples shown in Fig. 1). For each individual, the tumor core with the highest score was used for analysis.

Statistical analysis
Follow-up was calculated as age at completion of the baseline survey until age at first colorectal cancer diagnosis, age at move from Iowa, or age at death. If none of these events occurred, a woman was assumed to be alive, cancer-free, and living in Iowa through December 31, 2002.

Cox proportional hazard regression analysis was used to estimate relative risks (RR) and 95% confidence intervals (CI) for associations between exposures of interest and colorectal cancer subtypes defined by ESR2 protein expression status (negative, low, and high). Incidence was modeled as a function of age rather than time on study, because age is a better predictor of colorectal cancer risk on our cohort than follow-up time (36). Smoking was examined by overall status (never, ever, former, or current), average number of cigarettes smoked per day, and cumulative cigarette pack-years; MHT was examined by overall status (never, ever, former, or current) and duration of use; and folate was examined by quartiles of consumption. Tests for trend were carried out for each exposure variable by ordering the categorized values from lowest to highest category (e.g., never, former and current smoking groups for smoking status) and including the resulting variable as a linear term in the Cox regression model. Multivariable adjustments were applied. All models were adjusted for BMI, WHR, physical activity level, alcohol consumption, and daily intake of total calories, fat, sucrose, red meat, calcium, vitamin E, and methionine. Smoking analyses were also adjusted for MHT and folate. MHT analyses were also adjusted for smoking, folate, age at menarche, age at menopause, OC use, and history of DM. Folate analyses were additionally adjusted for smoking, MHT, and history of DM.

For all subtype analyses, the outcome variable was incident colorectal cancer with the ESR2 protein expression status of interest; all other colorectal cancer cases (including those with missing or unknown ESR2 status) were considered censored observations at the date of diagnosis. We determined whether risk ratios for smoking, MHT, and folate differed according to these cancer subtypes using a competing risk form of Cox proportional hazards analysis (37). This approach allowed us to model and test the interaction between an exposure (modeled as a covariate) and molecular/tumor subtype (modeled as a stratum variable).

Results
Informative environmental exposure and protein expression data were available for 491 of 563 (87%) incident colorectal cancer cases that met study criteria. Distribution by ESR2 expression level included 66 (13%) ESR2-negative, 126 (26%) ESR2-low, and 299 (61%) ESR2-high (Table 1).
associations failed to reach statistical significance ($P > 0.20$ for each), acknowledging low power for this test.

For MHT, a statistically significant, inverse association with ESR2-low tumors was observed when comparing never use with former (RR, 0.68; 95% CI, 0.44–1.07) and current (RR, 0.59; 95% CI, 0.28–1.23) use of MHT ($P_{\text{trend}} = 0.05$); there was also a trend with longer duration of MHT exposure (RR, 0.54; 95% CI, 0.26–1.13 for $\geq$5 years compared with no exposure; $P_{\text{trend}} = 0.04$).

Similar trends were observed in the ESR2-negative tumors, but numbers were very small (4 cases with current or $\geq$5 years use). No associations with MHT were observed for ESR2-high tumors. As with the smoking analyses, tests for heterogeneity in subtype-specific MHT associations did not reach statistical significance ($P > 0.40$).

Folate intake was not associated with colorectal cancer risks, either overall or for any ESR2 subtype.

### Discussion

In this prospective cohort study of older women, we found that increased smoking exposure appeared to influence ESR2-low and ESR2-high colorectal cancers to a greater degree than ESR2-negative tumors (although sample size was limited in some smoking categories and tests for heterogeneity were not statistically significant). In addition, longer duration of MHT use was associated with a decreased risk for colorectal cancers with ESR2-low and, to a lesser extent, ESR2-negative protein expression levels. Conversely, no statistically significant associations were observed for folate and ESR2-specific colorectal cancer subtypes. These novel data add to the body of literature from our previous molecular epidemiology studies of smoking, MHT, folate, and other exposure variables with colorectal cancer subtypes defined by MSI, CIMP, BRAF mutation, TP53 protein expression, or KRAS mutation status (2, 3, 15, 16, 18, 19).

Coupled with our previously published results, the IWHS molecular epidemiology data reported herein continue to support the hypothesis that smoking primarily influences colorectal cancer risk through the serrated pathway (12, 38–40). The serrated pathway appears to be initiated by $BRAF$ mutation and progresses through a serrated precursor (sessile serrated adenoma) to cancers characterized by mutant $BRAF$, high CIMP and, often, high MSI. Burnett-Hartman and colleagues found that serrated polyps were positively associated with cigarette smoking (41). Our group previously reported that smoking was associated with CIMP-positive, $BRAF$-mutated, and MSI-high tumors, linking this lifestyle habit to the serrated pathway of colorectal carcinogenesis (2). Interestingly, in lung tissue, smoking induces expression of $CYP1B1$, an enzyme that metabolizes both the tobacco carcinogens and estrogen and smoking also increases the carcinogenic estrogen metabolites (4-OHE). Some of the estrogen metabolites that are produced are known to activate the ER-mediated signaling pathways (27). Cleary and colleagues found a significant interaction between smoking status and $CYP1B1$ and other carcinogen metabolism gene variants in colorectal cancer (42). Together, these findings provide a biologically credible mechanism for the smoking-related risk associations observed in the IWHS cohort.

### Table 1. Distributions of cigarette smoking, hormone therapy, and folate intake by ESR2 tumor expression among incident colorectal cancer cases

<table>
<thead>
<tr>
<th>Attribute</th>
<th>ESR2-negative (N = 66)</th>
<th>ESR2-low (N = 126)</th>
<th>ESR2-high (N = 299)</th>
<th>Overall (N = 491)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age at baseline, mean (SD)</td>
<td>63.8 (4.3)</td>
<td>63.0 (3.8)</td>
<td>62.9 (4.1)</td>
<td>63.1 (4.3)</td>
</tr>
<tr>
<td>Age at colorectal cancer diagnosis, mean (SD)</td>
<td>73.6 (6.3)</td>
<td>73.9 (5.9)</td>
<td>73.9 (5.9)</td>
<td>73.9 (5.9)</td>
</tr>
<tr>
<td>Smoking status</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Never</td>
<td>47 (72.3%)</td>
<td>77 (61.6%)</td>
<td>191 (65.5%)</td>
<td>315 (65.9%)</td>
</tr>
<tr>
<td>Ever</td>
<td>19 (28.8%)</td>
<td>48 (38.4%)</td>
<td>103 (35%)</td>
<td>170 (35.1%)</td>
</tr>
<tr>
<td>Former</td>
<td>14 (21.2%)</td>
<td>29 (23.2%)</td>
<td>59 (20.1%)</td>
<td>102 (21%)</td>
</tr>
<tr>
<td>Current</td>
<td>5 (7.6%)</td>
<td>19 (15.2%)</td>
<td>44 (15%)</td>
<td>68 (14%)</td>
</tr>
<tr>
<td>Average number of cigarettes per day</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0</td>
<td>47 (72.3%)</td>
<td>77 (61.6%)</td>
<td>191 (65.2%)</td>
<td>315 (65.2%)</td>
</tr>
<tr>
<td>1–19</td>
<td>10 (15.4%)</td>
<td>23 (18.4%)</td>
<td>49 (16.7%)</td>
<td>82 (17%)</td>
</tr>
<tr>
<td>20–39</td>
<td>7 (10.8%)</td>
<td>19 (15.2%)</td>
<td>45 (15.4%)</td>
<td>71 (14.7%)</td>
</tr>
<tr>
<td>$\geq$40</td>
<td>1 (1.5%)</td>
<td>6 (4.8%)</td>
<td>8 (2.7%)</td>
<td>15 (3.1%)</td>
</tr>
<tr>
<td>Cumulative pack-years of cigarettes smoked</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0</td>
<td>47 (72.3%)</td>
<td>77 (61.6%)</td>
<td>191 (65.9%)</td>
<td>315 (65.9%)</td>
</tr>
<tr>
<td>1–19</td>
<td>10 (16.2%)</td>
<td>16 (13%)</td>
<td>36 (12.4%)</td>
<td>62 (13%)</td>
</tr>
<tr>
<td>20–39</td>
<td>7 (10.8%)</td>
<td>19 (15.2%)</td>
<td>45 (15.4%)</td>
<td>71 (14.7%)</td>
</tr>
<tr>
<td>$\geq$40</td>
<td>1 (1.5%)</td>
<td>6 (4.8%)</td>
<td>8 (2.7%)</td>
<td>15 (3.1%)</td>
</tr>
<tr>
<td>Hormone therapy</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Never</td>
<td>41 (62.1%)</td>
<td>87 (70.2%)</td>
<td>196 (66.7%)</td>
<td>324 (66.9%)</td>
</tr>
<tr>
<td>Ever</td>
<td>25 (37.9%)</td>
<td>37 (29.8%)</td>
<td>98 (33.3%)</td>
<td>160 (33.1%)</td>
</tr>
<tr>
<td>Former</td>
<td>21 (31.8%)</td>
<td>27 (21.8%)</td>
<td>67 (22.6%)</td>
<td>115 (23.8%)</td>
</tr>
<tr>
<td>Current</td>
<td>4 (6.1%)</td>
<td>10 (8.1%)</td>
<td>31 (10.5%)</td>
<td>45 (9.1%)</td>
</tr>
<tr>
<td>Duration of hormone therapy</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Never</td>
<td>41 (62.1%)</td>
<td>87 (70.2%)</td>
<td>196 (66.7%)</td>
<td>324 (66.9%)</td>
</tr>
<tr>
<td>$\leq$5 Years</td>
<td>21 (31.8%)</td>
<td>26 (21.1%)</td>
<td>68 (23.3%)</td>
<td>115 (23.9%)</td>
</tr>
<tr>
<td>$\geq$5 Years</td>
<td>4 (6.1%)</td>
<td>10 (8.1%)</td>
<td>28 (9.6%)</td>
<td>42 (8.7%)</td>
</tr>
<tr>
<td>Folate intake (mg/d)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>$\leq$250</td>
<td>16 (25.8%)</td>
<td>23 (20%)</td>
<td>70 (25.8%)</td>
<td>109 (24.3%)</td>
</tr>
<tr>
<td>251–350</td>
<td>17 (27.4%)</td>
<td>35 (30.4%)</td>
<td>82 (30.3%)</td>
<td>134 (29.3%)</td>
</tr>
<tr>
<td>351–573</td>
<td>14 (22.6%)</td>
<td>21 (18.5%)</td>
<td>66 (24.4%)</td>
<td>101 (22.5%)</td>
</tr>
<tr>
<td>$\geq$574</td>
<td>15 (24.2%)</td>
<td>36 (31.3%)</td>
<td>53 (19.6%)</td>
<td>104 (23.2%)</td>
</tr>
</tbody>
</table>

*Numbers may not sum to totals due to missing data.*
MHT has been shown to provide a protective effect on colorectal cancer risk (5–8, 15, 23). In our previous work, we found that MHT may reduce colorectal cancer risk in KRAS-WT tumors in the distal colorectum (16). We also found MHT to be associated with colorectal cancer risk in the proximal colorectum (17). These results seem to indicate that MHT in the distal colorectum may reduce colorectal cancer risk in people with a history of DM. We also found MHT to be associated with colorectal cancer risk in people with a history of DM. Folate analyses additionally adjusted for smoking, MHT, and history of DM.

Environmental Exposures and CRC Risk by ESR2 Expression Levels

<table>
<thead>
<tr>
<th>Attribute</th>
<th>Person years</th>
<th>N</th>
<th>RR (95% CI)</th>
<th>N</th>
<th>RR (95% CI)</th>
<th>N</th>
<th>RR (95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>从来没有</td>
<td>375,486</td>
<td>47</td>
<td>1.00 (Ref)</td>
<td>77</td>
<td>1.00 (Ref)</td>
<td>191</td>
<td>1.00 (Ref)</td>
</tr>
<tr>
<td>曾经吸烟</td>
<td>180,409</td>
<td>19</td>
<td>0.93 (0.53–1.64)</td>
<td>48</td>
<td>1.35 (0.91–2.00)</td>
<td>103</td>
<td>1.24 (0.96–1.61)</td>
</tr>
<tr>
<td>前吸烟</td>
<td>104,111</td>
<td>14</td>
<td>1.25 (0.67–2.28)</td>
<td>29</td>
<td>1.28 (0.81–2.03)</td>
<td>59</td>
<td>1.19 (0.87–1.61)</td>
</tr>
<tr>
<td>当前</td>
<td>76,297</td>
<td>5</td>
<td>0.54 (0.21–1.40)</td>
<td>19</td>
<td>1.46 (0.86–2.50)</td>
<td>44</td>
<td>1.33 (0.94–1.90)</td>
</tr>
</tbody>
</table>

| P-met | 0.40 | 0.12 | 0.08 |

平均每天吸烟数量

| 1–19 | 95,965 | 10 | 0.91 (0.45–1.85) | 23 | 1.14 (0.69–1.87) | 49 | 1.10 (0.79–1.52) |
| 20–39 | 73,546 | 7 | 0.82 (0.36–1.86) | 19 | 1.38 (0.81–2.37) | 45 | 1.35 (0.95–1.91) |
| ≥40 | 9,022 | 1 | 1.01 (0.14–7.38) | 6 | 4.24 (1.81–9.91) | 8 | 2.15 (1.05–4.41) |

Folate摄入（μg/d）

| ≤5 Years | 184,704 | 21 | 1.14 (0.66–1.97) | 26 | 0.69 (0.44–1.09) | 68 | 0.80 (0.60–1.08) |
| >5 Years | 60,064 | 4 | 0.41 (0.12–1.35) | 10 | 0.54 (0.26–1.13) | 28 | 0.89 (0.59–1.34) |

| P-met | 0.36 | 0.04 | 0.25 |

 smoking, age at menarche, age at menopause, OC use, and history of DM. Folate analyses additionally adjusted for smoking, MHT, and history of DM.

Further work is needed to determine the molecular mechanism for the possible protective effects of folate intake on colorectal cancer risk.

Relatively few prior studies have reported associations between the exposures of interest in this study and ESR2-defined colorectal cancer subtypes. Rudolph and colleagues found that colorectal cancer risk was significantly reduced with ESR2-positive tumor with current and longer duration MHT. Like our study, heterogeneity of association according to ESR2 status was not statistically significant (20). While we saw reduced colorectal cancer risk only with ESR2-low samples, it is hard to compare the results because we had a more complex category scale. It appears that our ESR2-low cases would fall into Rudolph’s ESR2-negative group (less than 10% strong staining or less than 50% weak staining). In both studies, there was the same correlation with at least some ESR2 expression. Although our population groups appear to be similar, there may be some subtle differences due to location, culture, or treatment protocol (Germany vs. Iowa). If we combine our ESR2-low and -negative categories, we have a higher proportion of patients with ESR2-high expression than Rudolph and colleagues (61% vs. 51%). We also used a different antibody in our study. Rudolph and colleagues used the 14C8 clone, which targets the N terminus of the protein, whereas our study used the PPGS/10 clone, which targets the C terminus of the protein. According to Skliris and colleagues, the loss of ESR2 is associated with more advanced stages of colorectal cancer, this could be a mechanism for the protective effect of ESR2-low expressions on colorectal cancer risk.

Our group previously reported no significant associations between folate intake and incidence colorectal cancer after adjustment for potential confounding factors, either overall or within molecular subtypes of MSI, CIMP, BRAF, TP53, or KRAS status (18, 19). In the current study, we also found no association between folate and colorectal cancer risk based on ESR2 status. Further work is needed to determine the molecular mechanism for the possible protective effects of folate intake on colorectal cancer risk.

Table 2. Associations of cigarette smoking, hormone therapy, and folate intake with incident colorectal cancer, by ESR2 tumor expression level

<table>
<thead>
<tr>
<th>Attribute</th>
<th>Person years</th>
<th>N</th>
<th>RR (95% CI)</th>
<th>N</th>
<th>RR (95% CI)</th>
<th>N</th>
<th>RR (95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Never</td>
<td>341,377</td>
<td>41</td>
<td>1.00 (Ref)</td>
<td>87</td>
<td>1.00 (Ref)</td>
<td>196</td>
<td>1.00 (Ref)</td>
</tr>
<tr>
<td>Ever</td>
<td>212,696</td>
<td>25</td>
<td>0.92 (0.54–1.56)</td>
<td>37</td>
<td>0.66 (0.44–0.99)</td>
<td>98</td>
<td>0.82 (0.63–1.07)</td>
</tr>
<tr>
<td>Former</td>
<td>151,535</td>
<td>21</td>
<td>1.02 (0.59–1.77)</td>
<td>27</td>
<td>0.68 (0.44–1.07)</td>
<td>67</td>
<td>0.87 (0.54–1.44)</td>
</tr>
<tr>
<td>Current</td>
<td>61,161</td>
<td>4</td>
<td>0.63 (0.22–1.78)</td>
<td>15</td>
<td>0.59 (0.28–1.23)</td>
<td>31</td>
<td>1.11 (0.74–1.64)</td>
</tr>
</tbody>
</table>

| P-met | 0.53 | 0.04 | 0.17 |

<table>
<thead>
<tr>
<th>Duration of hormone therapy</th>
<th>N</th>
<th>RR (95% CI)</th>
<th>N</th>
<th>RR (95% CI)</th>
<th>N</th>
<th>RR (95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>≤5 Years</td>
<td>184,704</td>
<td>21</td>
<td>1.14 (0.66–1.97)</td>
<td>26</td>
<td>0.69 (0.44–1.09)</td>
<td>68</td>
</tr>
<tr>
<td>&gt;5 Years</td>
<td>60,064</td>
<td>4</td>
<td>0.41 (0.12–1.35)</td>
<td>10</td>
<td>0.54 (0.26–1.13)</td>
<td>28</td>
</tr>
</tbody>
</table>

| P-met | 0.36 | 0.04 | 0.25 |

Cumulative pack-years of cigarettes smoked

| 1–19 | 74,225 | 10 | 1.27 (0.63–2.55) | 16 | 0.99 (0.55–1.78) | 36 | 1.09 (0.76–1.58) |
| 20–39 | 59,187 | 3 | 0.44 (0.15–1.43) | 15 | 1.30 (0.72–2.34) | 35 | 1.26 (0.86–1.84) |
| ≥40 | 42,566 | 5 | 0.90 (0.35–2.33) | 15 | 1.88 (1.05–3.36) | 28 | 1.42 (0.94–2.14) |

| P-met | 0.46 | 0.04 | 0.06 |

Folate intake (μg/d)

| ≤250 | 142,477 | 16 | 1.00 (Ref)  | 23 | 1.00 (Ref)  | 70 | 1.00 (Ref)  |
| >250 | 143,152 | 17 | 1.30 (0.61–2.77) | 35 | 1.65 (0.91–3.00) | 82 | 1.09 (0.75–1.58) |
| >573 | 142,999 | 14 | 1.37 (0.56–3.32) | 21 | 1.24 (0.60–2.56) | 66 | 0.76 (0.49–1.18) |
| >744 | 141,705 | 15 | 1.48 (0.52–4.16) | 36 | 2.09 (0.97–4.54) | 53 | 0.73 (0.44–1.20) |

| P-met | 0.46 | 0.12 | 0.31 |

NOTE: RRs and 95% CIs based on Cox proportional hazards regression analysis. All models adjusted for BMI, WHR, physical activity level, alcohol consumption, and daily intake of total calories, fat, sucrose, red meat, calcium, vitamin E, and methionine. Smoking analyses also adjusted for MHT and folate. MHT analyses also adjusted for smoking, FI, age at menarche, age at menopause, OC use, and history of DM.
some of the association trends that did not reach statistical
the exposure-subtype associations are relevant limitations to our
whole section approach, and reduce the run to run variability
replicates that would not have been feasible to assess using a
many more samples with normal and tumor cores, along with
By using TMAs for our IHC analyses, we were also able to stain
retrieved tissue samples from 58% of the colorectal cancer cases
population from which they were derived (47). In our study, we
are not representative of the broader subject cohort or target
accompanying mechanistic inferences. As cautioned by Ogino
colorectal cancer subtype-speci
fi
ology study design (47) permitted more focused evaluation of
cancer, likely due to the lack of suf
cient power. This is also
22.

We evaluated nuclear staining, but there are indications that
cytoplasmic staining may also be informative. Several groups
noticed a difference between normal tissue and tumor tissue
with the ESR2 staining location. Normal tissue tended to have
all nuclear staining, whereas tumor tissue had both nuclear and
cytoplasmic staining (22, 44). Examining this could help
explain the mechanism for loss of ESR2 protein in some
tumors. Traditionally, ERs are located in the nucleus where
they bind to estrogen and modulate gene expression. There are
also reports of plasma membrane ERs that induce more rapid
signaling (45, 46).

Notable strengths of our study include the detailed exposure
data and extended follow-up time available for IWHS subjects,
central pathology review, and near-complete colorectal cancer
case ascertainment. Use of the molecular pathologic epidemi-
ology study design (47) permitted more focused evaluation of
colorectal cancer subtype-specific exposure associations, with
accompanying mechanistic inferences. As cautioned by Ogino
and colleagues, selection bias can be introduced into molecular
pathologic epidemiology studies if the analyzed tumor samples
are not representative of the broader subject cohort or target
population from which they were derived (47). In our study, we
retrieved tissue samples from 58% of the colorectal cancer cases
requested (similar to other large cohort studies), without evi-
dence of selection bias based on specimen availability (2, 15).
By using TMAs for our IHC analyses, we were also able to stain
many more samples with normal and tumor cores, along with
replicates that would not have been feasible to assess using a
whole section approach, and reduce the run to run variability
that would have been present had each case been immuno-
tained separately.

The restricted demographic composition of our cohort (older
midwest women) and the relatively small sample sizes for some of
the exposure-subtype associations are relevant limitations to our
study. This can be seen in our tests for heterogeneity and with
some of the association trends that did not reach statistical
significance, likely due to the lack of sufficient power. This is also
evidenced by the large CIs in some of our comparisons performed
with limited sample numbers in category.

In addition, although we utilized a very extensive question-
naire, our study was still dependent on patient recall for the
analyzed exposure information, which may not be as reliable as
the molecular assay data. As discussed, the assessment of ESR2
status based on IHC results with one antibody rather than a more
comprehensive (and resource intensive) antibody panel to look at
different isoforms should be considered when interpreting our
results (42).

In conclusion, our data support the possibility of heteroge-
neous effects of MHT and smoking on ESR2-related pathways
of colorectal carcinogenesis in older women, while no clear associ-
betw between folate exposures and ESR2-related colorectal can-
tubertax subtypes was observed. These findings continue to support
the hypothesis that smoking primarily influences colorectal cancer
risk through the serrated pathway. Further evaluation of exposure-
related colorectal cancer risks based on independent and com-
bined molecular marker data in the IWHS cohort is ongoing,
which should provide additional clarity about the carcinogenic
mechanisms influenced by smoking, MHT, folate, and other
environmental factors.

Disclosure of Potential Conflicts of Interest
P.J. Limburg has ownership interest (including patents) in Exact Sciences. No potential conflicts of interest were disclosed by the other authors.

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Writing, review, and/or revision of the manuscript: I.S. Tillmans, R.A. Vier-
Administrative, technical, or material support (i.e., reporting or organizing
data, constructing databases): I.S. Tillmans, C.F. Lynch, J.R. Cerhan
Study supervision: P.J. Limburg

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Environmental Exposures and CRC Risk by ESR2 Expression Levels


Associations between Environmental Exposures and Incident Colorectal Cancer by ESR2 Protein Expression Level in a Population-Based Cohort of Older Women

Lori S. Tillmans, Robert A. Vierkant, Alice H. Wang, et al.

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