Resveratrol-Induced Gene Expression Profiles in Human Prostate Cancer Cells

Sunita B. Jones, 1 Samuel E. DePrimo, 1 Michael L. Whitfield, 2 and James D. Brooks 1

Departments of Urology and Genetics, Stanford University School of Medicine, Stanford, California

Abstract

Objective: The transhydroxystilbene resveratrol is found at high levels in red wine and grapes, and red wine consumption may be inversely associated with prostate cancer risk. To gain insights into the possible mechanisms of action of resveratrol in human prostate cancer, we did DNA microarray analysis of the temporal transcriptional program induced by treatment of the human prostate cancer cell line LNCaP with resveratrol.

Methods: Spotted DNA microarrays containing over 42,000 elements were used to obtain a global view of the effects of resveratrol on gene expression. Prostate-specific antigen (PSA) and androgen receptor (AR) expression were determined by Northern blot and immunoblot analyses. Cell proliferation was determined by the 3-(4,5-dimethylthiazolyl-2)-2, 5-diphenyltetrazolium bromide assay and cell cycle analysis by flow cytometry.

Results: We observed time-dependent expression changes in >1,600 transcripts as early as 6 hours after treatment with resveratrol. Most striking was the modulation of a number of important genes in the androgen pathway including PSA and AR. Resveratrol also down-regulated expression of cell cycle and proliferation-specific genes involved in all phases of the cell cycle, and inhibited cell proliferation in a time- and dose-dependent manner.

Conclusion: Resveratrol produces gene expression changes in the androgen axis and cell cycle regulators that may underlie its putative anticancer activities in prostate cancer. (Cancer Epidemiol Biomarkers Prev 2005; 14(3):596–604)

Introduction

The most diagnosed cancer among men, prostate cancer will claim an estimated 29,900 lives this year in the United States alone (1). Considerable effort has been devoted to detecting and treating localized prostate cancer, and limited progress has been made in the treatment of recurrent or advanced disease. Epidemiologic evidence and two intervention trials have fueled interest in developing chemopreventive strategies for prostate cancer (http://www.crab.org/select/; ref. 2). Thus far, selenium, vitamin E, lycopene, cruciferous vegetables, and antiandrogens have been proposed as potential prostate cancer chemopreventive agents (3, 4). The recent increase in association of red wine intake with prostate cancer risk (5) led us to wonder whether resveratrol, a polyphenol transhydroxystilbene found at high levels in grapes and red wine, might exert biological effects that protect against prostate cancer.

Resveratrol appears in substantial quantities in several foods. Red wine contains between 0.72 and 3.18 mg/L resveratrol (6-8). Resveratrol is rapidly absorbed by the gut and shows excellent tissue bioavailability (9-14). The effects of resveratrol in biological systems are wide-ranging, and several studies have shown that it can inhibit or modulate metabolic pathways, act as an anti-inflammatory agent or antioxidant, and block cell proliferation (15-21). In prostate cancer cell lines, resveratrol has been shown to block proliferation and possibly act as a negative regulator of androgen pathways (22, 23).

DNA microarray technology has provided insights into the molecular taxonomy of human tumors as well as the transcriptional underpinnings of the cell cycle, prostate cellular senescence, cellular response to stress, and androgen action (24-32). To gain insights into the possible mechanisms of action of resveratrol in human prostate cancer, we did DNA microarray analysis of the temporal transcriptional program induced by treatment of the human prostate cancer cell line LNCaP with resveratrol. Based on these findings, we further investigated the effects of resveratrol on androgen pathways and the cell cycle.

Materials and Methods

Cell Culture and Treatments. The LNCaP cell line was obtained from American Type Culture Collection (Manassas, VA) and maintained in RPMI medium with 10% fetal bovine serum (CDT; Hyclone Laboratories, Logan, UT) and penicillin/streptomycin (Mediatech, Herndon, VA) in an environment of 95% air and 5% CO2 at 37°C. Upon reaching 75% confluence, cells were treated with either DMSO control or purified resveratrol at several concentrations (LKT Laboratories, St. Paul, MN), dissolved in DMSO, and incubated for varying lengths of time. Final concentration of DMSO in media was 0.01%. Total RNA was prepared from cells using TRIZol (Invitrogen Life Technologies, Carlsbad, CA).

Microarray Hybridizations and Data Analysis. Stanford Human cDNA microarrays were used in this study. Each microarray contains 42,000 elements representing ~30,000 UNigene Clusters (32). Microarray hybridizations were done as previously described (The Brown Lab: http://brownlab.stanford.edu; ref. 33). Briefly, 100 μg total RNA from each sample was reverse-transcribed and labeled with fluorescein-tagged nucleotides (Cy3-dUTP for the DMSO control samples; Cy5-dUTP for the resveratrol-treated samples). Pairs of resveratrol-treated and DMSO control–labeled cDNA were hybridized to spotted DNA microarrays containing over 42,000 elements. Pairs of hybridizations were normalized by taking the mean of each spot. Raw data were converted to log2 of the ratio of Cy5 to Cy3 for each spot.

Received 5/31/04; revised 9/15/04; accepted 10/22/04.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Copyright © 2005 American Association for Cancer Research.
mixed and hybridized to microarray slides for 14 to 18 hours at 65°C. Microarrays were scanned with a GenePix microarray scanner (Axon Instruments, Union City, CA) and analyzed with GenePix software (34). Spots of insufficient quality were excluded from further analysis by visual inspection. All data are stored in the Stanford Microarray Database at http://genome-www.stanford.edu/microarray. Only spots with a fluorescent intensity in each channel >2.0-fold over background were selected for further analysis; data points that did not meet this criteria were excluded. Genes missing >20% of their data points were also excluded from further analysis. Genes were selected that exhibited a ≥2-fold change in expression level relative to the control sample in at least two experiments. Application of different filtering and data selection criteria resulted in highly reproducible gene clustering patterns demonstrating the robustness of the data set. All microarray data can be downloaded at http://genome-www.stanford.edu/microarray/ (35).

Results

To gain insights into the potential mechanisms through which resveratrol acts as a putative cancer preventive agent, we used DNA microarrays to assess global gene expression patterns in a prostate cancer cell line, LNCaP, after treatment with resveratrol. As a first test of whether resveratrol could modulate gene expression patterns, LNCaP cells were treated with 10 nmol/L, 100 nmol/L, 1 µmol/L, 10 µmol/L, 25 µmol/L, 40 µmol/L, and 100 µmol/L resveratrol. Total RNA was harvested between 18 and 40 hours. These RNAs (labeled with the Cy-5 fluorophore) were cohybridized with RNA from cells treated in parallel with vehicle alone (labeled with Cy-3) on spotted cDNA microarrays containing 42,264 elements representing ~27,286 unique genes (estimated by UNIGene clusters). Only minor gene expression changes were noted at low doses (10 nmol/L–1 µmol/L); however, consistent, dose-dependent changes in gene expression were observed starting at 10 µmol/L that increased up to 100 µmol/L (Fig. 1; Supplementary Fig. S1).

Based on the observed expression changes in our dose-escalation experiments, we assessed the genome-wide patterns of gene expression induced after treatment with either 75 or 150 µmol/L resveratrol in LNCaP cells at time points between 0 and 60 hours after treatment (Fig. 1). We selected 1,656 transcripts whose expression levels changed at least 2-fold from the control samples on at least two arrays. These 1,656 transcripts were further analyzed by hierarchical clustering, revealing groups of genes that varied both in their magnitude and temporal patterns of gene expression in a resveratrol-dependent manner. Six hundred fourteen transcripts (37%) were induced and 1,044 transcripts (63%) were repressed following treatment with resveratrol. Changes in transcript levels were detected as early as 1 hour in a few genes and was apparent in most genes by 8 hours. Many transcripts represented named genes, although most were poorly characterized, and 198 (12%) of the genes were uncharacterized expressed sequence tags.

The human prostate cancer cell line LNCaP expresses the androgen receptor (AR) and responds to androgen stimulation (36, 37). DNA microarrays have been used to characterize the transcriptional program induced by treatment of LNCaP cells with dihydrotestosterone and R1881, a synthetic androgen analogue (31, 35, 38). Deprimo et al. (31) reported 567 androgen-responsive genes and, of these, 517 showed greater repression at 75 µmol/L than at 150 µmol/L. The 105 genes normally repressed by androgens, 92 (88%) were induced by resveratrol. A subset of genes (19%) was induced both by resveratrol and androgen and included genes involved in lipid metabolism, protein trafficking, vesicle formation, and stress response.
PSA, a well-characterized androgen-regulated gene, was repressed >2-fold by 12 hours after treatment with resveratrol. In agreement with transcript levels measured on the microarrays, Northern blot analysis on RNA from LNCaP cells treated with either 75 or 150 μmol/L resveratrol showed significant decreases in PSA mRNA levels (Fig. 3A). PSA protein levels were measured in the media of LNCaP cells treated with either 75 or 150 μmol/L resveratrol (Fig. 3B). In accord with the gene expression data, resveratrol-treated cells failed to accumulate PSA in the media compared with DMSO-treated control cells. PSA is regulated by the AR, and AR transcript levels were reduced >2-fold within 12 hours of treatment in the microarray data set. We tested whether resveratrol modulated AR levels using Western blot analysis and found a decrease in AR protein levels within 24 hours after treatment with 150 μmol/L resveratrol and within 36 hours in cells exposed to 75 μmol/L resveratrol (Fig. 3C). Somewhat surprisingly, however, decreased expression of AR target genes occurred by 12 hours, well before AR protein levels decreased.

In addition to its effects on the androgen axis, resveratrol produced complex temporal changes in the expression of genes involved in the cell division cycle. Whitfield et al. (28) have comprehensively identified genes whose expression varies periodically as synchronized cells traverse the cell cycle. Genes identified as cell cycle regulated were further grouped according to the phase of the cell cycle at which their expression is highest. By comparing the genes that change expression in the resveratrol-treated cells with those periodically expressed in the cell cycle, we identified 442 transcripts that are cell cycle regulated and change at least 2-fold in expression after treatment with resveratrol (Fig. 4A). Changes in expression levels were observed in genes associated with every phase of the cell cycle. Among the 442 genes in the cluster, 78 (18%) were G1-S phase specific, 81 (18%) S phase, 99 (22%) G2 phase, 120 (27%) G2-M phase, and 64 (14%) were M-G1 phase–specific genes (Fig. 4B). Approximately 20% of the 442 cell cycle–regulated genes showed increased expression after treatment with resveratrol, including several...
negative regulators of proliferation (PA26, TSG101, PCAF, and HDAC3). Most genes (80%) showed decreased expression after resveratrol treatment by 8 hours and remained repressed over the remainder of the time course. Transcript levels were suppressed over the entire time course and include many of the genes necessary for all the basic processes required to duplicate a human cell (cell cycle control, DNA replication, spindle assembly, mitosis; Fig. 4B). Also seen were proapoptotic genes JUND, IPLA2, TP53INP1, BOK, PA26, MDM2, RRM2B, PIGPC1, SARS, PDCD4, and STK17A (Fig. 1B) that were induced after treatment with 75 and 150 μmol/L resveratrol.

Because resveratrol produced large-scale changes in expression of cell cycle–regulated genes, we tested whether resveratrol affected LNCaP cell growth by using an MTT assay (ATCC). In agreement with the microarray data, resveratrol produced a dose- and time-dependent inhibition of cell growth (Fig. 5). Resveratrol (25 μmol/L) inhibited cell growth by only 3% after 24 hours and 16% by 60 hours, corresponding well with limited effects on gene expression seen at that dose. More dramatic growth inhibition was seen with 75 and 150 μmol/L doses that produce significant changes in gene expression of the cell cycle–regulated genes. Resveratrol (75 μmol/L) inhibited cell growth by 16% at 24 hours and 83% by 60 hours after treatment, whereas 150 μmol/L inhibited growth by 53% at 24 hours and 86% by 60 hours.

Flow cytometry was used to further characterize the effects of resveratrol on cell growth. As expected, 25 μmol/L resveratrol produced only minor changes in the cell cycle, with the portion of cells in S phase increasing from 12% in control cells to 18% in treated cells by 60 hours (Fig. 6). A much more dramatic effect was evident at 75 and 50 μmol/L resveratrol, where disappearance of the G2-M peak and accumulation of cells in S phase was observed. By 48 hours, cells in S phase comprised 27% of the total when treated with 75 μmol/L resveratrol and 30% in 150 μmol/L resveratrol-treated cells. A sub-G1 peak was observed in cells treated with 75 and 150 μmol/L resveratrol after 48 hours, although a corresponding sub-G1 peak was absent in cells treated with 25 μmol/L resveratrol even after 60 hours. These findings suggest that resveratrol induces apoptosis at high doses and this may, in part, account for the significant effects seen on MTT assay.

Discussion

The effects of resveratrol on the AR and androgen-responsive genes seem to be complex. In LNCaP cells, resveratrol has been shown to suppress secretion of PSA, although controversy exists as to whether this decrease is due to decreased expression of the AR or is independent of AR signaling.
Resveratrol Treatment of Prostate Cancer Cells

The gene expression data suggests that the down-regulation of androgen-responsive genes is not solely because of decreased levels of the AR. Resveratrol treatment produced an early and sustained decreased expression of many androgen-responsive genes (KLK2, KLK3, KLK4, AlbZIP, NXX3, FKB5, TMEPA1) long before AR protein levels were diminished. Furthermore, the decreased expression of androgen-responsive genes occurred even at low doses of resveratrol, whereas the decreases in AR transcript levels occurred only at very high doses. Finally, resveratrol did not oppose all transcriptional changes induced by androgen. A subset of 153 genes was up-regulated by both resveratrol and androgen. However, many of these genes, such as JUNB, HSP40, SERP1, and STCH seem to reflect cellular stress. In LNCaP, androgen treatment is known to produce cellular stress by inducing an oxidative burst, and this stress pattern has been observed in other gene expression profiles (31, 40). Resveratrol treatment undoubtedly places these cells under stress because they undergo cell cycle arrest and, at higher doses, apoptosis.

How resveratrol might affect expression of androgen-regulated genes, aside from its effects on AR protein levels, is unclear. It is possible that resveratrol acts, in part, as an androgen-receptor antagonist, or by blocking androgen-signaling pathways downstream of AR. Resveratrol has been shown to have partial agonist effects in estrogen-responsive mammary cancer cells and estrogens have been used to treat prostate cancer (41-43). This raises the possibility that resveratrol exerts its effects through other steroid signaling pathways. Whether resveratrol acts directly as an antiandrogen by binding to the AR, or indirectly through its estrogenic effects, awaits further study. Other putative prostate cancer-preventive agents, including vitamin E, lycopene, and methlyselenic acid, an organic selenium compound, have been reported to modulate androgen signaling in LNCaP cells through unknown mechanisms (44-49).

We have found that resveratrol inhibits the proliferation of LNCaP cells in a dose- and time-dependent manner. Cell growth arrest occurred in G1 and S phase of the cell cycle as shown by flow cytometry combined with a concomitant decrease in the number of cells in G2-M (Fig. 6). Dramatic gene expression changes accompany this cell cycle arrest and are distributed throughout all phases of the cell cycle. Resveratrol suppressed transcript levels for genes involved in cell cycle control, such as cyclins D, E, A, and B (Fig. 4). Negative regulators of proliferation, such as cyclins G1 and G2 and PA26 and HDAC3 (Fig. 1B), were induced in response to resveratrol and likely contribute to S-phase arrest. Therefore, resveratrol-induced growth arrest seems to be mediated by a complex network of cell cycle regulatory genes. The results observed here confirm other studies of gene expression demonstrating that cell cycle arrest results in decreased expression of the cell cycle-regulated genes (28). The cell cycle-regulated genes seem to be regulated such that expression occurs primarily when cells are dividing and not at any other time.

Considerable work has been published on resveratrol and its effect on the cell cycle and is consistent with the results presented here. Most studies suggest that resveratrol arrest cells in S phase and at the S/G2 transition. In a study using HL-60 cells, resveratrol exposure resulted in accumulation of cells at G1 and at the S phases with and an absence of the G2-M peak (20). This was attributed to an increase in the levels of cyclins A and E along with accumulation of phosphorylated cdk2. Hsieh et al. (50) reported that resveratrol induced NO synthase in pulmonary epithelial cells with suppression of the cell cycle through the S and G2 phases. Cell cycle arrest was accompanied by a corresponding increase in the expression of p53 and p21 and apoptosis. Inhibition in cell cycle progression by resveratrol by inducing S-phase arrest was also reported in osteoblasts, breast, colon, and prostate cancer cells (51, 52). In breast cancer cells, resveratrol caused an accumulation of cells in the S phase with a concomitant reduced expression of Rb and increased expression of p53 and bcl-2 proteins (53). Resveratrol-mediated growth inhibition and apoptosis in prostate cancer were observed in androgen nonresponsive cell lines with a disruption in the G1-S phase transition (54) and S-phase arrest in androgen responsive LNCaP cells (55).

Resveratrol has been reported to induce apoptosis via several pathways including DNA damage (56, 57) through p53-dependent and p53-independent pathways (16, 56-59) and by affecting lipoxygenase and cyclooxygenase activities (60). Narayanan et al. (61), using highly purified resveratrol and a different microarray platform, reported induction of apoptosis at 10 μmol/L in LNCaP that is correlated with modulation of p53-mediated molecular targets. We see similar regulation of p53 genes, but only at relatively high levels of resveratrol. In addition, we see apoptosis only at the highest concentration used in our study. In our gene expression data set, a few apoptosis genes were modulated, but not enough to elucidate pathways involved in apoptosis. It is, therefore, possible that critical apoptotic signals occur posttranscriptionally.

**Figure 3.** Effects of resveratrol on PSA and AR. A, Northern blot analysis of PSA mRNA in LNCaP cells shows decreased PSA expression over time. Equal loading was confirmed by rehybridization of the stripped membrane with radiolabeled β-actin (bottom). B, dose- and time-dependent inhibition of accumulation of secreted PSA in media of LNCaP cells. Culture medium was collected at indicated time intervals after addition of resveratrol (75 and 150 μmol/L) for measurement of total PSA. PSA levels were normalized to cell density. C, immuno blot analysis of AR protein levels in LNCaP cells shows decreased expression after exposure to resveratrol (25, 75, and 150 μmol/L). Equal loading was determined by glyceraldehyde-3-phosphate dehydrogenase immunoblotting (not shown).
The microarray results reported here provide additional insights into the mechanisms of action of this compound in prostate cells. Resveratrol produced striking induction of quinone reductase (NQO1) transcript levels. Quinone reductase is tightly regulated at the transcriptional level, and has served as a surrogate for phase 2 enzyme responsiveness (62). Indeed, we observe coordinate induction of other phase 2 enzymes (MGST1, TXNRD1, GSTA2, and PRDX1) and glutathione synthetic pathways (UGDH). Induction of phase 2 enzyme activity by resveratrol has been reported in other model systems, but may be particularly relevant to prostate cancer. From its earliest stages, human prostate cancers lose expression of a critical carcinogen defense enzyme, glutathione S-transferase-π or GSTP1, because of extensive methylation of deoxycytidine residues in the 5'-regulatory regions of the GSTP1 gene (63). Loss of GSTP1 could render prostate cells susceptible to carcinogenesis by compromising their defenses against endogenous or exogenous electrophilic mutagens. Compensation for loss of GSTP1 expression by induction of global carcinogen defenses could protect against the DNA damage that contributes to prostate cancer initiation or progression.

We recognize that our findings, whereas provocative, have several limitations. We have analyzed comprehensively the gene expression changes induced in a single, androgen-sensitive prostate cancer cell line, LNCaP, and recognize that these findings should be validated in other model systems.

Figure 4. Expression of cell cycle and proliferation genes in LNCaP cells exposed to resveratrol. Color bands, saturation scales and treatment times are the same as in Fig. 1. A, overview of the cluster diagram generated by querying microarray data using a gene list containing cell cycle–regulated genes. The full image of this cluster diagram is viewable as Supplementary Fig. S4. B, transcript profiles of selected genes involved in each phase of the cell cycle. Resveratrol treatments, color bands, and saturation scales are the same as in Fig. 1. The full image of this cluster diagram is viewable as Supplementary Fig. S4.
An important factor in explaining the efficacy of resveratrol is the comparison of dosages used in vitro and in animal studies, with dosages that can be expected to be clinically effective in humans. Tissue bioavailability of resveratrol in rat kidney has been reported at 77.75 ng/h/mL (14) following a single administration of red wine containing 28.24 μg resveratrol. Animal and human studies have shown that resveratrol is rapidly absorbed in the gut, attaining highest concentration in the blood in 1 hour, and its accumulation in organs vary (64-66). No information exists regarding resveratrol levels in the prostate tissue for any species. In this study, we have shown changes in gene expression at 10 to 100 μmol/L, physiologically attainable levels in rats (64). However, the levels of resveratrol used in this study that result in gene expression modulation in vitro may or may not be attainable in man. At the very least, the relatively low concentration of resveratrol in grapes or wine will likely make it necessary for it to be given as a dietary supplement. Further clinical translational work is urgently needed to clarify the relationship between serum concentrations and levels achieved in the prostate and whether similar gene expression changes are observed in vivo.

Our data provides a global view of the potential mechanisms through which resveratrol may act in protecting against prostate cancer and serves as a resource for future investigations into its mechanisms of action. Resveratrol exerts antiandrogenic effects not strictly attributable to repression of AR expression, inhibits the cell cycle, induces apoptosis, and up-regulates enzymes of carcinogen defense. This data set serves as a resource for understanding the effects of resveratrol in the prostate and as a potential source of biomarkers of response in vivo.

Figure 5. Time course of resveratrol-induced inhibition of LNCaP cell proliferation determined using the MTT assay. Results are expressed as a percent of inhibition in treated cells compared with control cells treated with the vehicle DMSO only. Data represent means for three separate experiments. *P < 0.05 compared with control.

Figure 6. Fluorescence-activated cell sorting analysis of LNCaP cells after treatment with resveratrol. Cell cycle phase distributions were quantified by staining cells with propidium iodide. Results are expressed as percent of cells in G1, S, G2-M, and sub-G1 phase at each time point after exposure.
Acknowledgments

We thank Dr. Zijie Sun, Department of Urology, Stanford University School of Medicine, for generous donation of PSA and AR probes, Dr. Manju Sharma for assistance with the assays and many useful discussions, and Stanford Microarray Database and Core Facility for providing arrays and technical support with data storage and retrieval.

References


Resveratrol-Induced Gene Expression Profiles in Human Prostate Cancer Cells

Sunita B. Jones, Samuel E. DePrimo, Michael L. Whitfield, et al.


Updated version
Access the most recent version of this article at:
http://cebp.aacrjournals.org/content/14/3/596

Supplementary Material
Access the most recent supplemental material at:
http://cebp.aacrjournals.org/content/suppl/2005/04/06/14.3.596.DC1

Cited articles
This article cites 65 articles, 22 of which you can access for free at:
http://cebp.aacrjournals.org/content/14/3/596.full#ref-list-1

Citing articles
This article has been cited by 4 HighWire-hosted articles. Access the articles at:
http://cebp.aacrjournals.org/content/14/3/596.full#related-urls

E-mail alerts
Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions
To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.