Variation of the *N-Acetyltransferase 2* Gene in a Romanian and a Kyrgyz Population

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Abstract

As part of a project on environmental disasters in minority populations, this study aimed to evaluate differences in the sequence of *N-acetyltransferase 2* (*NAT2*) as a metabolic susceptibility gene in yet unexplored ethnicities. Eight single nucleotide polymorphisms (SNP) in the *NAT2* coding region and a variant in the 3' flanking region were analyzed in 290 unrelated Kyrgyz and 140 unrelated Romanians by SNP-specific PCR analysis. The variants 341C, 481T, and 803G were less and 857A more prevalent in Kyrgyz (*P* < 0.0001). The variant at site 857 indicates Asian descent. 282C>T and 590G>A showed no significant variation by ethnicity. 364G>A and 411A>T turned out to be monomorphic. Database comparisons of the *NAT2* minor allele frequencies support that Romanians belong to Caucasians and Kyrgyz are in between Caucasians and East Asians. The distributions of predicted haplotypes differed significantly between the two ethnicities where the Kyrgyz showed a higher genetic diversity. The haplotype without mutations was more common in Kyrgyz (40.1% in Kyrgyz, 29.3% in Romanians). Accordingly, the imputed slow acetylator phenotype was less prevalent in Kyrgyz (35.2% versus 51.4% in Romanians). We found pronounced ethnic differences in *NAT2* genotypes with yet unknown effect on the health risks for environmental or occupational exposures in minority populations. (Cancer Epidemiol Biomarkers Prev 2006;15(1):138–41)

Introduction

The role of susceptibility genes in the development of chronic diseases is an important issue in occupational and environmental medicine. Arylamine *N-acetyltransferase 2* (*NAT2*) catalyzes the addition of an acetyl group from acetyl-CoA to a terminal nitrogen on substrates (1). *NAT2* is a gene with a 870-bp coding region mapped to chromosome 8p22. The GenBank accession number X14672 is commonly used as reference sequence (URL: http://www.ncbi.nlm.nih.gov). As of June 5, 2003, 36 human *NAT2* alleles were reported based on variations at 15 nucleotide positions (URL: http://www.louisville.edu/medschool/pharmacology/NAT.html). We further refer to this database as NAT database. An additional single nucleotide polymorphism (SNP) in the 3' flanking region was documented in the SNP500Cancer database of Cancer Genome Anatomy Project (ref. 2; URL: http://snp500cancer.ncbi.nlm.nih.gov) and in the database of Perlegen Sciences (ref. 3; URL: http://genome.perlegen.com). We further refer to these databases as SNP500 database and Perlegen database. First results of an international project on genetic susceptibility to environmental carcinogens revealed a large variation of allele frequencies in control populations by ethnicity (4). Here, we report on the genetic variation of *NAT2* in a European (Romanian) and Central-Asian (Kyrgyz) study population analyzed as a part of the European Commission-funded project “Investigation of the Risk of Cyanide in Gold Leaching on Health and Environment in Central Asia and Central Europe” (IRCYL; ref. 5).

Materials and Methods

Study Population. Unrelated subjects were selected for the analyses of *NAT2* gene variants from two population surveys of IRCYL. *NAT2* genotypes were obtained from whole blood samples collected with informed consent. The study was approved by the Ethics Committee of the Kyrgyz Medical Academy and by the Romanian Ministry of Health. In total, 140 Romanian and 290 Kyrgyz subjects were included.

Genotyping of NAT2 Variants. Genomic DNA from frozen blood samples was prepared at the Kyrgyz Scientific Center for Haematology (Bishkek, Kyrgyzstan) and the Institute of Public Health (Cluj-Napoca, Romania) using the QIAamp DNA Blood Maxi Kit (Qiagen, Hilden, Germany) according to the protocol of the manufacturer. DNA was then shipped to Institut für umweltmedizinische Forschung (Düsseldorf, Germany) and tested for its applicability in PCR assays. *NAT2* variants documented as polymorphic in the SNP500 database were selected for genotyping with the MassARRAY system (Sequenom, San Diego, CA; ref. 6). These variant sites comprised two synonymous polymorphisms (282C>T and 481C>T), six nonsynonymous SNPs (341T>C, 364G>A, 411A>T, 590G>A, 803A>G, and 857G>A), and a C>T polymorphism in 3' untranslated region (rs2552). Two variants (364G>A and 411A>T) turned out to be monomorphic in the IRCYL populations. No assay could be established for the rare polymorphism 191G>A. Genotyping call rates were ≥96.8%.

Statistical Analyses. *NAT2* analyses were calculated using SAS 8.02 (Cary, NC). Deviations from Hardy-Weinberg
equilibrium were examined with exact tests. The minor allele was defined as the less frequent nucleotide at the polymorphic site for which frequencies are shown with 95% confidence limits. To investigate differences in genotype distributions by χ² test, we used a Caucasian and a Chinese population from public databases (SNP500 database and Perlegen database). Haplotypes were inferred using PHASE version 2.0.2 based on coding SNPs with positional information (7, 8).

Further information on genotyping and the prediction of haplotypes can be found on the internet (URL: http://www.bgfa.ruhr-uni-bochum.de/specials/NAT2.php).

Discussion

As part of a project on environmental disasters (5), this study aimed to evaluate ethnic differences in genes involved in susceptibility to environmental agents. We described NAT2 genotypes, predicted haplotypes, and deduced phenotypes in a Central Asian (Kyrgyz) and a European (Romanian) study population. During the last decade, a high degree of variability in NAT2 alleles has been found at 15 nucleotide positions and documented in the NAT database. The SNP500Cancer Project verified 9 nucleotide positions of these 15 and an additional SNP in the 3' flanking region. A first decision has therefore to be made on the selection of sequence variants in studies of genetic susceptibility (10). We selected all NAT2 sequence variants which were verified as polymorphic by the SNP500Cancer Project for genotyping with matrix-assisted laser desorption/ionization time-of-flight mass spectrometry. Two variants (364G>A and 411A>T) turned out to be monomorphic in the Kyrgyz and Romanian study groups. No assay could be established for 191G>A, which characterizes African descent. Cascorbi and Roots (11) critically reviewed pitfalls in NAT2 genotyping and considered 282C>T and 341T>C in non-Africans to be sufficient to predict the acetylation phenotype. This has at least to be updated for 857G>A, which characterizes Asian descent.

Romanians settle in Central Europe and speak a Latin language from the time of colonization of Dacia by Roman ancestors. They were repeatedly invaded by Turkish-Mongolians. Overall, the NAT2 SNP distribution in Romanians was similar to other Caucasians with high frequencies of 481T and 803G (SNP500 database; ref. 12). Compared with the distribution of NAT2 SNPs in a group of Germans, the Romanians had a lower frequency of 341T (13). The Asian-specific SNP at site 857 is slightly more prevalent in Romanians which may reflect Mongolian invasions. The Kyrgyz settle in Central Asia, have been nomads, and speak a Turk language. They have Turkish, Scytho-Siberian, and Mongolian influences. The study population lives at Lake Issyk-Kul near China. The distribution of NAT2 SNPs in the

Table 1. Minor allele frequencies of NAT2 polymorphisms in Romanians, Kyrgyz, Caucasians, and Chinese

<table>
<thead>
<tr>
<th>Sequence variant</th>
<th>Minor allele frequency, % (95% confidence interval)</th>
<th>P, χ² test</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Romanian, n = 140</td>
<td>Caucasian*, n = 31</td>
</tr>
<tr>
<td>282T</td>
<td>37.1 (25.2-50.3)</td>
<td>30.0 (24.7-35.7)</td>
</tr>
<tr>
<td>590A (197Q)</td>
<td>33.9 (22.3-47.0)</td>
<td>28.6 (23.4-34.3)</td>
</tr>
<tr>
<td>341C (114T)</td>
<td>43.5 (31.0-56.7)</td>
<td>37.8 (32.2-43.8)</td>
</tr>
<tr>
<td>481C (268K)</td>
<td>43.3 (30.6-56.8)</td>
<td>38.2 (32.5-44.2)</td>
</tr>
<tr>
<td>803G (268K)</td>
<td>40.3 (281.3-53.6)</td>
<td>36.8 (31.1-42.7)</td>
</tr>
<tr>
<td>857A (286E)</td>
<td>3.2 (0.4-11.2)</td>
<td>1.4 (0.4-3.6)</td>
</tr>
</tbody>
</table>


### Notes
- N.d., no data available.
- Caucasians are defined as a population with a greater than 20% frequency of 282C>T and 590G>A (3.2% of 857A has been observed among the Kyrgyz (12.1% versus 3.2%)).
- Sixteen Romanians (11.4%) and 47 Kyrgyz (16.2%) were homozygous for the reference sequence. A total of 54 study participants (12.6%) of either Romanian or Kyrgyz origin were estimated to carry a SNP combination not prevalent in the other study group. For all cases with more than one heterozygous SNP, many potential alleles could be related. Expected frequencies of predicted haplotypes based on coding SNPs are reported in Table 2. In analogy to the allele nomenclature [prefix asterisk (*)], we used the prefix number (#) to indicate predicted haplotypes. In both populations, the haplotypes #4, #5B, and #6A account for >80% of the predicted haplotypes. Overall, the distribution of haplotypes in the two study populations was different (permutation test, P = 0.01). We found ethnic differences in expected haplotype frequencies of >10% for #4, #5B, and #7B. The latter haplotype was mainly found in Kyrgyz subjects with an expected frequency of 11.6% based on 857A.

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Chinese investigated for Perlegen Sciences (http://genome.perlegen.com).
Kyrgyz study population resembled more than that of Caucasians although the allele frequencies were in between these populations (Perlegen database; ref. 14).

Comparing Romanians with Kyrgyz, there was a slightly larger proportion of Kyrgyz than Romanians which were homozygous carriers of the reference sequence at the investigated sites. The reference sequence was found more common in Asians than in Europeans and is even predominant in Amerindians but rare in Africans (12-17). We found a pronounced variation in four NAT2 polymorphisms which might indicate characteristics of population genetics. There was no obvious association of ethnic differences on whether a SNP results in an exchange of an amino acid or not. The mutations 282C>T and 590G>A showed no significant differences. The synonymous mutation 282C>T is common in a variety of ethnicities with low variation (SNP500 database; refs. 12, 13, 16). The nonsynonymous mutation 590G>A occurred frequently in both Caucasians and Asians but is rare in Amerindians (16). The alleles 341C, 481T, and 803G were less prevalent in Kyrgyz than in Romanians but they occur in higher frequencies in Chinese and even more in Amerindians (Perlegen database; ref. 16). Interestingly, the allele frequencies of these three variants were similar in each population: ≥40% in Caucasians, ~20% in Kyrgyz, 6% in Chinese, and <3% in Ngawbe Indians. The allele 857A is rare in Caucasians. Its prevalence was 12% in Kyrgyz but >20% in Pacific Rim and Amerindians (16, 18).

Regarding genetic diversity, the majority of subjects in the IRCYL study populations were carriers of more than one heterozygous NAT2 variant. For this group, alleles can be determined by cloning and consecutive sequencing of the single DNA strands. With SNP-based PCR methods, haplotypes can only be deduced. Thirty haplotypes were predicted in the Kyrgyz and nine haplotypes in the Romanian study population, indicating a higher genetic diversity of the Kyrgyz. Haplotypes #4, #5A, and #5B, and #6B were not observed in the Romanian study population, indicating a higher genetic diversity of the Kyrgyz. Haplotypes #4, #5B, and 6A comprised 92% of the Romanian NAT2 variants. These and haplotype #7B with the mutation at site 857 explained 95% of the Kyrgyz variants. The haplotype #5B occurred with a higher frequency in Caucasians and with a lower frequency in the Kyrgyz from Central Asia but was rare in East Asians and Amerindians (12-17).

The human NAT2 gene is supposed to be a susceptibility factor in the metabolism of xenobiotics and, in particular, in the development of bladder cancer where slow acetylators have been associated with an excess risk (19). NAT2 genotyping has been growingly employed to predict the phenotype. Slow acetylators vary by geographic region. About 50% of Caucasians are potential slow acetylators but there is a lower fraction in Asian populations (20). We also found a lower frequency of predicted slow acetylators in Kyrgyz in comparison with Romanians. In Chinese, there is an even lower fraction of slow acetylators (14, 21). Slow acetylators can be underestimated when only a few SNPs are used to predict the phenotype (10). The degree of misclassification in the prediction of the phenotype from the NAT2 genotype has been estimated to be up to 7% (13, 22-24).

In summary, we confirmed a large ethnic variation of NAT2 gene sequence. From the evolutionary perspective, this points to a selectively neutral character of the NAT2 gene (16). The variant at site 857 characterizes Asian descent. The NAT2 genotype distribution in Romanians was comparable to other Caucasians whereas Kyrgyz had minor allele frequencies in between Caucasians and East Asians. Ethnic differences in metabolic susceptibility genes indicate differences in metabolic pathways with yet unknown effect on the metabolic capacity and disease risks.

Acknowledgments

We thank the Romanian and Kyrgyz IRCYL teams for excellent field work, Tina Müller for statistical assistance, and Lucia Jorge-Nebert and Daniel Nebert for very fruitful comments.

Table 2. Expected frequencies for predicted NAT2 haplotypes in the IRCYL study populations

<table>
<thead>
<tr>
<th>Haplotype</th>
<th>Expected frequency, % (SE)†</th>
<th>Romansians</th>
<th>Kyrgyz</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>282C&gt;T</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>C</td>
<td>T</td>
<td>90.0 (0.11)</td>
<td>90.0 (0.11)</td>
</tr>
<tr>
<td>341T&gt;C</td>
<td>C</td>
<td>90.0 (0.11)</td>
<td>90.0 (0.11)</td>
</tr>
<tr>
<td>481C&gt;T</td>
<td>G</td>
<td>90.0 (0.11)</td>
<td>90.0 (0.11)</td>
</tr>
<tr>
<td>590G&gt;A</td>
<td>A</td>
<td>&lt;0.1 (0.03)</td>
<td>&lt;0.1 (0.03)</td>
</tr>
<tr>
<td>803A&gt;G</td>
<td>G</td>
<td>&lt;0.1 (0.03)</td>
<td>&lt;0.1 (0.03)</td>
</tr>
<tr>
<td>857G&gt;A</td>
<td>G</td>
<td>&lt;0.1 (0.03)</td>
<td>&lt;0.1 (0.03)</td>
</tr>
</tbody>
</table>

*NAT2 haplotype nomenclature with prefix number (#) in analogy to NAT2 allele nomenclature with prefix asterisk (*).
†Standard error.
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