

## Interaction of Glutathione *S*-Transferase *M1* and *T1* Genotypes and Malignant Melanoma<sup>1</sup>

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### Abstract

The  $\mu$  and  $\theta$  classes of glutathione *S*-transferases (GST) may affect the development of cutaneous malignant melanoma (CMM) by decreasing cellular oxidative stress in skin. These isozymes are absent in a large proportion of the population because of germ-line homozygous deletions in the genes encoding *GSTM1* and *GSTT1*. To determine the association between *GSTM1* and *GSTT1* homozygous deletions (*GSTM1 null* and *GSTT1 null*, respectively) and CMM, we studied 212 patients with CMM, 150 patients with CMM and dysplastic nevi (DN), 147 patients with DN alone, and 124 healthy persons without CMM or DN. Comparing CMM cases ( $n = 362$ ) to participants without CMM ( $n = 271$ ), we found no association with *GSTM1 null* [odds ratio (OR), 1.2; 95% confidence interval (CI), 0.86–1.6] or *GSTT1 null* (OR, 0.82; 95% CI, 0.56–1.2), either independently or in combination (OR, 1.4; 95% CI, 0.81–2.2), after adjusting for age. However, among the subset of participants with red or blond hair, those with CMM were twice as likely to carry *GSTM1 null* (OR, 2.2; 95% CI, 1.2–4.2) and nearly 10-fold more likely to carry both *GSTM1 null* and *GSTT1 null* (OR, 9.5; 95% CI, 1.2–73) compared with those without CMM. These data suggest that among persons with hair colors traditionally associated with increased risk for melanoma, absence of both *GSTM1* and *GSTT1* may act to further elevate CMM risk.

### Introduction

Increased levels of cellular oxidative stress can be harmful to the faithful replication of DNA. Oxidative stress resulting in lipid peroxidation and DNA hydroperoxide formation can be induced in skin by UV radiation (1). Because one function of

GST<sup>3</sup> is the reduction of these potential mutagens (2, 3), GST isozymes active in skin, including *GSTM1* and *GSTT1*, may play a role in protection against development of cutaneous neoplasms, including CMM. Polymorphisms in members of the GST family have been shown to influence risk of cancers at several sites, including nonmelanoma skin cancers (4) and multiple cutaneous skin cancers (4–6). Lafuente *et al.* (7) measured leukocytic *GSTM1* levels by enzyme-linked immunoassay and noted that CMM patients were twice as likely to have decreased levels of *GSTM1* compared with controls. Two other studies (4, 8) genotyped *GSTM1* using PCR and reported no significant difference in the proportion of those homozygous for the *GSTM1* deletion (*GSTM1 null*) between CMM patients and controls. One additional study (9) published in abstract form only reported a significant association between CMM and carriage of the *GSTT1* homozygous deletion (*GSTT1 null*) but not *GSTM1 null*. DN are atypical moles that are epidemiological risk markers of CMM and nonobligate precursors (10). Their association with GST status has not been reported.

To gain additional insight into the complex etiology of CMM, we designed a case-control study to evaluate the associations among *GSTM1* and *GSTT1*, pigmentation characteristics, DN, and CMM.

### Materials and Methods

#### Setting and Population

Participants in the present study were accrued into a case-control study at the Pigmented Lesion Clinic of the Hospital of the University of Pennsylvania between September 1997 and September 1999. The Institutional Review Board of the University of Pennsylvania that oversees research involving human beings approved this study, and informed consent was obtained from each participant. Any clinic patient with incident CMM, defined as CMM histologically diagnosed within a year of their clinic visit, was eligible to participate. Patients with a first-time clinical diagnosis of DN (within 1 year of their clinic visit) were also eligible to participate. Patients who had a clinical or histological diagnosis of DN 1 year or more before the date of their study participation were excluded from study. We did not select patients based on prior knowledge of family history of melanoma or other melanoma risk factors, and those with more than one primary CMM were not eligible to participate regardless of the date of their first CMM. For each clinic patient enrolled, we asked for the name of a healthy nonblood relative or acquaintance to contact as a potential control subject.

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<sup>3</sup> The abbreviations used are: GST, glutathione *S*-transferase; CI, confidence interval; CMM, cutaneous malignant melanoma; DN, dysplastic nevi; IQR, interquartile range; OR, odds ratio; SULT, sulfotransferase.

### Data Collection and Measurement

Information on pigmentation characteristics and sun exposure history was obtained from a brief self-administered questionnaire. We asked about natural hair color as a teenager, natural eye color, initial skin reaction after exposure to the first strong sunlight of summer, skin reaction after long and repeated sun exposure, and freckling on the face and upper back. One research nurse (R. H.) completed a full skin examination, excluding the scalp and genitalia, for all of the study participants, and the following information was recorded: total mole count, total number of large moles (>8 mm), number of DN, and presence or absence of freckling. A buccal sample was self-collected by each study participant using sterile cheek swabs (Cyto-Pak Cytosoft Brush; Medical Packaging Corporation, Camarillo, CA) to collect a germ-line DNA sample.

### *GSTM1* and *GSTT1* Genotyping

Genomic DNA from the buccal swab was extracted using the protocol of Walker *et al.* (11), and the complete gene deletion at *GSTT1* and *GSTM1* was determined by using PCR-based assays modified from the protocol of Rebbeck *et al.* (12). The modified protocol used exon 4 of the sulfotransferase (*SULT1E1*) gene to serve as a positive internal control for the amplification of *GSTM1* rather than the  $\beta$ -globin-positive internal control reported previously. These primers were *SULT* exon 4 forward, 5' TGG AGT TGC TTA ACC TTT ACT-3'; and *SULT* exon 4 reverse, 5' GAG AAC ACT TGA CTC TGG TTA C 3'. We did not distinguish the homozygous active from the heterozygous active genotype for either *GSTM1* or *GSTT1*.

### Data Coding

**Case Status.** We grouped participants according to the presence or absence of CMM. The CMM case group included patients with CMM alone and those with both CMM and DN. The control group included patients with DN alone and healthy participants without CMM or DN.

**GST Genotypes.** We dichotomized GST genotypes based on the presence of at least one active allele. "*GSTM1 active*" indicates the presence of at least one nondeleted *GSTM1* allele, whereas "*GSTM1 null*" indicates the presence of two deleted *GSTM1* alleles. Similarly, "*GSTT1 active*" indicates the presence of at least one nondeleted *GSTT1* allele, and "*GSTT1 null*" indicates the presence of two deleted *GSTT1* alleles. A composite GST genotype was created to reflect overall GST activity. "GST active" indicates the presence of at least one nondeleted active allele in either *GSTM1* or *GSTT1*, whereas "GST null" indicates both *GSTM1 null* and *GSTT1 null*.

**Nevi Outcomes.** We categorized the total number of DN and the total number of large nevi as 0, 1, 2–5, 6–9, and 10 or more. All of the classes of nevi, including banal nevi, DN, and congenital nevi, were included in the count of large nevi. Total number of nondysplastic nevi was categorized as 1–24, 25–49, 50–74, and 75 or more and includes counts of large and small ( $\leq 8$  mm) banal nevi and congenital nevi.

**Pigmentation Characteristics.** Questionnaire information on hair color was coded as red (including reddish-brown), blond, or dark (including light brown, medium brown, dark brown, gray, and black); on eye color as blue or gray, green or hazel, or dark (including light brown, dark brown, and black); on freckling as many, few, or none; on skin reaction to acute sun as burn and blister, burn without blister, or tan (including mild sunburn followed by a tan, no sunburn and no tan, tan with no

Table 1 Adjusted odds of CMM by pigmentation characteristics

Pigmentation characteristic	Without CMM <i>n</i> = 271		CMM <i>n</i> = 362		OR <sup>a</sup>	95% CI
	<i>N</i>	(%)	<i>N</i>	(%)		
<b>Hair color</b>						
Red or reddish-brown	8	(3)	43	(12)	4.6	(2.1–10.2)
Blond	55	(20)	76	(20)	1.3	(0.85–1.9)
Dark	208	(77)	243	(67)	1.0	
<i>P</i> = 0.0002						
<b>Eye color<sup>b</sup></b>						
Blue	101	(38)	158	(44)	1.6	(1.1–2.4)
Green, gray, or hazel	64	(24)	104	(29)	1.8	(1.2–2.7)
Light or dark brown	104	(39)	96	(27)	1.0	
<i>P</i> = 0.01						
<b>Skin reaction to acute sun<sup>c</sup></b>						
Burn and blister	26	(10)	49	(14)	1.5	(0.88–2.6)
Burn without blister	100	(37)	136	(38)	1.2	(0.87–1.8)
Tan	143	(53)	177	(49)	1.0	
<i>P</i> = 0.11						
<b>Skin reaction to chronic sun<sup>d</sup></b>						
No tan	10	(4)	26	(7)	2.6	(1.2–5.6)
Light tan	58	(22)	121	(34)	2.1	(1.5–3.1)
Medium or dark tan	198	(74)	209	(59)	1.0	
<i>P</i> = 0.0001						
<b>Freckling</b>						
Many	69	(25)	162	(45)	3.7	(2.4–5.8)
Few	104	(38)	134	(37)	1.9	(1.3–2.9)
None	98	(36)	66	(18)	1.0	
<i>P</i> = 0.0001						

<sup>a</sup> Adjusted for age and total number of DN.

<sup>b</sup> Excludes six persons with missing data on eye color.

<sup>c</sup> Excludes two persons who responded "never exposed to strong sunlight."

<sup>d</sup> Excludes eleven persons who responded "never had repeated exposure to sun."

sunburn, and no change in skin color); and on skin reaction to chronic sun as no tan, light tan, or medium to dark tan.

### Statistical Analysis

Median counts of nevi outcomes were compared nonparametrically using Mann-Whitney *U* Wilcoxon rank-sum tests. Using unconditional logistic regression models, we calculated age-adjusted ORs and 95% CIs for associations between pigmentation characteristics and CMM, freckling and nevi types and GST genotype, and GST genotype and CMM. For each model, independent variables were entered as (0,1) indicator variables, where 1 specified the presence of the variable and 0 specified the absence of the variable. Independent variables with more than two levels were considered as ordinal variables to test for linear trend. For all of the analyses of pigmentation characteristics and CMM, total number of DN was additionally adjusted for in the logistic model.

To evaluate effect modification, we determined age-adjusted ORs and 95% CIs of CMM within pigmentation categories. Because the stratum-specific number of subjects for some pigmentation characteristics was too small to support subgroup analysis, we combined categories to create dichotomous variables (hair color: red, reddish-brown, blond *versus* light brown, medium brown, dark brown, gray, and black; eye color: blue, gray, green, hazel *versus* light brown, dark brown, and black; freckling: any *versus* none; skin reaction to acute sun: burn without tanning, regardless of blistering *versus* tanning or no effect, regardless of burning; and skin reaction to chronic sun: no or light tan *versus* medium or dark tan). We chose to combine the moderate and high risk categories and

Table 2 Adjusted odds of *GSTM1* null, *GSTT1* null, and *GST* null by freckling and nevi type

	<i>GSTM1</i> active n = 330		<i>GSTM1</i> null n = 303				<i>GSTT1</i> active n = 491		<i>GSTT1</i> null n = 142				<i>GST</i> active n = 491		<i>GST</i> null n = 78			
	N	(%)	N	(%)	OR <sup>a</sup>	95% CI	N	(%)	N	(%)	OR <sup>a</sup>	95% CI	N	(%)	N	(%)	OR <sup>a</sup>	95% CI
<b>Freckling</b>																		
None	86	(26)	78	(26)	1.0		128	(26)	36	(25)	1.0		147	(26)	17	(22)	1.0	
Few	121	(37)	117	(39)	1.1	(0.65–1.4)	177	(36)	61	(43)	1.2	(0.77–2.0)	204	(37)	34	(44)	1.4	(0.78–2.7)
Many	123	(37)	108	(36)	0.97	(0.72–1.6)	186	(38)	45	(32)	0.86	(0.53–1.4)	204	(37)	27	(35)	1.1	(0.60–2.2)
					P = 0.83				P = 0.46				P = 0.79					
<b>Number of non-DN</b>																		
1–24	157	(48)	158	(52)	1.0		248	(51)	67	(47)	1.0		281	(51)	34	(44)	1.0	
25–49	68	(21)	46	(15)	0.64	(0.41–1.0)	82	(17)	32	(23)	1.5	(0.88–2.4)	98	(18)	16	(21)	1.4	(0.72–2.6)
50–74	30	(9)	37	(12)	1.2	(0.67–2.0)	53	(11)	14	(10)	0.99	(0.51–1.9)	58	(10)	9	(12)	1.3	(0.59–3.0)
74 or more	76	(23)	62	(20)	0.76	(0.49–1.2)	108	(22)	29	(20)	1.0	(0.60–1.7)	118	(21)	19	(24)	1.4	(0.72–2.7)
					P = 0.35				P = 0.96				P = 0.32					
<b>Number of DN</b>																		
0	175	(53)	157	(52)	1.0		257	(52)	75	(53)	1.0		294	(53)	38	(49)	1.0	
1	44	(13)	37	(12)	0.93	(0.57–1.5)	62	(13)	19	(13)	1.1	(0.59–1.9)	69	(12)	12	(15)	1.4	(0.68–2.8)
2–5	51	(15)	53	(17)	1.1	(0.72–1.8)	89	(18)	15	(11)	0.59	(0.32–1.1)	97	(17)	7	(9)	0.58	(0.25–1.4)
6–9	27	(8)	19	(6)	0.77	(0.41–1.4)	38	(8)	8	(6)	0.73	(0.33–1.7)	43	(8)	3	(4)	0.56	(0.16–1.9)
10 or more	33	(10)	37	(12)	1.2	(0.70–2.1)	45	(9)	25	(18)	2.0	(1.1–3.5)	52	(9)	18	(23)	2.8	(1.4–5.6)
					P = 0.73				P = 0.29				P = 0.08					

<sup>a</sup> Adjusted for age.

keep separate the low risk category in an attempt to highlight subgroups that identify persons at reduced risk of melanoma and persons at some increased risk of melanoma.

## Results

Our study sample included 362 CMM cases and 271 controls without CMM. All of the participants were Caucasian, and only five (two CMM cases and three controls) reported being of Hispanic origin. The gender distribution was similar among those with CMM (50% male) and those without (53% male;  $\chi^2 = 1.02$ ;  $P = 0.31$ ). The mean age of CMM cases ( $50.6 \pm 14.0$ ) was statistically significantly older than that of controls without CMM ( $45.1 \pm 13.4$ ;  $P = 0.0001$ ). No difference in ever-smoking was noted between the groups ( $\chi^2 = 0.28$ ;  $P = 0.60$ ).

**Association of Pigmentation Characteristics and CMM.** Adjusted ORs for CMM in relation to self-reported pigmentation characteristics are given in Table 1. The strongest predictors of CMM were freckling and skin reaction to chronic sun exposure. CMM cases were nearly four times as likely to report many freckles (OR, 3.7; 95% CI, 2.4–5.8) and nearly two times as likely to report few freckles (OR, 1.9; 95% CI, 1.3–2.9) compared with controls. Compared with controls, CMM cases were over twice as likely to report both an inability to tan (OR, 2.6; 95% CI, 1.2–5.6) and a light tanning ability (OR, 2.1; 95% CI 1.5–3.1) in response to chronic sun. A statistically significant linear trend was observed for these associations. Patients with CMM were over four times as likely to be redheaded (OR, 4.6; 95% CI, 2.1–10.2) compared with controls without CMM. Blond hair was not predictive of CMM (OR, 1.3; 95% CI, 0.85–1.9).

The total number of DN and total number of large nevi were highly and significantly correlated (Pearson  $r = 0.76$ ;  $P = 0.001$ ). Because the total number of large nevi was a composite measure that did not distinguish among large banal nevi, large congenital nevi, and large DN, it was not possible for us to separate the affects of large banal nevi from that of other types of large nevi. Thus, this measure was not considered in subsequent analyses.

Table 3 Adjusted odds of CMM by GST genotype

GST genotype	Without CMM n = 271		CMM n = 362		OR <sup>a</sup>	95% CI
	N	(%)	N	(%)		
<i>GSTM1</i> active	147	(54)	183	(51)	1.0	
<i>GSTM1</i> null	124	(46)	179	(49)	1.2	(0.86, 1.6)
<i>GSTT1</i> active	205	(76)	286	(79)	1.0	
<i>GSTT1</i> null	66	(24)	76	(21)	0.82	(0.56, 1.2)
<i>GST</i> active	241	(89)	314	(87)	1.0	
<i>GST</i> null	30	(11)	48	(13)	1.3	(0.76, 2.1)

<sup>a</sup> Adjusted for age.

**Association of Cutaneous Markers of CMM Risk and GST Genotype.** Comparing participants who were *GST* null ( $n = 78$ ) to those who were *GST* active ( $n = 555$ ), there were no statistically significant differences in the median number of total nevi (32.5; IQR, 8–74; and 24; IQR, 8–66, respectively) or median number of DN (1; IQR, 0–6; and 0; IQR, 0–3, respectively). Results were similar comparing median nevus counts across *GSTM1* and *GSTT1* status (data not shown). Table 2 shows age-adjusted ORs for associations of nondysplastic nevi, DN, and freckling with GST genotype. Persons with 10 or more DN were nearly three times as likely (OR, 2.8; 95% CI, 1.4–5.6) to be *GST* null. The linear trend for increasing categories of DN count did not reach significance at the  $\alpha = 0.05$  level. There was no association of reported freckling and GST genotypes.

**Association of CMM and GST Genotype.** We did not find a significant association with *GSTM1* null, *GSTT1* null, or *GST* null genotypes and CMM status (Table 3).

After stratifying the sample on pigmentation characteristics, we found a significant association of GST genotype and CMM in the subgroup of persons with light hair color (red, reddish-brown, or blond; Table 4). Among those with red or blond hair, cases with CMM were nearly 10-times more likely

Table 4 Age-adjusted odds of CMM by GST genotype stratified by pigmentation characteristic

	Hair color			Eye color			Skin reaction to acute sun			Skin reaction to chronic sun			Freckling					
	Brown or black	Red, reddish-brown, or blond	Light or dark brown	Blue, gray, green, or hazel	Burn then tan or tan	Burn without tanning	Medium or dark tan	Light or no tan	None	Any	Case/control		OR <sup>a</sup> 95% CI		Case/control		OR <sup>a</sup> 95% CI	
											Case/control	OR <sup>a</sup> 95% CI	Case/control	OR <sup>a</sup> 95% CI	Case/control	OR <sup>a</sup> 95% CI	Case/control	OR <sup>a</sup> 95% CI
<i>GSTM1</i>	126/105 1.0	57/42 1.0	41/49 1.0	138/96 1.0	87/69 1.0	96/77 1.0	103/103 1.0	75/40 1.0	34/52 1.0	149/95 1.0								
<i>GSTM1</i> active	117/103 0.99 (0.68–1.4)	62/21 2.2 (1.2–4.2)	55/55 1.3 (0.72–2.2)	124/69 1.3 (0.85–1.9)	90/74 0.99 (0.63–1.6)	89/49 1.5 (0.94–2.4)	106/95 1.1 (0.76–1.7)	72/28 1.4 (0.79–2.6)	32/46 1.1 (0.57–2.1)	147/78 1.2 (0.84–1.8)								
<i>GSTM1</i> null	194/154 1.0	92/51 1.0	75/82 1.0	207/121 1.0	143/112 1.0	143/91 1.0	162/151 1.0	118/50 1.0	52/76 1.0	234/129 1.0								
<i>GSTT1</i> active	49/54 0.71 (0.45–1.1)	27/12 1.3 (0.58–2.7)	21/22 1.1 (0.54–2.1)	55/44 0.71 (0.45–1.1)	34/31 0.83 (0.48–1.5)	42/35 0.77 (0.45–1.3)	47/47 0.94 (0.59–1.5)	29/18 0.66 (0.33–1.3)	14/22 0.84 (0.38–1.9)	62/44 0.79 (0.50–1.2)								
<i>GSTT1</i> null	211/179 1.0	103/62 1.0	81/93 1.0	229/146 1.0	152/128 1.0	162/111 1.0	179/176 1.0	129/60 1.0	57/90 1.0	257/151 1.0								
<i>GST</i> active	32/29 0.97 (0.56–1.7)	16/1 9.5 (1.2–73)	15/11 1.7 (0.72–3.9)	33/19 1.1 (0.60–2.0)	25/15 1.4 (0.69–2.8)	23/15 1.1 (0.54–2.2)	30/22 1.4 (0.77–2.6)	18/8 0.99 (0.40–2.5)	9/8 1.8 (0.61–5.1)	39/22 1.1 (0.61–1.9)								
<i>GST</i> null																		

<sup>a</sup> Adjusted for age.

(OR, 9.5; 95% CI, 1.2–73) to be *GST null* compared with controls without CMM. The association between *GSTM1 null* and CMM was also significant in those with light hair color, although to a more modest degree (OR, 2.2; 95% CI, 1.2–4.2). No other significant interactions with pigmentation characteristics were noted.

**Discussion**

The etiology of CMM is complex and likely involves multiple low penetrance susceptibility genes, interactions among these genes, the influences of environmental exposures such as UV light, and the interaction of genotypes and environments. *GSTM1* and *GSTT1* represent only two of numerous potential candidate CMM susceptibility genes. Similar to other published reports (4, 8), our data do not support a strong role of *GSTM1* or *GSTT1* in the development of CMM, either individually or in combination. However, *GSTM1 null*, predominantly when in combination with *GSTT1 null*, may exacerbate CMM risk when it occurs in the context of specific phenotypic backgrounds known to increase susceptibility to the development of CMM.

A biological interaction between GST genotype and hair color is quite plausible. Persons with lighter hair color, *i.e.*, red and blond hair, tend to have fairer skin complexion and a corresponding greater proportion of pheomelanin in the epidermis (13). This is in contrast to those with brown or black hair who tend to have darker skin complexion and a greater amount of eumelanin. Pheomelanin is a poorer protector against cellular oxidative stress than is eumelanin because reactive oxygen species may be more likely to form when pheomelanin is exposed to UV radiation (14).

Our data support a moderate yet significant association with *GSTM1 null* and CMM among persons with light (*i.e.*, red, reddish-brown, and blond) hair color. It is possible that *GSTM1* is a more efficient reducer of oxidized DNA and lipids than *GSTT1* in skin. It is clear, however, that the combination of *GSTM1 null* and *GSTT1 null* confers the greatest risk of CMM in light-haired individuals. The association between *GST null* and CMM among persons with light hair color remained statistically significant even after further adjustment for either total number of DN alone or in combination with all of the other pigmentation characteristic variables included in Table 4 (data not shown). Because *GSTM1* and *GSTT1* have complementary yet somewhat redundant biochemical properties, the absence of an active form of both isozymes may represent a more limited ability to reduce cellular oxidative stress. Kerb *et al.* (15) noted that the minimal erythematous dose of UVB irradiation was lowest among three healthy subjects who were both *GSTM1 null* and *GSTT1 null*.

We acknowledge that our ability to draw strong conclusions based on these data is limited because of the small number of persons who were *GST null* and the potential for spurious associations resulting from the number of tests of significance performed.

We speculate that the association between *GST null* and CMM would be even greater among a subgroup consisting of only redheads. We were unable to directly test this in our data. The number of redheads in our study population totaled 51, of whom only 5 were controls and only 3 had DN. None of these redheaded controls were genotyped as *GST null*. In fact, only one subject without CMM (who was blond) was *GST null*, which contributed to the imprecision of the parameter estimate.

Of interest was the lack of a clear association between GST genotypes and measures of cutaneous risk markers for CMM, specifically DN. Because some, but not all, CMM arise

from DN precursors, it may be that *GSTM1 null* and *GSTT1 null* play a more crucial role in later stage melanocytic carcinogenesis by contributing to the continued deregulation of cellular functions within an already functionally and morphologically abnormal DN.

Seemingly inconsistent was the moderate association between *GST null* and floridly expressed (10 or more) DN. Because these persons are at greatest risk for development of CMM (10), it is possible that this association is indicative of an undiagnosed CMM rather than increased number of DN *per se*. To determine whether the lack of association between *GST null* and DN count was, in part, determined by the presence of CMM, we repeated analyses after stratification on case-control status. The Cochran-Mantel-Haenszel statistic (17.5;  $P = 0.002$ ) indicated that the association of *GST null* and DN was the same in cases and controls. However, we did note an increased likelihood of *GST null* (OR, 2.0; 95% CI, 0.92–4.5) among CMM cases with a solitary DN, compared with a decreased likelihood of *GST null* (OR, 0.29; 95% CI, 0.04–2.3) among controls with a solitary DN.

We have extensively evaluated our control group to better understand the potential impact of our sampling design on our reported associations. The majority of controls were partners of clinic patients (87%), 10% were friends, and 3% were in-laws or adoptive relatives. Statistical tests did not reveal any difference between partner controls and friend controls for either pigmentation characteristic or GST genotyping results (data not shown). The prevalence of *GSTM1 null* in CMM-free controls (46%) was well within the range published for this null polymorphism, whereas the prevalence of *GSTT1 null* among this group (24%) was slightly higher than that previously reported in Caucasians (reviewed in Ref. 16). Comparing DN controls with referred healthy controls without DN or CMM, there were no statistically significant differences in the proportions with *GSTM1 null* ( $\chi^2 = 0.84$ ;  $P = 0.36$ ), *GSTT1 null* ( $\chi^2 = 1.9$ ;  $P = 0.17$ ), or *GST null* ( $\chi^2 = 0.01$ ;  $P = 0.92$ ).

It is possible that a shared etiology exists between DN and CMM. Under this assumption, bias introduced by including DN into the referent group would likely make it more difficult to detect significant associations. Reassuringly, the associations we found between CMM and pigmentation characteristics were similar to those published previously (reviewed in Ref. 17).

Additional evaluation will be required to further explore the apparent interaction between GST genotypes and hair color phenotype (18) and to relate these results to additional factors determining melanogenesis and melanocytic carcinogenesis, *e.g.*, melanocortin-1 receptor allelic variation (18–20). Phenotypic pigmentation alone is not sufficient to predict CMM risk. The additional information provided by genotyping may contribute significantly to better models of prediction, prevention, and treatment of CMM.

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### References

1. Griffiths, H. R., Mistry, P., Herbert, K. E., and Lunec, J. Molecular and cellular effect of ultraviolet light-induced genotoxicity. *Crit. Rev. Clin. Lab. Sci.*, 35: 189–237, 1998.

2. Ketterer, B., and Meyer, D. J. Glutathione transferases: a possible role in the detoxication and repair of DNA and lipid hydroperoxides. *Mutat. Res.*, 214: 33–40, 1989.
3. Wang, W., and Ballatori, N. Endogenous glutathione conjugates: occurrence and biological functions. *Pharmacol. Rev.*, 50: 335–355, 1998.
4. Heagerty, A. H. M., Fitzgerald, D., Smith, A., Bowers, B., Jones, P., Fryer, A. A., Zhao, L., Alldersea, J., and Strange, R. C. Glutathione *S*-transferase *GSTM1* phenotypes and protection against cutaneous tumors. *Lancet*, 343: 266–268, 1994.
5. Heagerty, A., Smith, A., English, J., Lear, J., Perkins, W., Bowers, B., Jones, P., Gilford, J., Alldersea, J., Fryer, A., and Strange, R. C. Susceptibility to multiple cutaneous basal cell carcinomas: significant interaction between glutathione *S*-transferase *GSTM1* genotypes, skin type, and male gender. *Br. J. Cancer*, 73: 44–48, 1996.
6. Lear, J. T., Heagerty, A. H. M., Smith, A., Bowers, B., Payne, C. R., Smith, C. A. D., Jones, P. W., Gilford, J., Yengi, L., Alldersea, J., Fryer, A. A., and Strange, R. C. Multiple cutaneous basal cell carcinomas: glutathione *S*-transferase (*GSTM1*, *GSTT1*) and cytochrome P450 (*CYP2D6*, *CYP1A1*) polymorphisms influence tumour numbers and accrual. *Carcinogenesis (Lond.)*, 17: 1891–1896, 1996.
7. Lafuente, A., Molina, R., Palou, J., Castel, T., Moral, A., and Trias, M. Phenotype of glutathione *S*-transferase  $\mu$  (*GSTM1*) and susceptibility to malignant melanoma. *Br. J. Cancer*, 72: 324–326, 1995.
8. Shanley, S. M., Chenevix-Trench, G., Palmer, J., and Hayward, N. Glutathione *S*-transferase *GSTM1* null genotype is not over-represented in Australian patients with nevoid basal cell carcinoma syndrome or sporadic melanoma. *Carcinogenesis (Lond.)*, 16: 2003–2004, 1995.
9. Lee, J. E., Cheng, L., Mansfield, P. F., Ross, M. I., Gershenwald, J. E., Char, D., Chen, M., Lu, M., and Wei, Q. *GSTT1* and *GSTM1* polymorphisms and risk of melanoma. *Proc. Am. Soc. Clin. Oncol. Annu. Meet.*, 18: 540a, 1999.
10. Tucker, M. A., Halpern, A., Holly, E. A., Hartge, P., Elder, D. E., Sagebiel, R. W., Guerry, D., IV, and Clark, W. H., Jr. Clinically recognized dysplastic nevi. A central risk factor for cutaneous melanoma. *JAMA*, 277: 1439–1444, 1997.
11. Walker, A. H., Najarian, D., White, D. L., Jaffe, J. M., Kanetsky, P. A., and Rebbeck, T. R. Collection of genomic DNA by buccal swabs for polymerase chain reaction-based biomarker assays. *Environ. Health Perspect.*, 107: 3–7, 1999.
12. Rebbeck, T. R., Walker, A. H., Jaffe, J. M., White, D. L., Wein, A. J., and Malkowicz, S. B. Glutathione *S*-transferase- $\mu$  (*GSTM1*), and - $\theta$  (*GSTT1*) genotypes in the etiology of prostate cancer. *Cancer Epidemiol. Biomark. Prev.*, 8: 283–287, 1999.
13. Hunt, G., Kyne, S., Ito, S., Wakamatsu, K., Todd, C., and Thody, A. Eumelanin and pheomelanin contents of human epidermis and cultured melanocytes. *Pigm. Cell Res.*, 8: 202–208, 1995.
14. Menon, I. A., Persad, S., Ranadive, N. S., and Haberman, H. F. Photobiological effects of eumelanin and pheomelanin. In: J. Bagnara, S. N. Klaus, E. Paul, and M. Scharl (eds.), *Biological, Molecular, and Clinical Aspects of Pigmentation*, pp. 77–85. Tokyo: University of Tokyo Press, 1985.
15. Kerb, R., Brockmoller, J., Reum, T., and Roots, I. Deficiency of glutathione *S*-transferases T1 and M1 as heritable factors of increased cutaneous UV sensitivity. *J. Invest. Dermatol.*, 108: 229–232, 1997.
16. Rebbeck, T. R. Molecular epidemiology of the human glutathione *S*-transferase genotypes *GSTM1* and *GSTT1* in cancer susceptibility. *Cancer Epidemiol. Biomark. Prev.*, 6: 733–743, 1997.
17. Bliss, J. M., Ford, D., Swerdlow, A. J., Armstrong, B. K., Cristofolini, M., Elwood, J. M., Green, A., Holly, E. A., Mack, T., MacKie, R. M., Østerlind, A., Walter, S. D., Peto, J., and Easton, D. F. Risk of cutaneous melanoma associated with pigmentation characteristics and freckling: systematic overview of 10 case-control studies. *Int. J. Cancer*, 62: 367–376, 1995.
18. Strange, R. C., Ellison, T., Ichii-Jones, F., Bath, J., Hoban, P., Lear, J. T., Smith, A. G., Hutchinson, P. E., Osborne, J., Bowers, B., Jones, P. W., and Fryer, A. A. Cytochrome P450 *CYP2D6* genotypes: association with hair color, Breslow thickness, and melanocyte stimulating hormone receptor alleles in patients with malignant melanoma. *Pharmacogenetics*, 9: 269–276, 1999.
19. Valverde, P., Healy, E., Jackson, I., Rees, J. L., and Thody, A. J. Variants of the melanocyte stimulating hormone receptor gene are associated with red hair and fair skin in humans. *Nat. Genet.*, 11: 328–330, 1995.
20. Palmer, J. S., Duffy, D. L., Box, N. F., Aitken, J. F., O’Gorman, L. E., Green, A. C., Hayward, N. K., Martin, N. G., and Sturm, R. A. Melanocortin-1 receptor polymorphisms and risk of melanoma: is the association explained solely by pigmentation phenotype? *Am. J. Hum. Genet.*, 66: 176–186, 2000.

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